

WO 2005/017148

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Fig. 3

SDS-PAGE Analysis of
2H7 scFvIgG1 (SSS-S)H WCH2 WCH3 Protein.

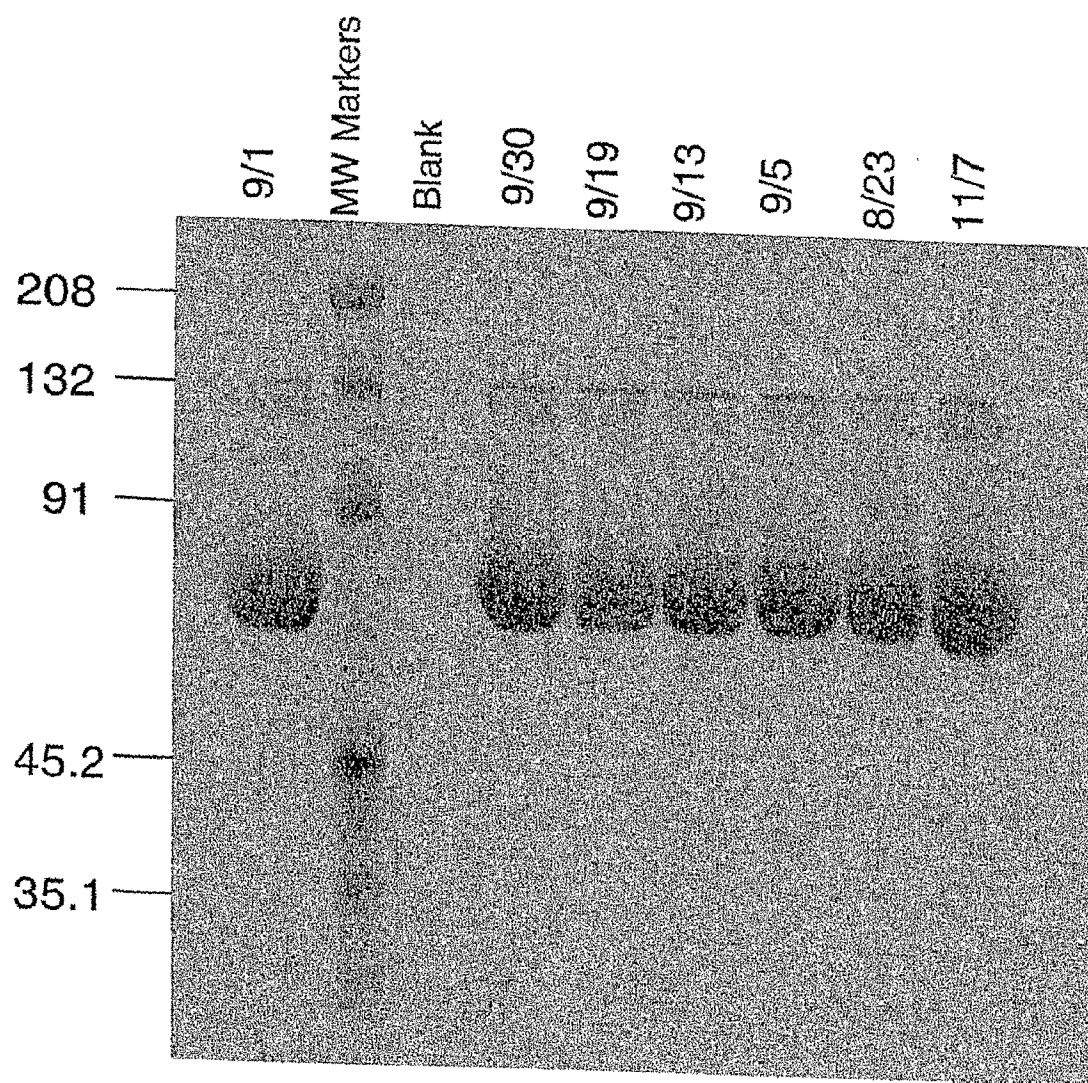


Fig. 4A

Complement Mediated B Cell Killing After Binding of
CD20-targeted 2H7 scFvIgG1 (SSS-S)H WCH2 WCH3:

2H7scFv-Ig Concentration	RAMOS		BJAB	
	# live cells/total cells		# live cells/total cells	
20 µg/ml + complement	-	0.16	-	0.07
5 µg/ml + complement	-	0.2	-	N.D.
1.25 µg/ml + complement	-	0.32	-	0.1
Complement alone	-	0.98	-	0.94

*Viability was determined by trypan blue exclusion and is tabulated as the fraction of viable cells out of the total number of cells counted.

**N.D. (not determined).

Fig. 4B

Antibody-dependent cellular cytotoxicity (ADCC) mediated by 2H7scFv-IgG1
(SSS-S)H WCH2 WCH3:

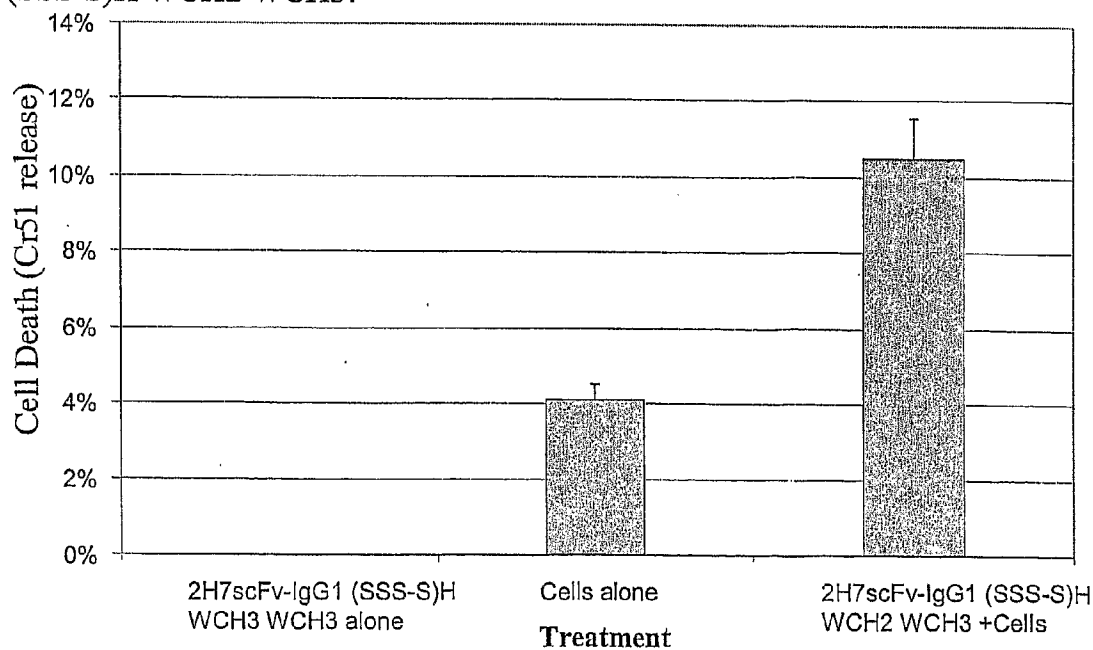
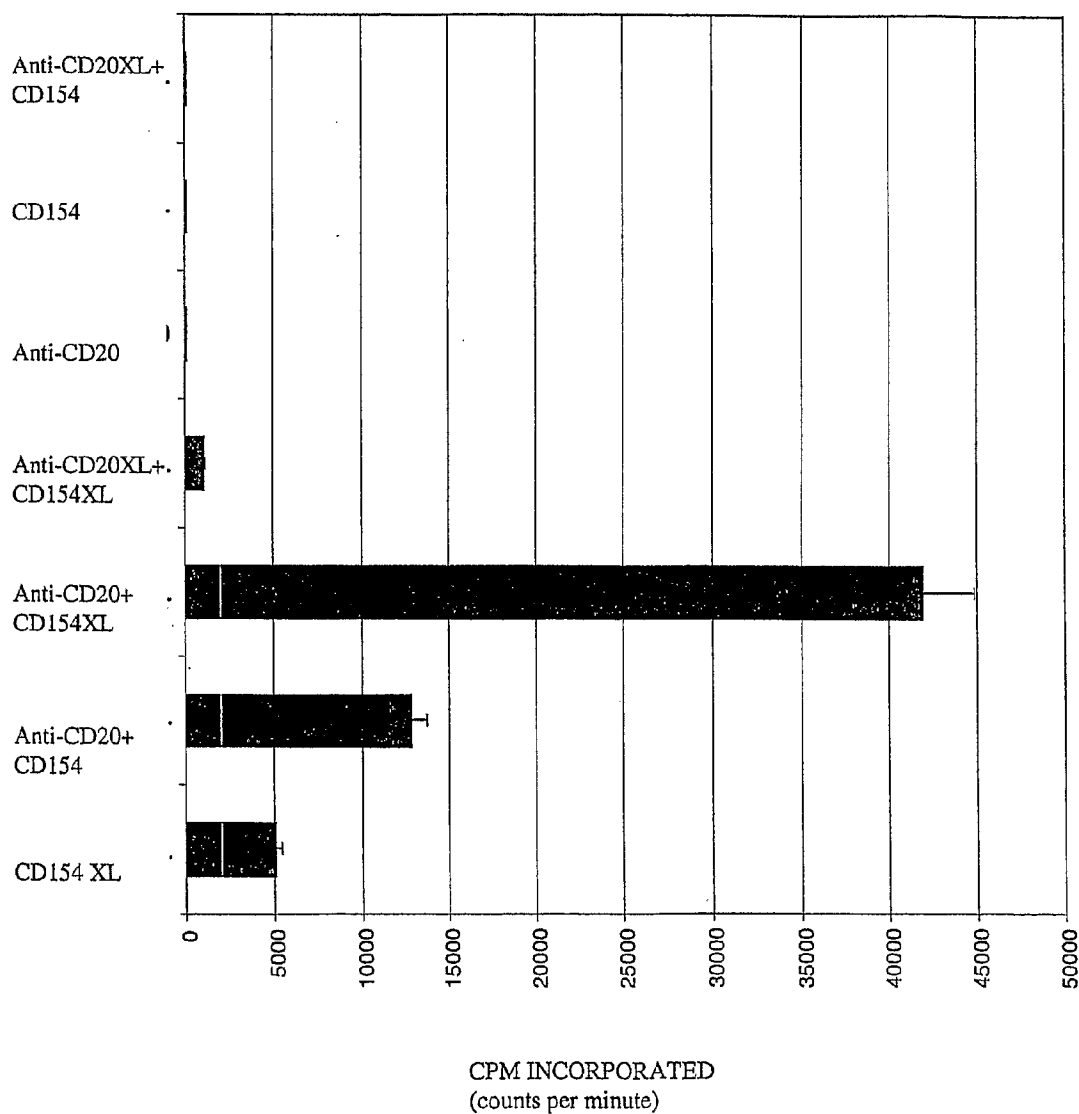


Fig. 5

Effects of Crosslinking of CD20 and CD40 Cell Surface Receptors
on B Cell Proliferation:



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Fig. 6

Effect of Simultaneous ligation of CD20 and CD40
on CD95 and apoptosis.

Fig. 6A.

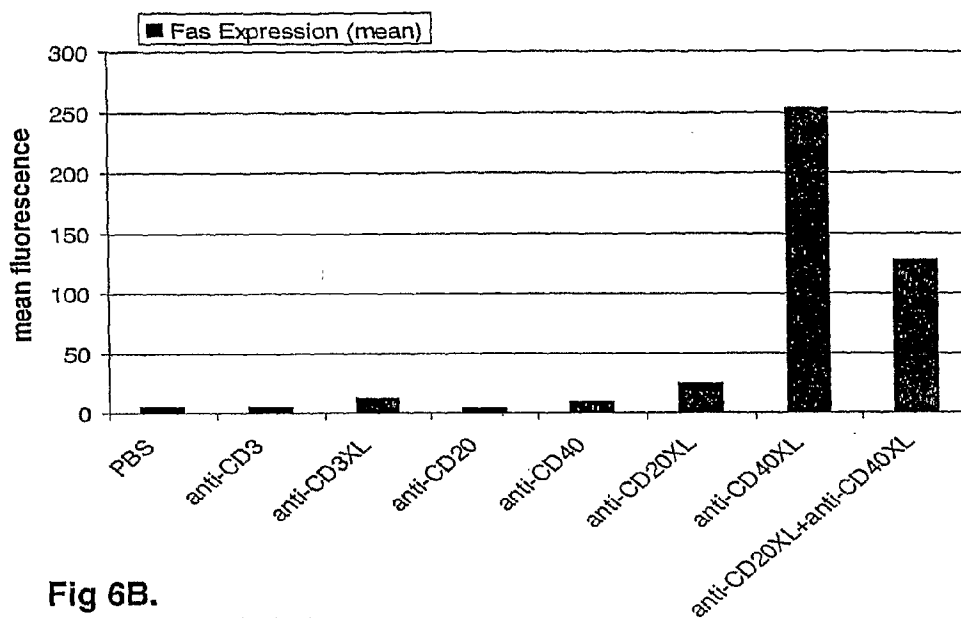
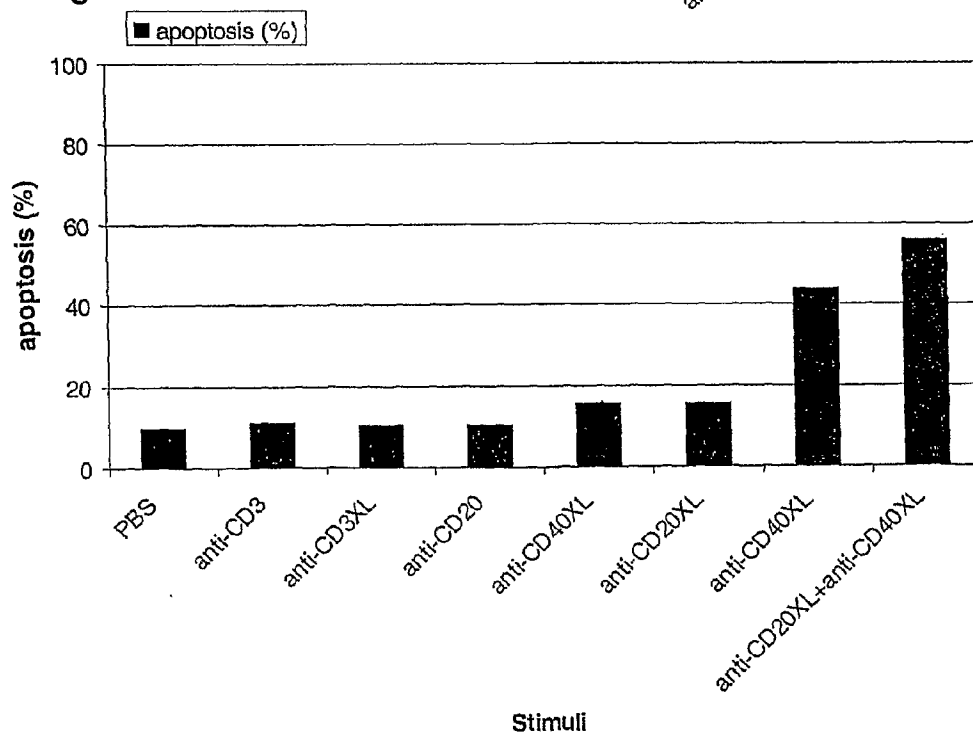


Fig 6B.



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2H7-CD154 L2 cDNA and predicted amino acid sequence:

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Fig. 7A (continued)

human CD154/amino acid 48→

```

                                Bcl/Bam hybrid site
781   G T G T T V T V S D P R R L D K I E D E
      GGCACAGGGA CCACGGTCAC CGTCTCTGAT CCAAGAAGGT TGGACAAGAT AGAAGATGAA

      R N L H E D F V F M K T I Q R C N T G E
841   AGGAATCTTC ATGAAGATTT TGTATTCATG AAAACGATAC AGAGATGCAA CACAGGAGAA

      R S L S L L N C E E I K S Q F E G F V K
901   AGATCCTTAT CCTTACTGAA CTGTGAGGAG ATTAAAAGCC AGTTTGAAGG CTTTGTGAAG

                                BclI
961   D I M L N K E E T K K E N S F E M Q K G
      GATATAATGT TAAACAAAGA GGAGACGAAG AAAGAAAACA GCTTTGAAAT GCAAAAAGGT

      BclI
      ~~~~~
1021  D Q N P Q I A A H V I S E A S S K T T S
      GATCAGAATC CTCAAATTGC GGCACATGTC ATAAGTGAGG CCAGCAGTAA AACAAATCTT

      V L Q W A E K G Y Y T M S N N L V T L E
1081  GTGTTACAGT GGGCTGAAAA AGGATACTAC ACCATGAGCA ACAACTTGGT AACCTTGGAA

      N G K Q L T V K R Q G L Y Y I Y A Q V T
1141  AATGGGAAAC AGCTGACCGT TAAAAGACAA GGACTCTATT ATATCTATGC CCAAGTCACC

                                HindIII
                                ~~~~~
1201  F C S N R E A S S Q A P F I A S L C L K
      TTCTGTTCCA ATCGGGAAGC TTCGAGTCAA GTCCTATTTA TAGCCAGCCT CTGCCTAAAG

      S P G R F E R I L L R A A N T H S S A K
1261  TCCCCGGTA GATTCGAGAG AATCTTACTC AGAGCTGCAA ATACCCACAG TTCCGCCAAA

      P C G Q Q S I H L G G V F E L Q P G A S
1321  CCTTGCGGGC AACAAATCCAT TCACTTGGGA GGAGTATTTG AATTGCAACC AGGTGCTTCG

                                NcoI
                                ~~~~~
1381  V F V N V T D P S Q V S H G T G F T S F
      GTGTTTGTCA ATGTGACTGA TCCAAGCCAA GTGAGCCATG GCACTGGCTT CACGTCCTTT

                                XhoI                                XbaI
                                ~~~~~                                ~~~~~
1441  G L L K L E * * S R
      GGCTTACTCA AACTCGAGTG ATAATCTAGA

```

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2H7scFv-CD154 S4 cDNA and predicted amino acid sequence:

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Fig. 7B

human CD154/amino acid 108 →

```

                                Bcl/Bam hybrid site
BclI
  G T G T T V T V S D P E N S F E M Q K G
781 GGCACAGGGA CCACGGTCAC CGTCTCTGAT CCAGAAAACA GCTTTGAAAT GCAAAAAGGT

BclI
~~~~~
  D Q N P Q I A A H V I S E A S S K T T S
841 GATCAGAATC CTCAAATTGC GGCACATGTC ATAAGTGAGG CCAGCAGTAA AACACATCT

  V L Q W A E K G Y Y T M S N N L V T L E
901 GTGTTACAGT GGGCTGAAAA AGGATACTAC ACCATGAGCA ACAACTTGGT AACCTGGAA

  N G K Q L T V K R Q G L Y Y I Y A Q V T
961 AATGGGAAAC AGCTGACCGT TAAAAGACAA GGACTCTATT ATATCTATGC CCAAGTCACC

                                HindIII
                                ~~~~~
  F C S N R E A S S Q A P F I A S L C L K
1021 TTCTGTTCCA ATCGGGAAGC TTCGAGTCAA GCTCCATTTA TAGCCAGCCT CTGCCTAAAG

  S P G R F E R I L L R A A N T H S S A K
1081 TCCCCCGGTA GATTCGAGAG AATCTTACTC AGAGCTGCAA ATACCCACAG TTCCGCCAAA

  P C G Q Q S I H L G G V F E L Q P G A S
1141 CCTTGCGGGC AACAATCCAT TCACTTGGGA GGAGTATTTG AATTGCAACC AGGTGCTTCG

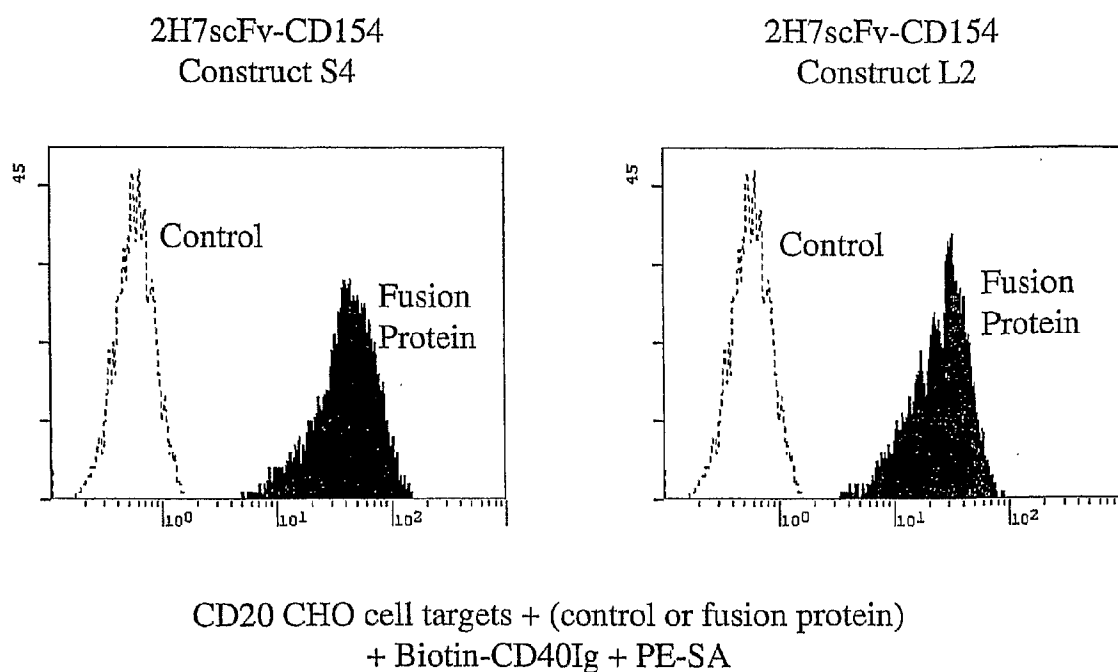
                                NcoI
                                ~~~~~
  V F V N V T D P S Q V S H G T G F T S F
1201 GTGTTTGTCA ATGTGACTGA TCCAAGCCAA GTGAGCCATG GCACTGGCTT CACGTCCTTT

                                XhoI                                XbaI
                                ~~~~~                                ~~~~~
  G L L K L E * * S R
1261 GGCTTACTCA AACTCGAGTG ATAATCTAGA

```


Fig. 8

Simultaneous Binding of 2H7scFv-CD154
Fusion Proteins to CD20 and CD40

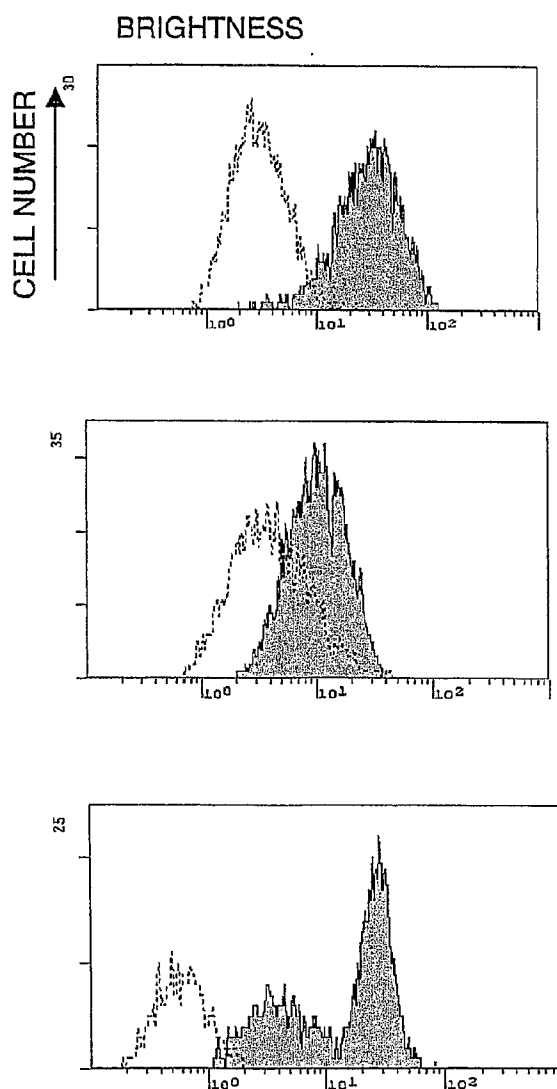


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Fig. 9

Induction of Apoptosis Measured by Binding of Annexin V after incubation with 2H7scFv-CD154



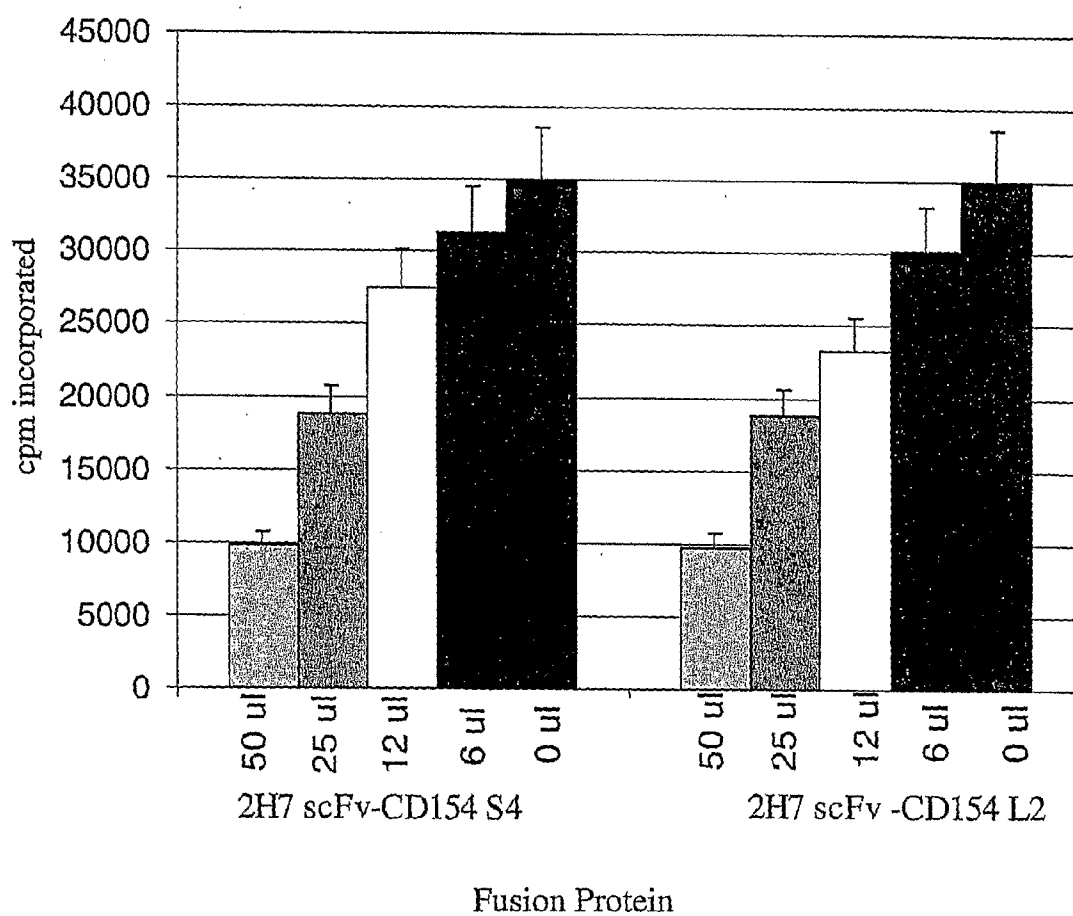
.....control supernatant ____2H7scFv-CD154 supernatant

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Fig. 10

Proliferation of T51 B Cell Line After Incubation with 2H7
scFv-CD154 S4 or 2H7 scFv-CD154 L2 Constructs



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Fig. 11

Schematic Representation of 2H7 scFvIg Constructs

2H7 scFvIgG (SSS-S)H WCH2 WCH3

OR 2H7 scFvIgG1 (SSS-S)H P238SCH2 WCH3 : 2H7 scFv

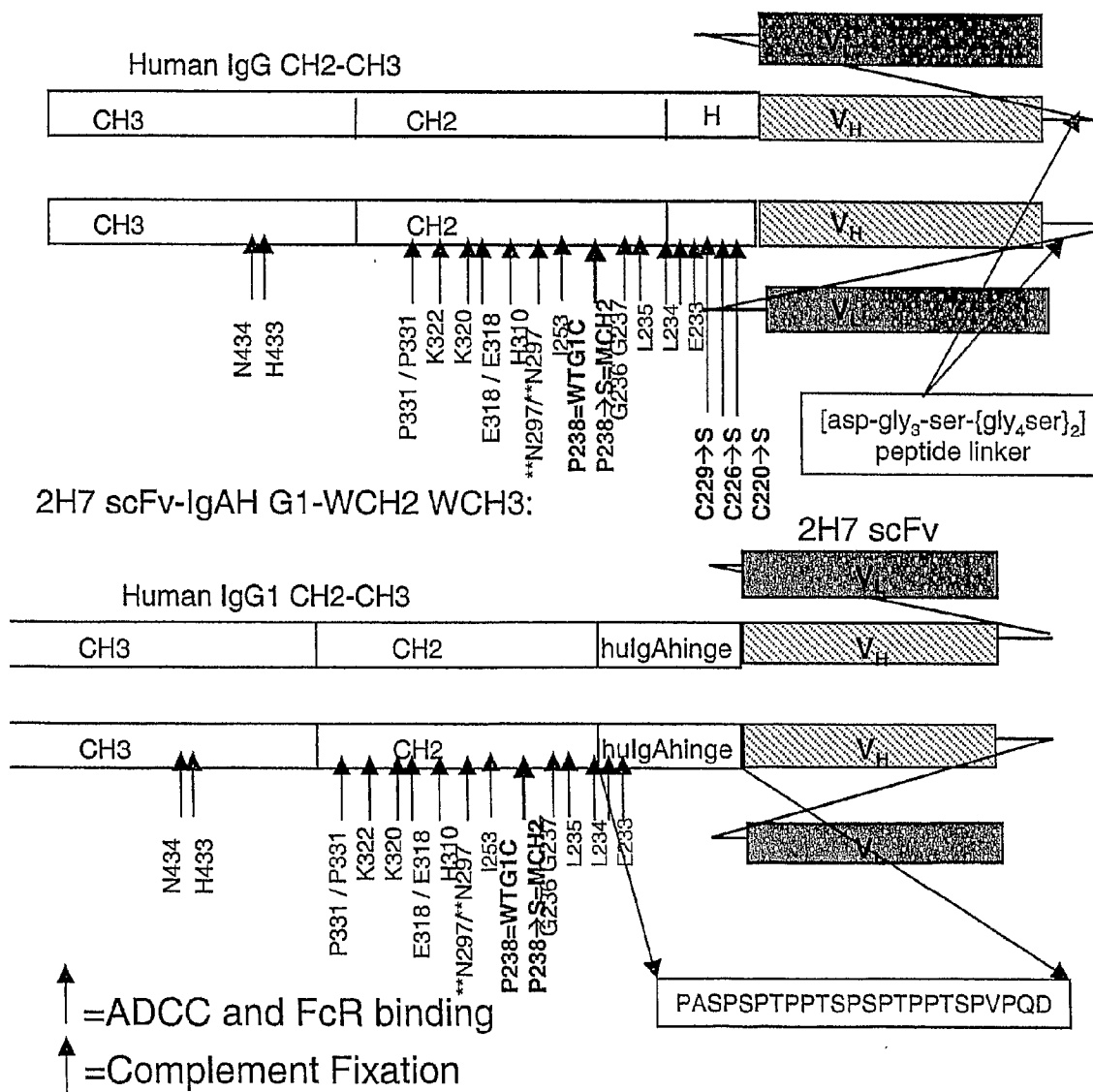


Fig. 12

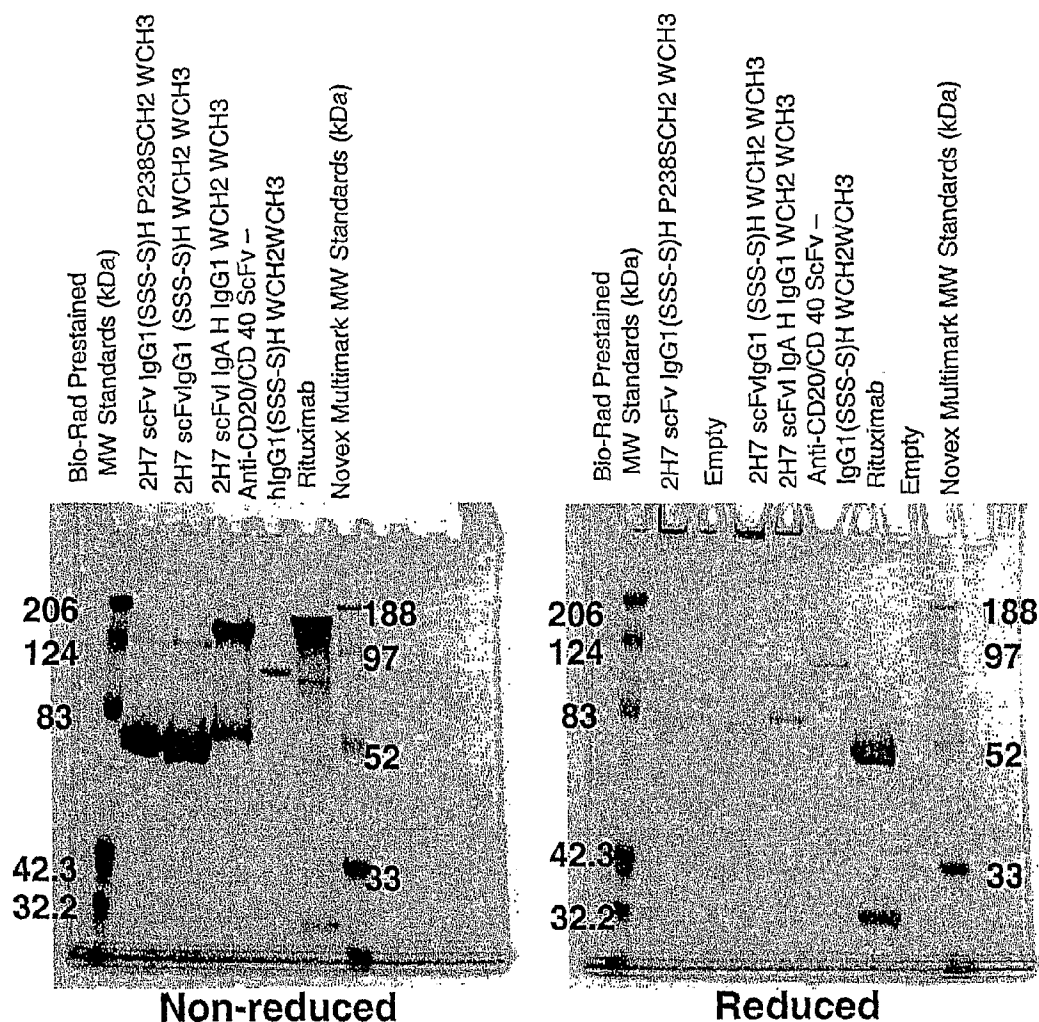


Figure 12: SDS-PAGE Analysis of CytosB Derivatives. Purified fusion protein derivatives of CytosB-scFvIg molecules and Rituximab were resuspended in SDS sample buffer, boiled, loaded onto 10% Novex Tris-Bis gels (Invitrogen, San Diego, CA) and subjected to nonreducing (left panel) or reducing (right panel) SDS-PAGE electrophoresis at 175 volts. Two different molecular weight markers, BioRad prestained markers, and Novex Multimark molecular weight markers were also loaded onto each gel and the approximate size in kDa of each marker band is indicated along each side of the photographed gels. Gels were stained in Coomassie Blue stain and photographed with a SONY Mavica Digital camera. The mutant hinge forms of 2H7 scFvIgG1 migrate at approximately 70 kDa under both nonreducing and reducing conditions, indicating that these molecules are monomeric rather than dimeric in structure. The IgA hinge form of 2H7scFvIg migrates at approximately 75 kDa under reducing conditions, but migrates predominately as a dimer of 140 kDa with a fraction of the protein migrating at 75 kDa under nonreducing conditions. Under nonreducing conditions, rituximab migrates as a diffuse band of between 150 and 200 kDa. The heavy and light chains resolve into separate bands of approximately 32 and 50 kDa when rituximab is reduced and subjected to SDS-PAGE.

Fig. 13

ADCC Activity of CytoxB (2H7 scFvIg) Constructs.

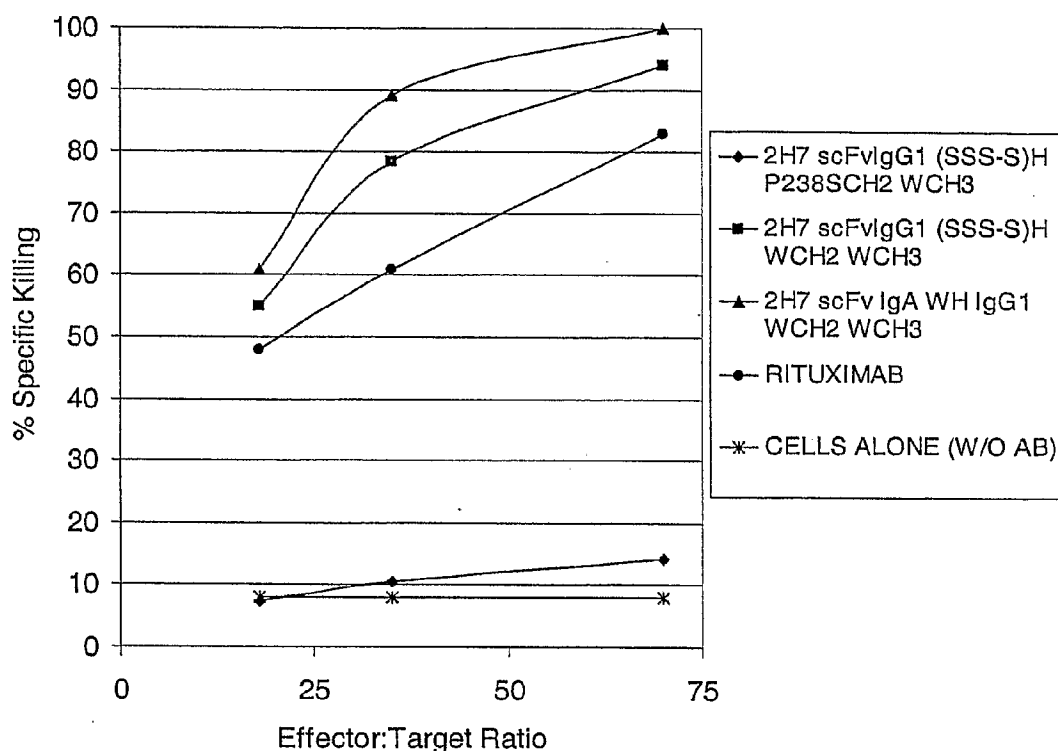


Figure 13: ADCC Activity of CytoxB Derivatives Compared to Rituximab. ADCC activity of CytoxB Derivatives or Rituximab was measured *in vitro* against BJAB B lymphoma cell line as target and using fresh human PBMC as effector cells. Effector to target ratios were varied as follows: 70:1, 35:1, and 18:1, with the number of BJAB cells per well remaining constant but varying the number of PBMC. Bjab cells were labeled for 2 hours with ^{51}Cr and aliquoted at a cell density of 5×10^4 cells/well to each well of flat-bottom 96 well plates. Purified fusion proteins or rituximab were added at a concentration of 10 mg/ml, and PBMC were added at 9×10^5 cells/well (18:1), 1.8×10^6 cells/well (35:1), or 3.6×10^6 cells/well (70:1), in a final volume of 200 μl . Spontaneous release was measured without addition of PBMC or fusion protein, and maximal release was measured by the addition of detergent (1% NP-40) to the appropriate wells. Reactions were incubated for 4 hours, and 100 μl culture supernatant harvested to a Lumaplate (Packard Instruments) and allowed to dry overnight prior to counting cpm released on a Packard Top Count NXT Microplate Scintillation Counter.

Fig. 14

CDC of Cytos B (2H7 scFvIg) Constructs

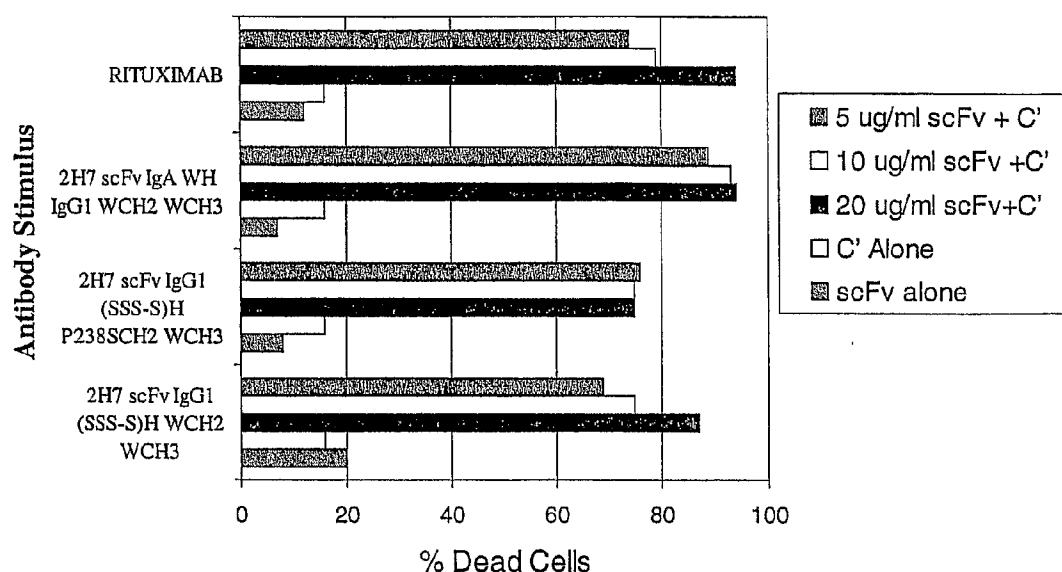
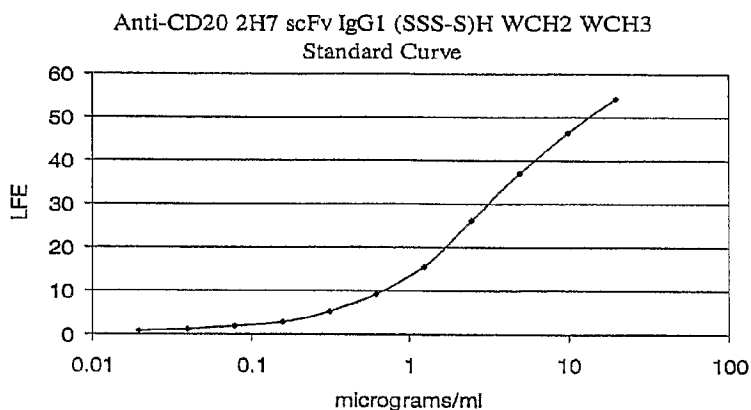


Figure 14: Complement Dependent Cytotoxicity (CDC) Activity of CytosB Derivatives Compared to Rituximab. 2H7 scFvIgG1 (SSS-S)H WCH2 WCH3, 2H7 scFvIgG1 (SSS-S)H WCH2 WCH3, and 2H7scFv IgA WH IgG1 WCH2 WCH3 derivatives and Rituximab were compared for their ability to mediate complement dependent cytotoxicity. Rabbit complement (Pel-Freez) was diluted 1:10 and added to BJAB cells along with dilutions of each antibody derivative (20 μ g/ml, 10 μ g/ml, and 5 μ g/ml). Controls were also included without addition of complement (C') or scFv derivative. Reactions were allowed to continue for 1 hour, and cells from each well were then stained with trypan blue and the cell viability counted using a hemacytometer. Data is graphed as % of dead cells/total cells counted for each condition assayed.

Fig. 15

2H7 (anti-CD20) scFv IgG1 (SSS-S)H WCH2 WCH3
In Vivo Half Life



Macaque A99314

	Day	Binding intensity (LFE) @ 1:50 dilution of serum	estimated concentration (µg/ml)
Injection #1	-7	0.213	<0.1
	0	0.227	<0.1
	1	7.79	25.1
	3	5.51	15.6
Injection #2	7	3.37	9.4
	8	11.33	41.7
	10	5.45	15.4
	14	0.27	<0.1

Macaque F98081

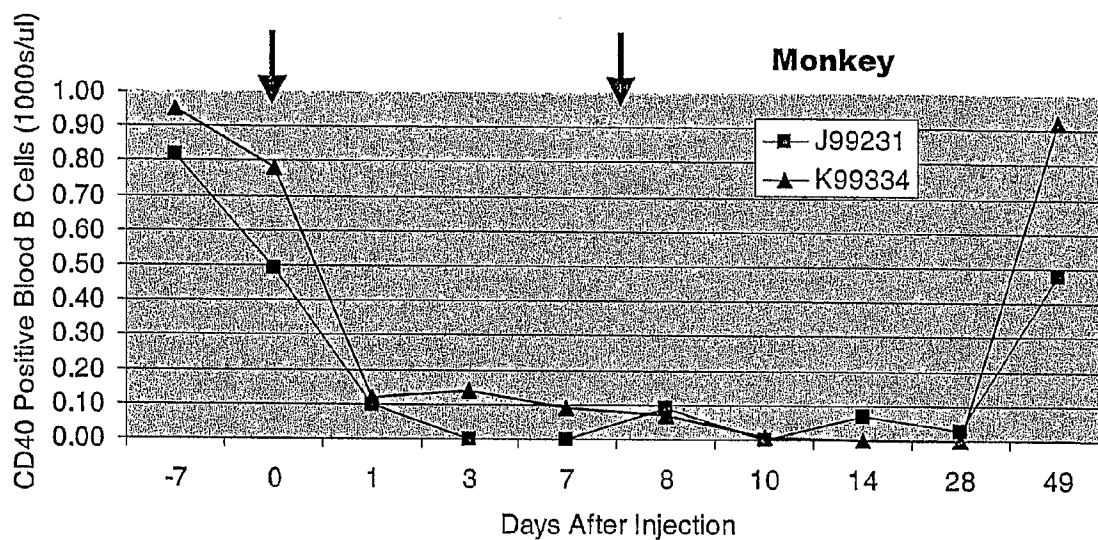
	Day	Binding intensity (LFE) @ 1:50 dilution of serum	estimated concentration (µg/ml)
Injection #1	-7	0.208	<0.1
	0	0.219	<0.1
	1	6.73	21.9
	3	6.14	19.3
Injection #2	7	3.04	8.7
	8	9.83	33.8
	10	4.77	14.4
	14	0.231	<0.1

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Fig. 16

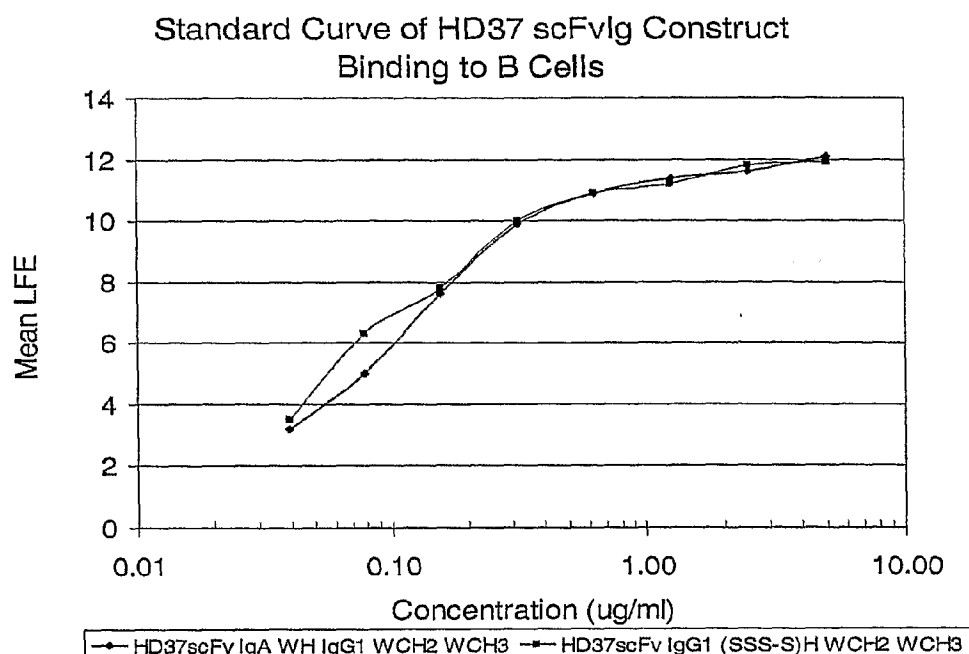
B Cell Depletion in macaques mediated by Cytos B20
(2H7 scFv IgG1 (SSS-S)H WCH2 WCH3) Construct



- CytosB20 injections of 6mg/kg yields 3 week B-cell depletion
- 3-4 day half-life *in vivo*
- CD20 saturation in lymph node B-cells at d14
- No first dose effects
- No anti-chimeric antibody development

Fig. 17

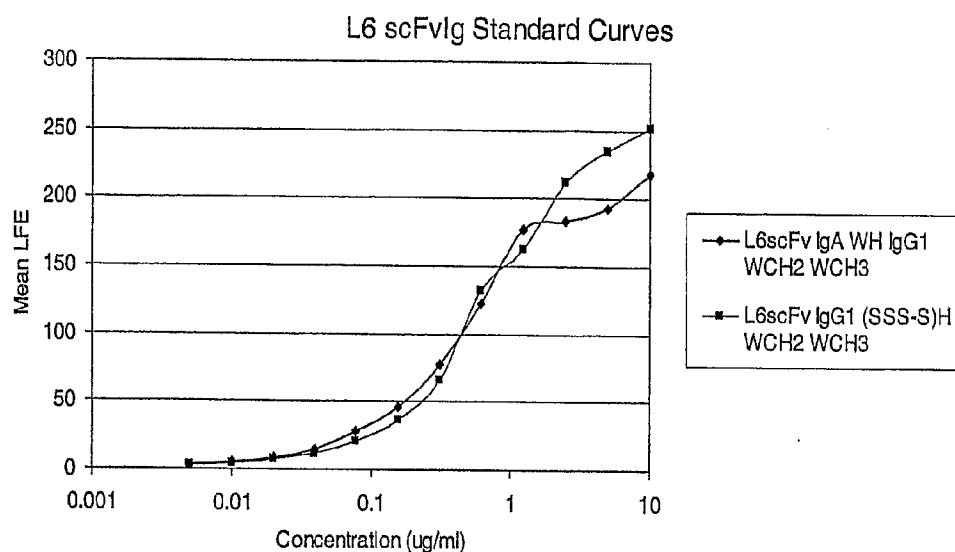
Production Levels of HD37 scFvIg Constructs by CHO Cell Lines



Clone/Isolate	Mean LFE at 1:100	Estimated Concentration
Bulk HD37 scFv		
IgA WH IgG1 WCH2 WCH3	11.2	> 60 ug/ml
1B2	10.4	>50 ug/ml
6C5	10.5	>50 ug/ml
4B1	8.6	>40 ug/ml
Bulk HD37 scFv		
IgG1 (SSS-S)H WCH2 WCH3	10.9	> 50 ug/ml
2G8	10.6	> 50 ug/ml
3F3	8.3	>40 ug/ml
3D9	11.1	> 60 ug/ml

Fig. 18

Production of L6 scFvIg constructs by CHO Cells



Construct	Mean LFE 1:20	Estimated Concentration
L6scFv IgA WH IgG1 WCH2 WCH3 unamplified CHO sup	51.1	6.25 ug/ml
L6scFv IgG1(SSS-S)H WCH2 WCH3 unamplified CHO sup	23.0	3.2ug/ml

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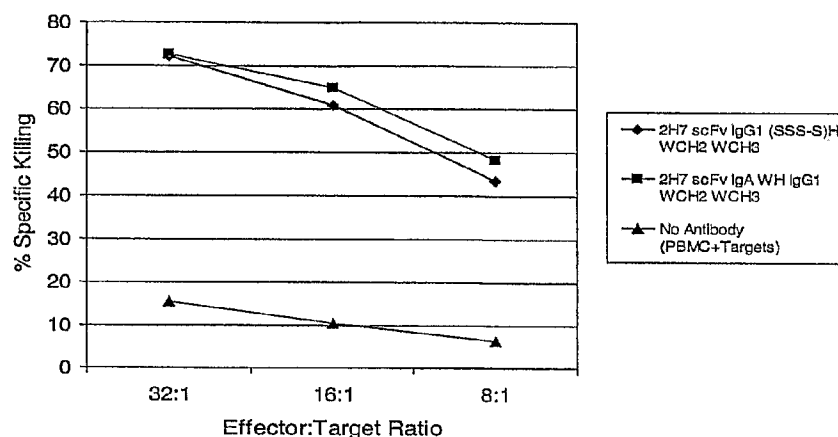
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Fig. 19

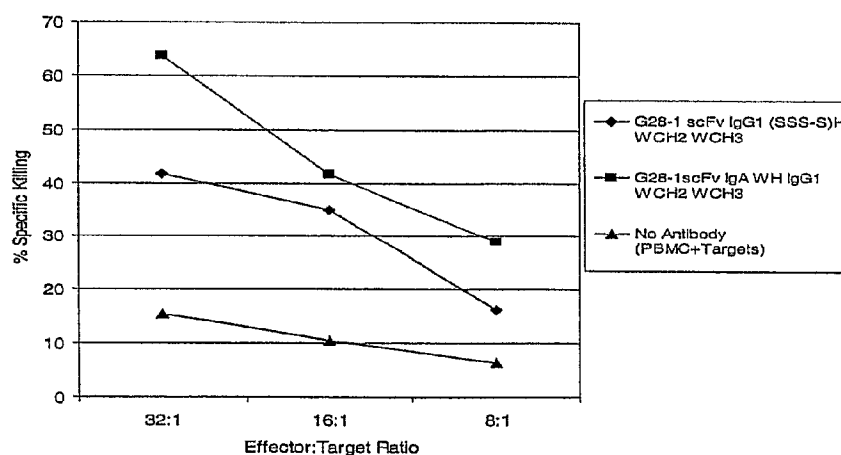
ADCC Activity of 2H7 scFvIg, G28-1 scFvIg, and HD37 scFvIg Constructs

ADCC Activity of scFvs Targeted to B Cell Antigens

A. 2H7 (anti-CD20) scFv constructs



B. G28-1 (anti-CD37) scFv constructs



C. HD37 (anti-CD19) scFv constructs

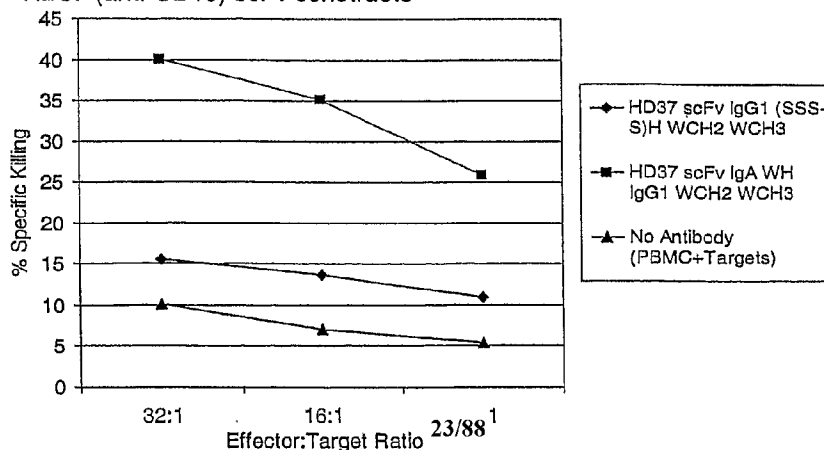
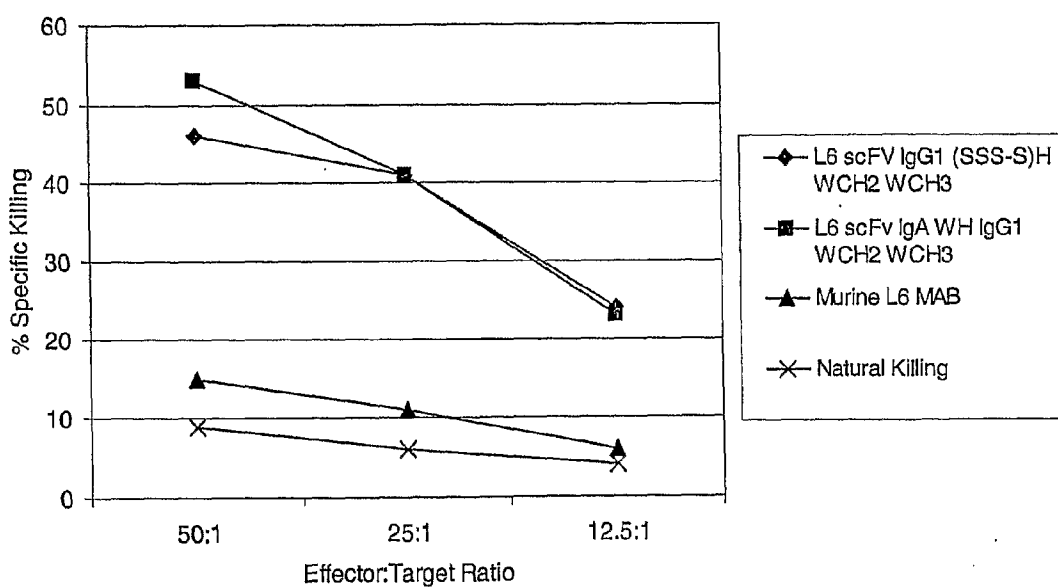


Fig. 20

ADCC Activity of L6 scFvIg Constructs

ADCC Activity of L6scFvIg Constructs with 2981 Targets

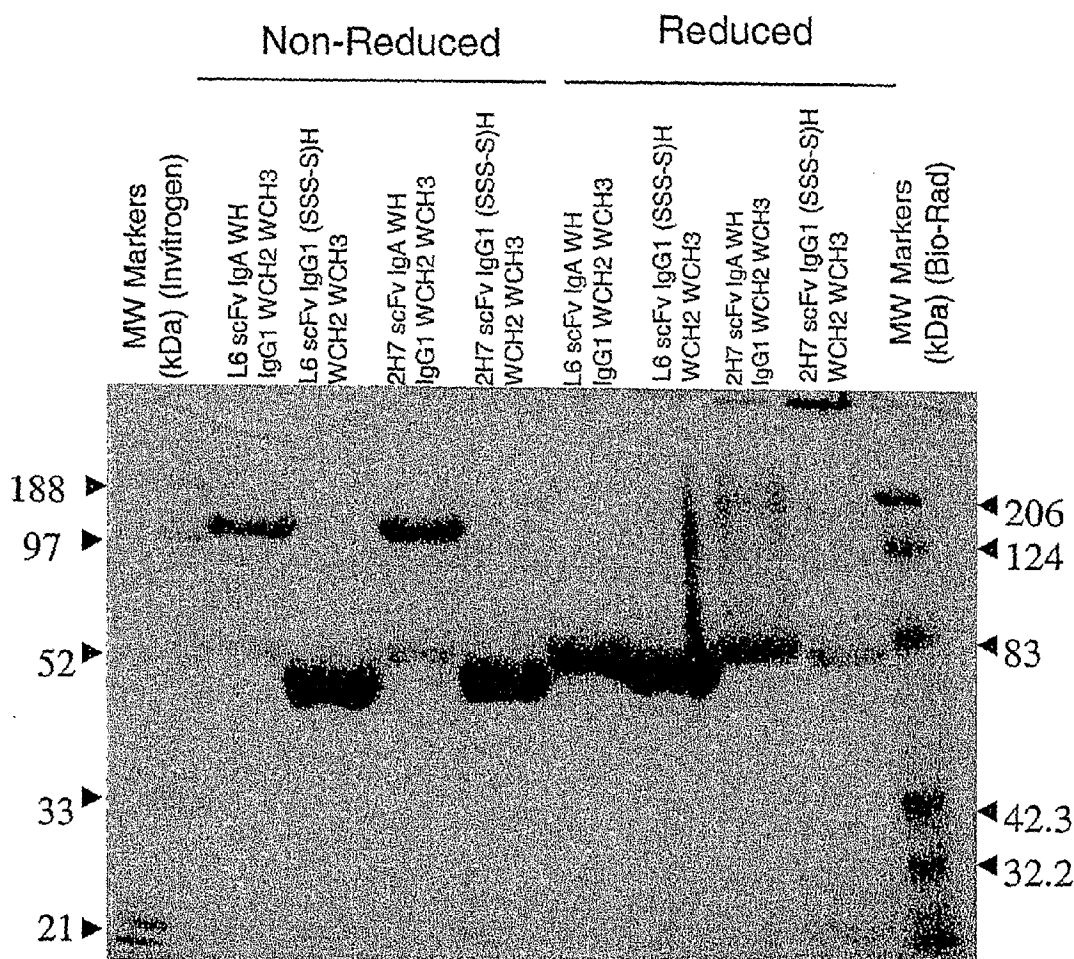


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Fig. 21

SDS-PAGE Analysis of L6 and 2H7
scFvIg Fusion Proteins.

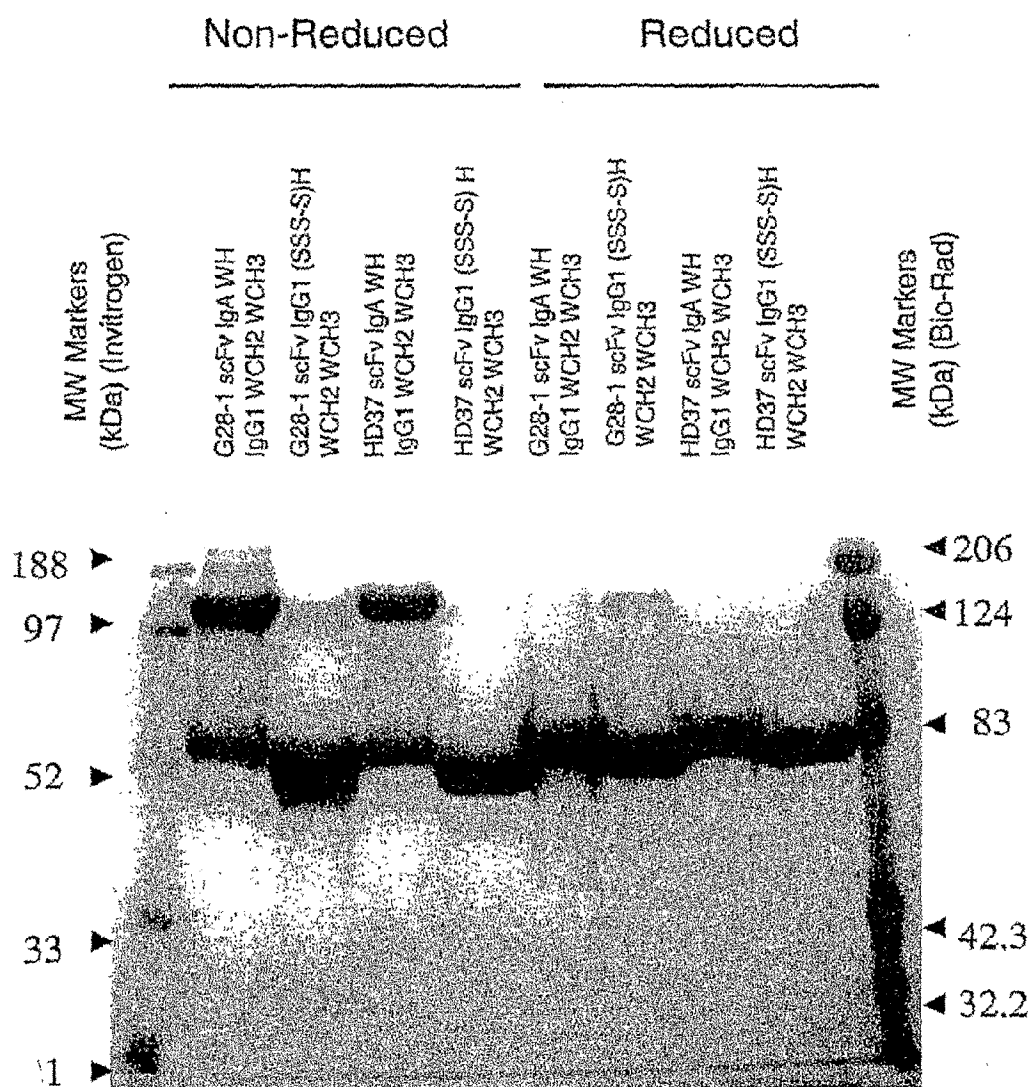


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Fig. 22

SDS-PAGE Analysis of G28-1 and HD37
scFvlg Constructs.



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Fig. 23

Sequence alignment of human and llama Fc regions.

	HINGE	CH2→
an IgG1:	DQEPKSCDKT-----HTCPPC	PAPELLGGPSVFLFPPKPKDTLMISRTPEVTCVVVDVSHEDPEVKFNWYVDG
ma IgG2:	DQEPKTPKPPQPPQPNPTTESKCPKC	PAPELLGGPSVFIFFPPKPKDVLSTISGRPEVTCVVVDVGQEDPEVSFNWYIDG
ma IgG1:	--EPHGG-----CTCPQC	PAPELPGGPSVFVFPPKPKDVLSTISGRPEVTCVVVDVGKEDPEVNFNWIYIDG
ma IgG3:	--AHHSEDPT-----SKCPKC	PGPELLGGPTVFIFPPKAKDVLSTTRKPEVTCVWVWVKKTLRSSSSWSVDD

VEVHNAKTKPREEQYNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKALPAPIEKTISKAKGQPREPQVYTLPPSRDELTKNQVSLT
TAEVIRANTRPKEEQFNSTYRVVSVLPPIQHQQDWTGKEFKCKVNNKALPAPIEKTISKAKGQTRFPQVYTLAPHREELAKDTVSVT
VEVIRANTKPKKEQFNSTYRVVSVLPPIQHQQDWTGKEFKCKVNNKALPAPIERTISKAKGQTRFPQVYTLAPHREELAKDTVSVT
TEVHTAETKPKKEQFNSTYRVVSVLPPIQHQQDWTGKEFKCKVNNKALPAPIERTISKAKGQTRFPQVYTLAPHREELAKDTVSVT

CLVKGFYPSDIAVEWESNGQPEN--NYKTTFPVLDSDGSFFLYSKLTVDKSRWQQGNVVFSCSVMEALHNHYTQKSLSLSPGK
CLVKGFYPPDINVEWQRNGQPESXGTYATTPQLDNDGTYFLXSKKSVGKNTWQQGETFTCVVMHEALHNHYTQKSITQSSGK
CLVKGFYPADINVEWQRNGQPESEGTYANTPPQLDNDGTYFLYRLSVGKNTWQRGETLTGVVMHEALHNHYTQKSITQSSGK
CLVKGFPPADINVEWQRNGQPESEGTYANTPPQLDNDGTYFLYSLSVGKNTWQQGEVFTCVVMHEALHNHSTQKSITQSSGK

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Fig. 24

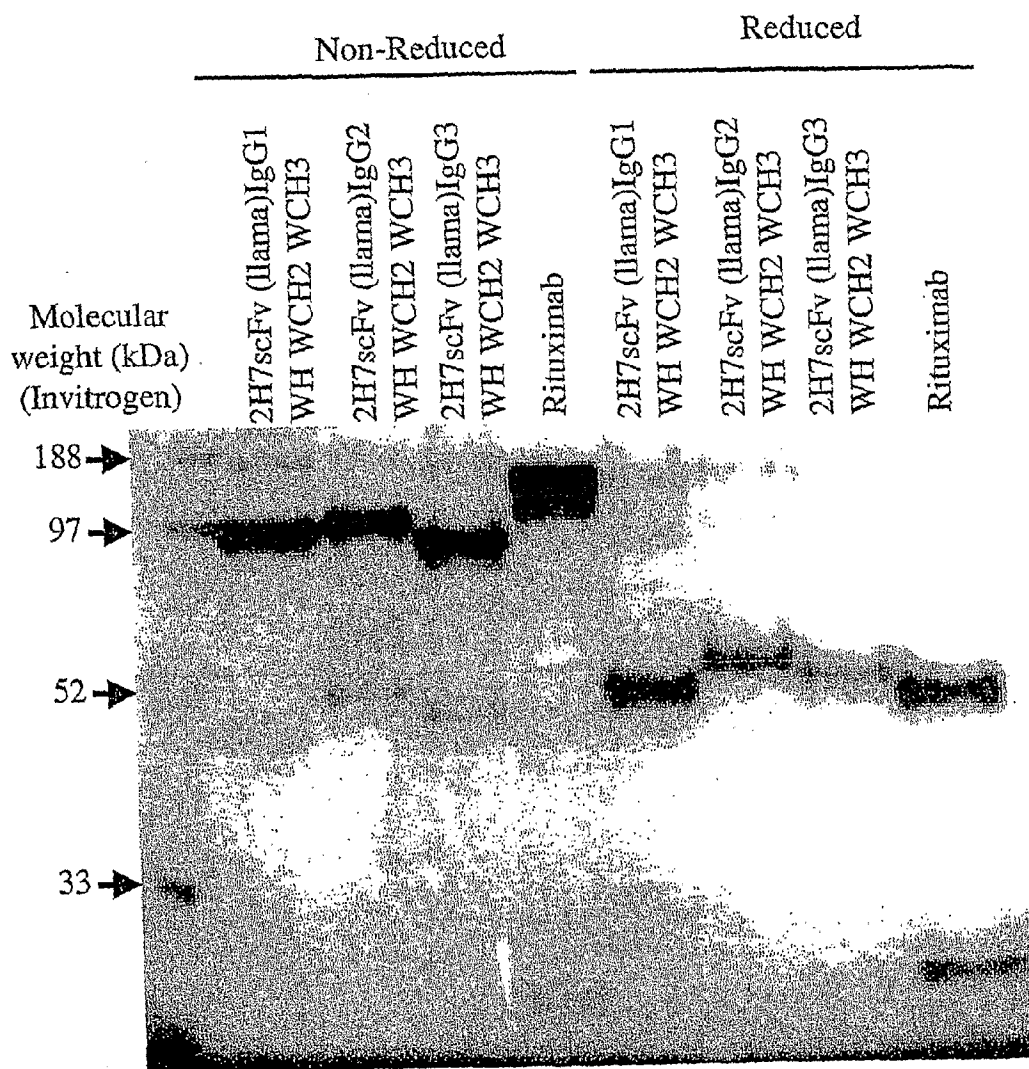


Fig. 25

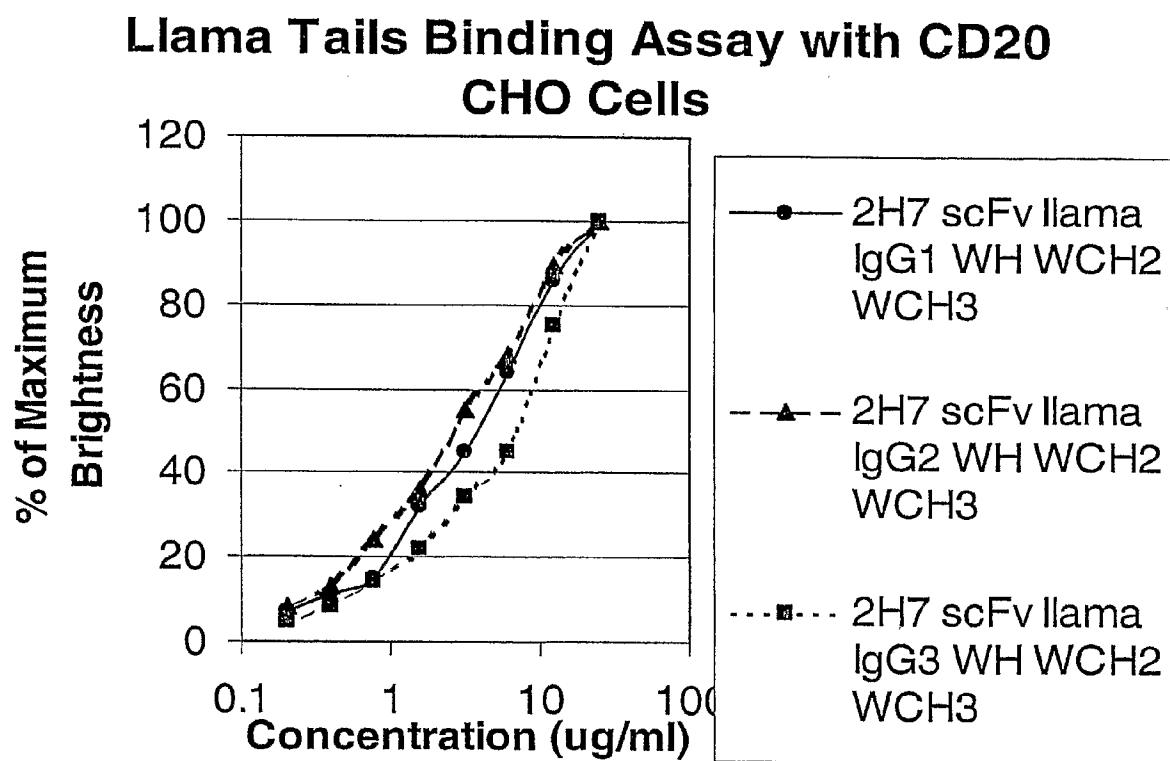


Fig. 26

2H7 scFvIg Llama Tails binding Assay with CD20 CHO Cells

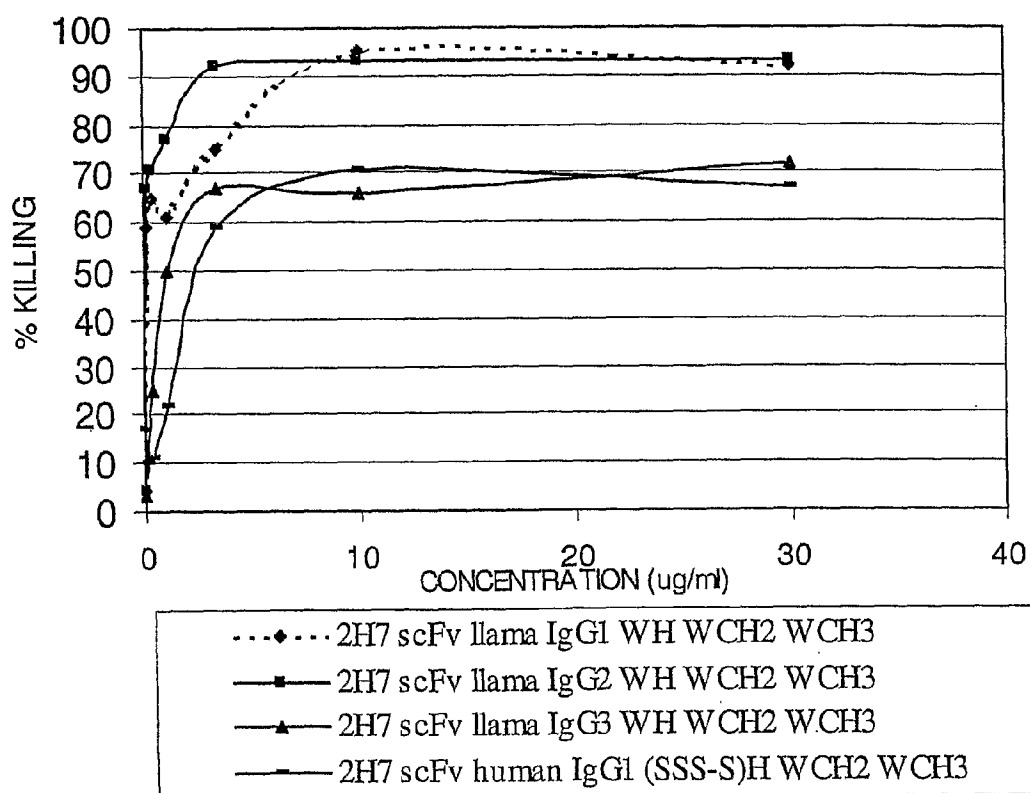


Fig. 27

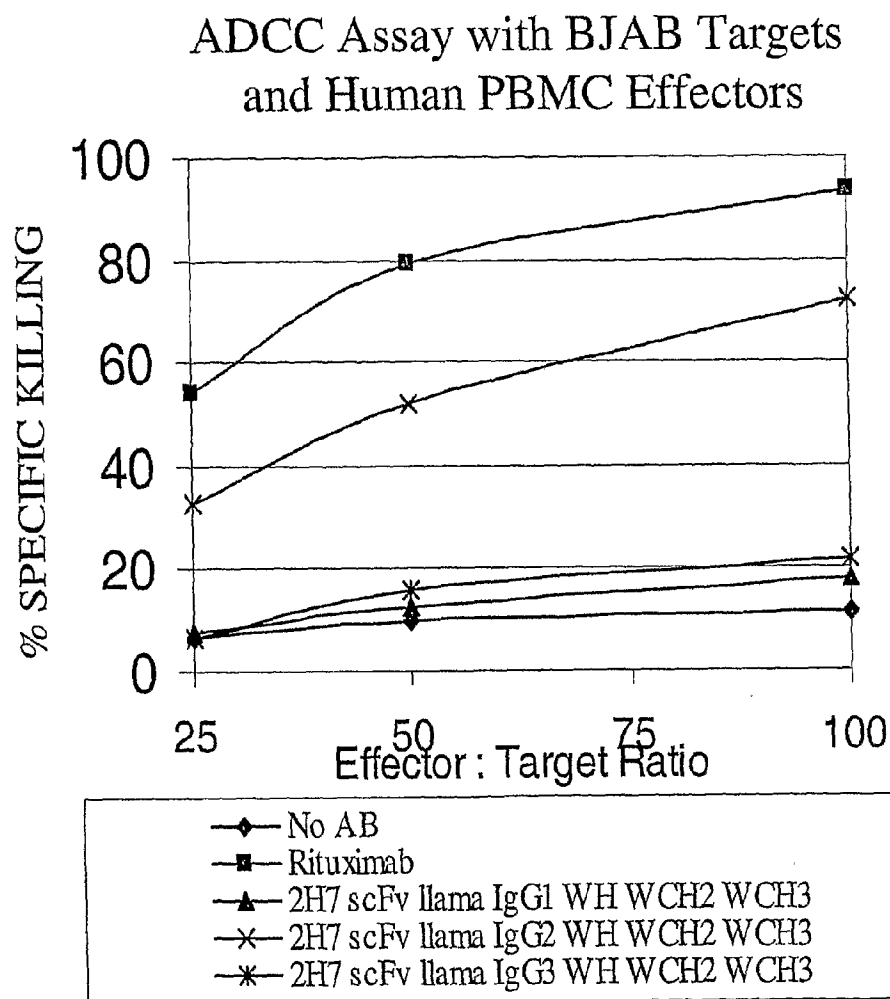


Fig. 28

ADCC Assay with BJAB Cells
And Llama PBMC Effectors

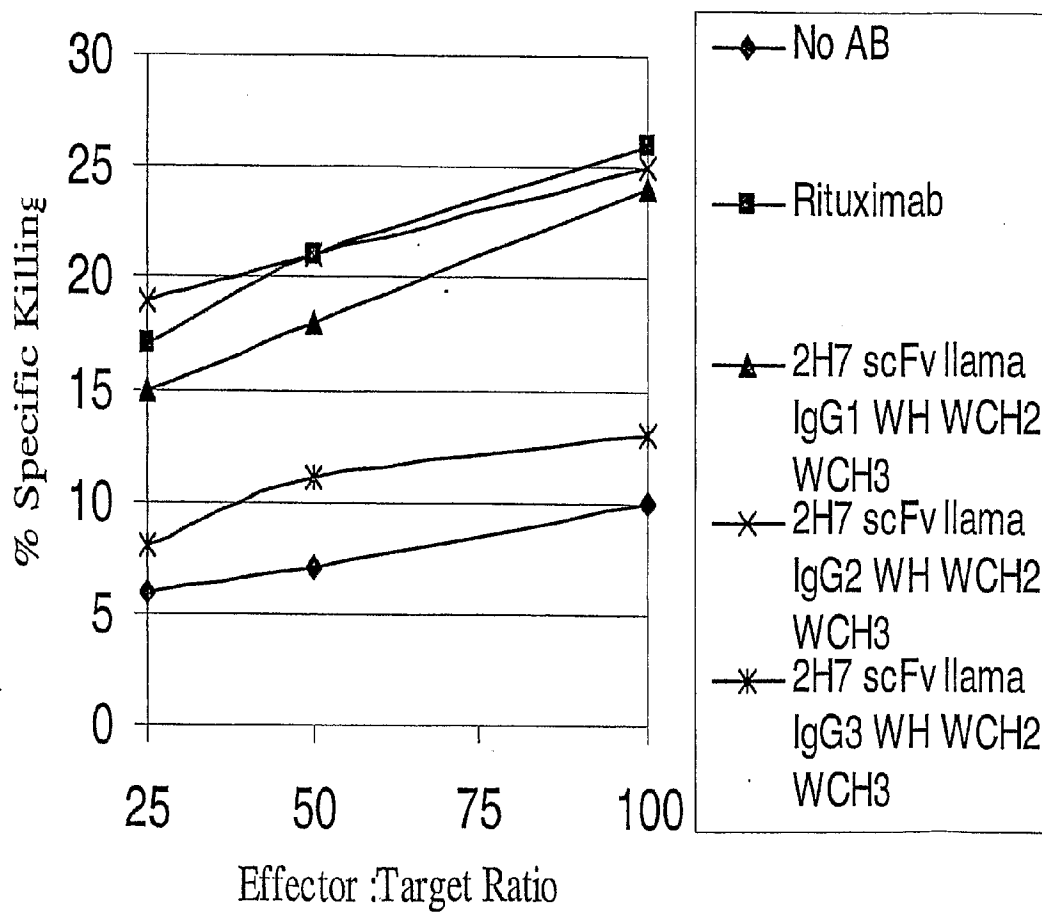


Fig. 29

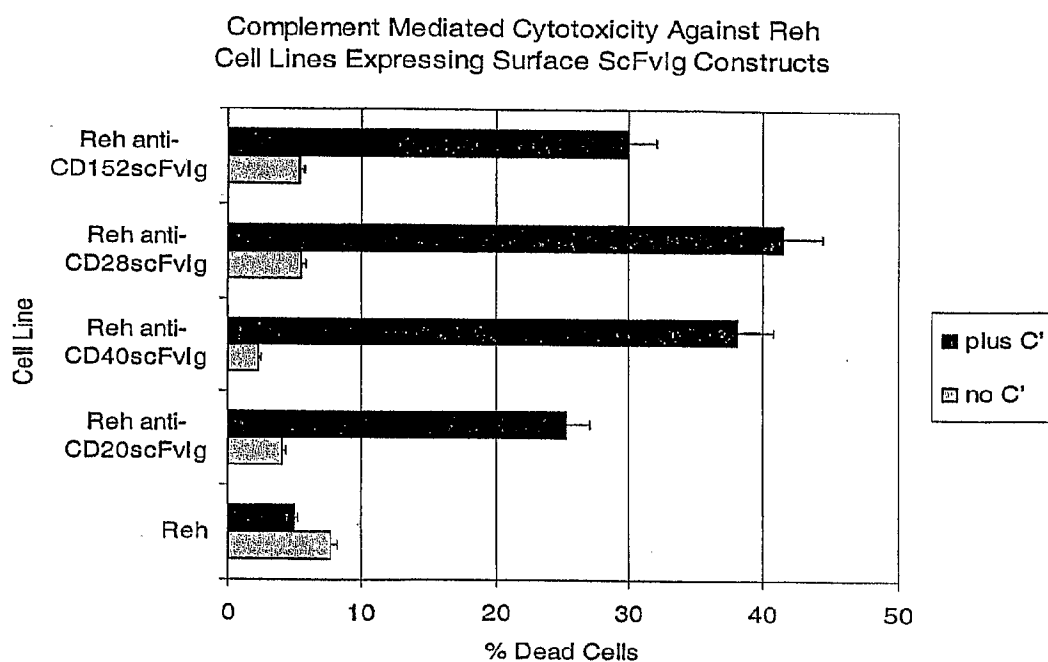


Fig. 30

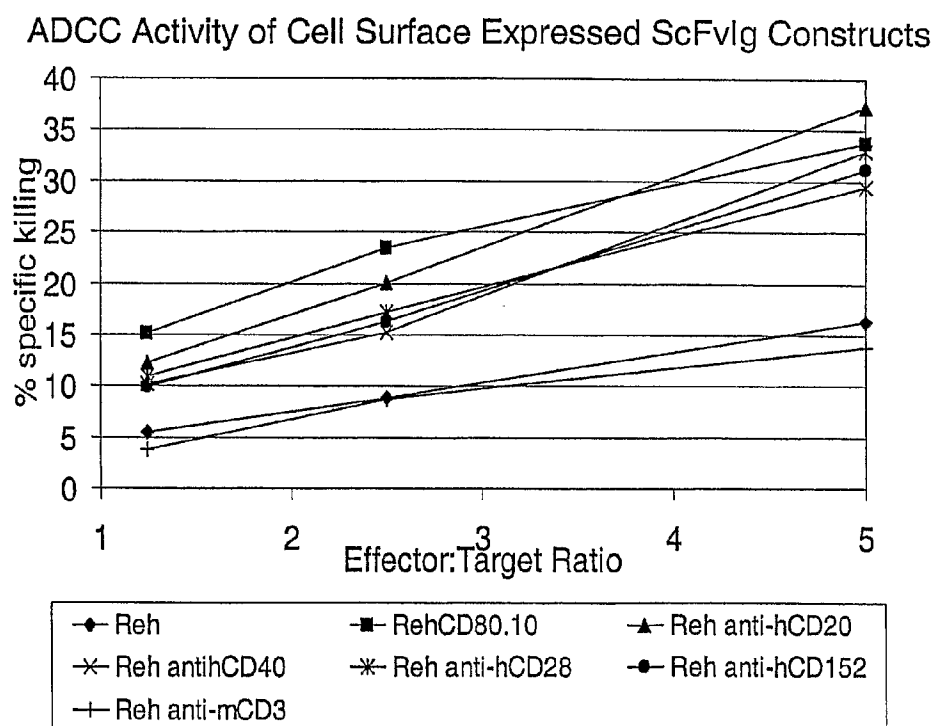


Fig. 31

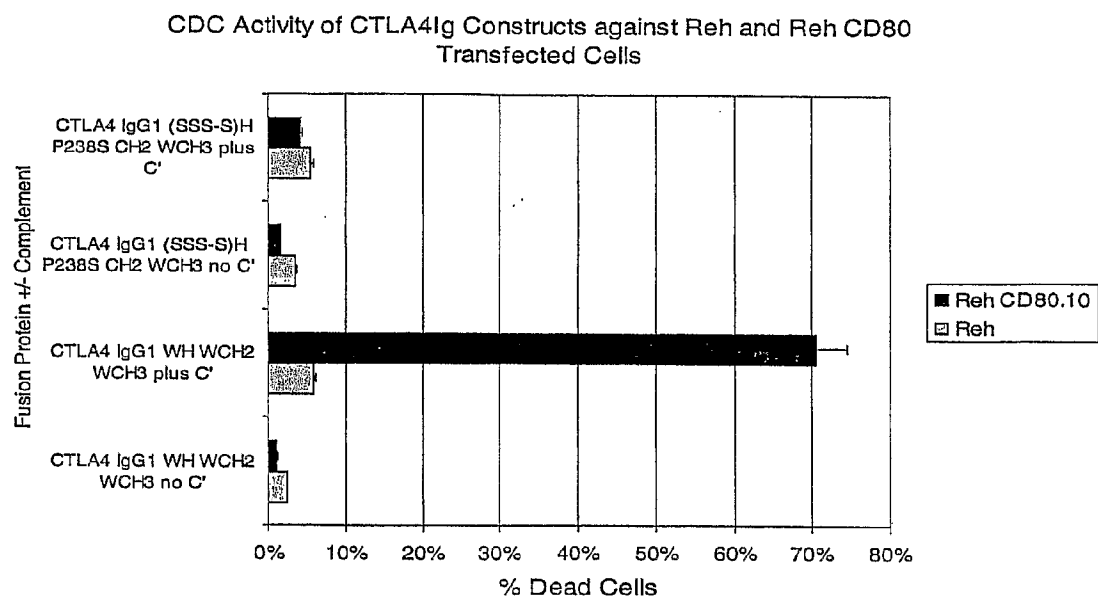
Ig Constructs and Nomenclature:

Name Identifier	Hinge Sequence	CH2 Sequence	CH3 Sequence
hIgG1 (CCC-P)H WCH2 WCH3	IgG1 WT Hinge (CCC-P)	Wild Type CH2	Wild Type CH3
hIgG1 (SSS-S)H WCH2 WCH3	IgG1 Mutant Hinge (SSS-S)	Wild type CH2 (IgG1)	Wild type CH3 (IgG1)
VH L11S hIgG1 (SSS-S)H WCH2 WCH3	IgG1 Mutant Hinge (SSS-S)	Wild type CH2 (IgG1)	Wild type CH3 (IgG1)
IgG1 (SSC-S)H WCH2 WCH3	IgG1 Mutant Hinge (SSC-S)	Wild type CH2 (IgG1)	Wild type CH3 (IgG1)
IgG1 (SCS-S)H WCH2 WCH3	IgG1 Mutant Hinge (SCS-S)	Wild type CH2 (IgG1)	Wild type CH3 (IgG1)
IgG1 (CSS-S)H WCH2 WCH3	IgG1 Mutant Hinge (CSS-S)	Wild type CH2 (IgG1)	Wild type CH3 (IgG1)
IgG1 (SSS-S)H P238S CH2 WCH3	IgG1 Mutant Hinge (SSS-S)	Mutant CH2 (IgG1) Pro→Ser 238	Wild type CH3 (IgG1)
IgA WH hIgG1 WCH2 WCH3	IgA Hinge	Wild type CH2 (IgG1)	Wild type CH3 (IgG1)
IgA WH IgA WCH2 WCH3	IgA Hinge	Wild type CH2 (IgA)	Wild type CH3 (IgA)
IgA WH IgA WCH2 T4CH3	IgA Hinge	Wild type CH2 (IgA)	Truncated CH3 (IgA) Missing 4 aa at COOH

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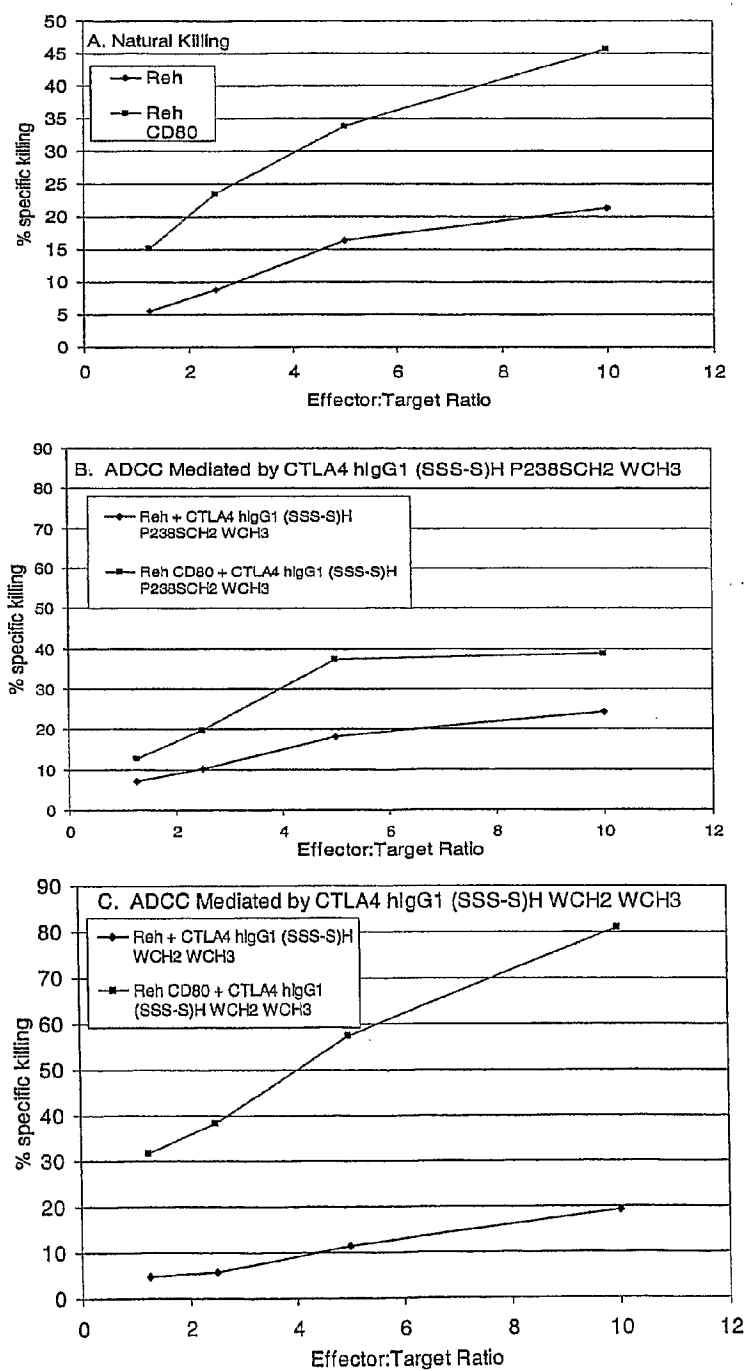
Fig. 32



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Fig. 33



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Fig. 34

Binding of 2H7 scFvIg Constructs
with Alternative Tails to CD20 CHO Cells

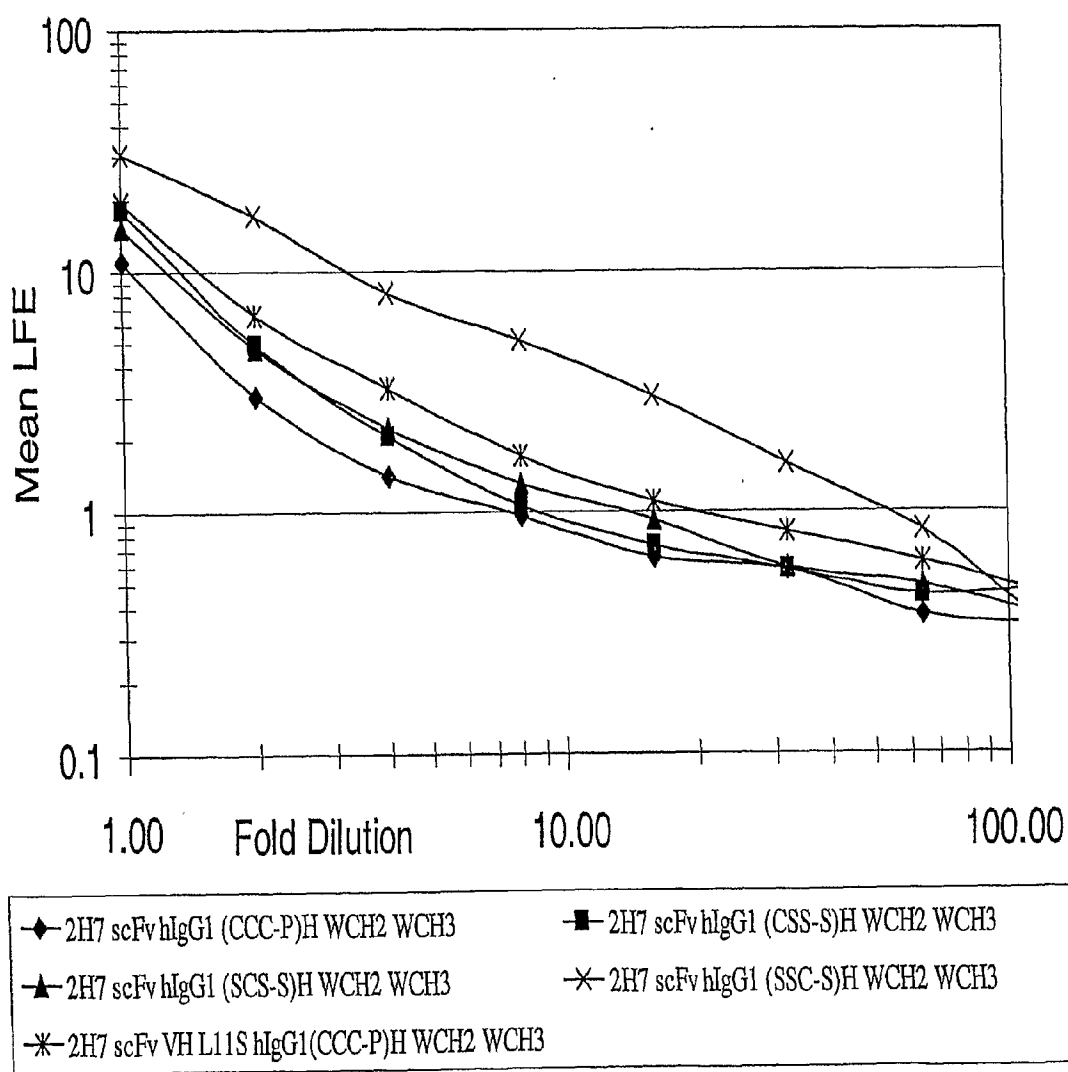
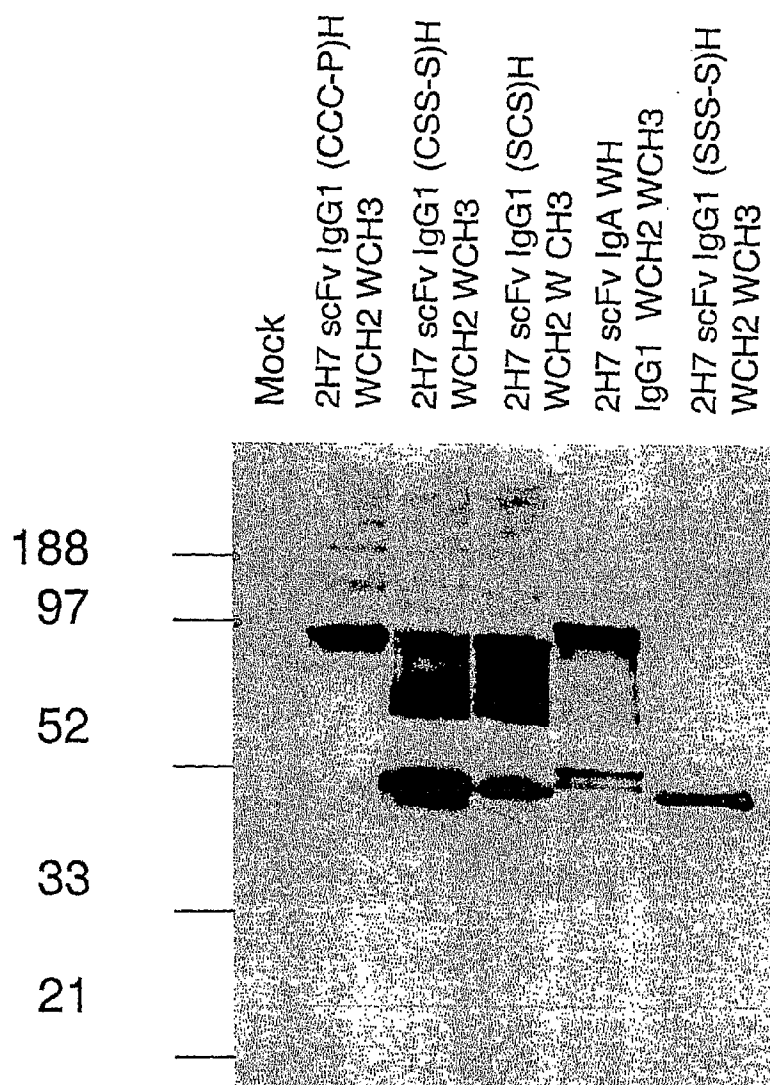


Fig. 35

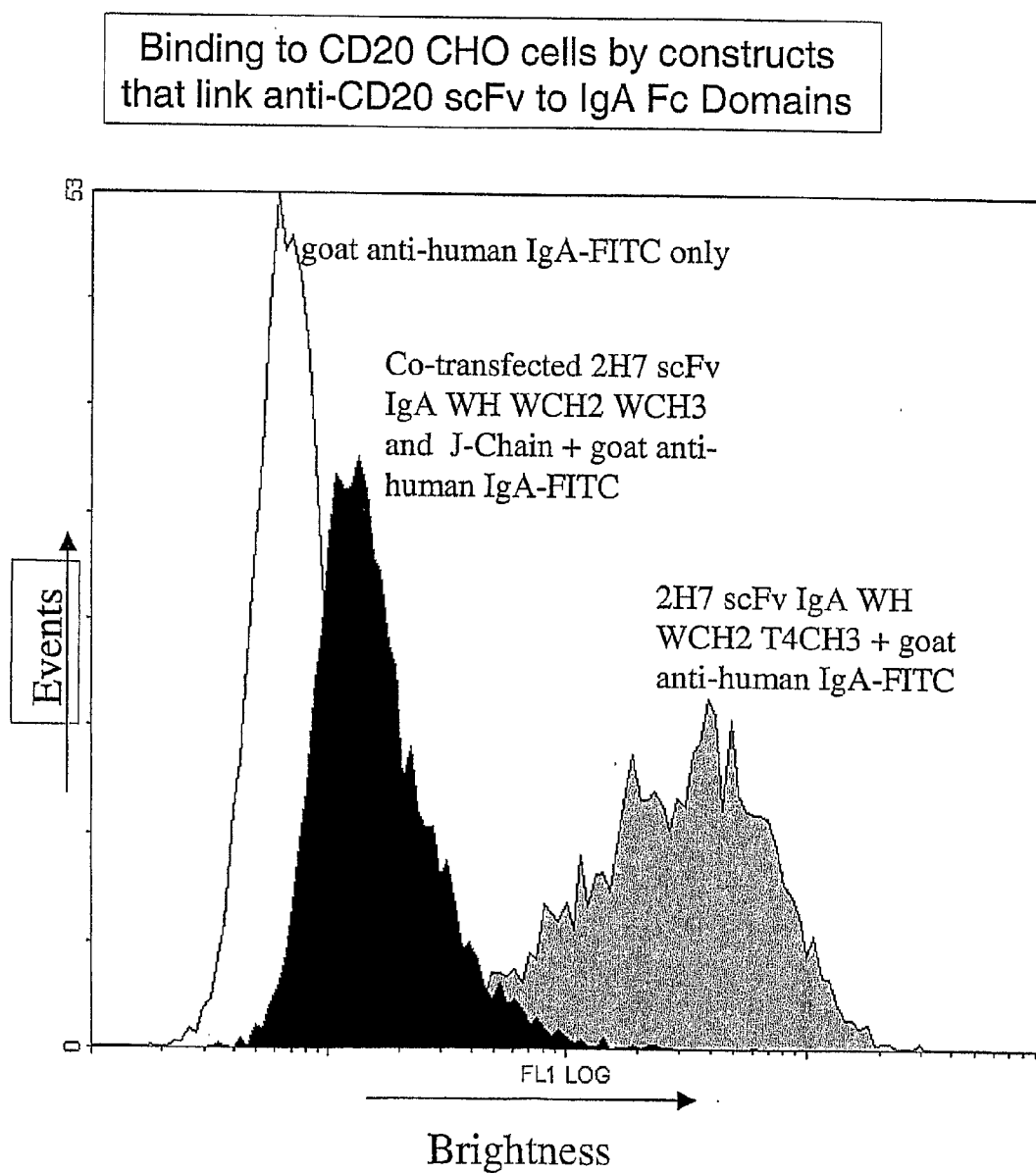
Immunoblot Analysis of protein immunoprecipitates
from COS transfections of 2H7 scFvIg Constructs



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Fig. 36



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Fig. 37

Titration of CD20 specific scFvIg Constructs
for ADCC Activity Using Whole Blood Effectors

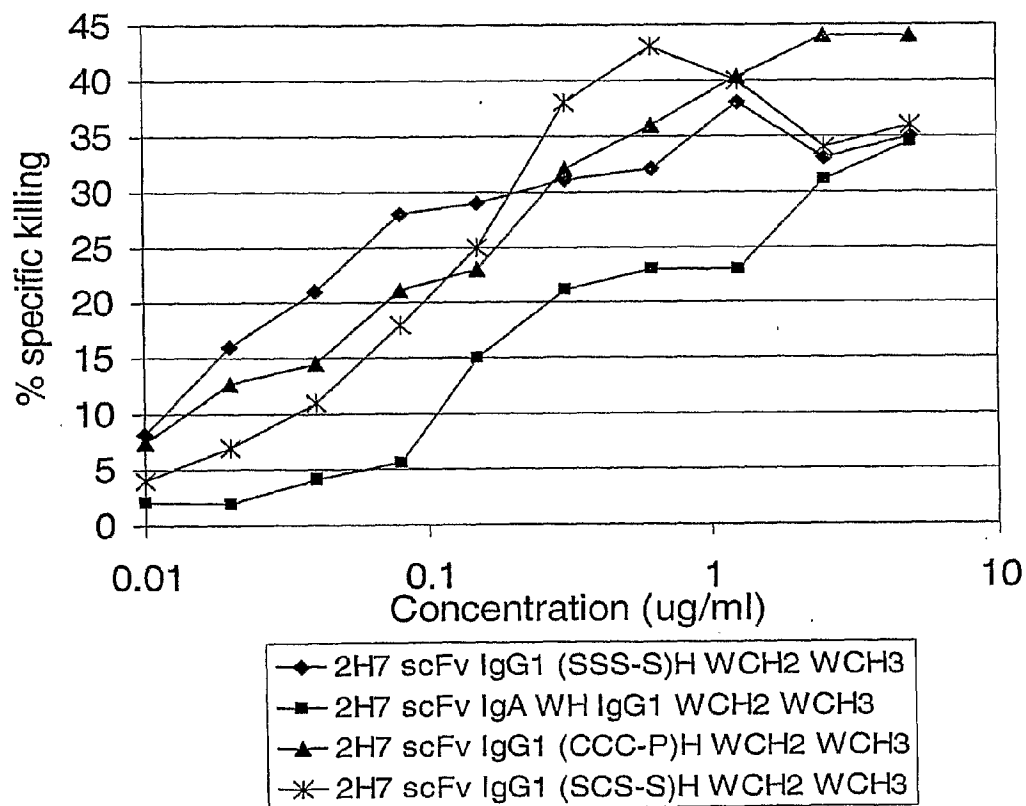
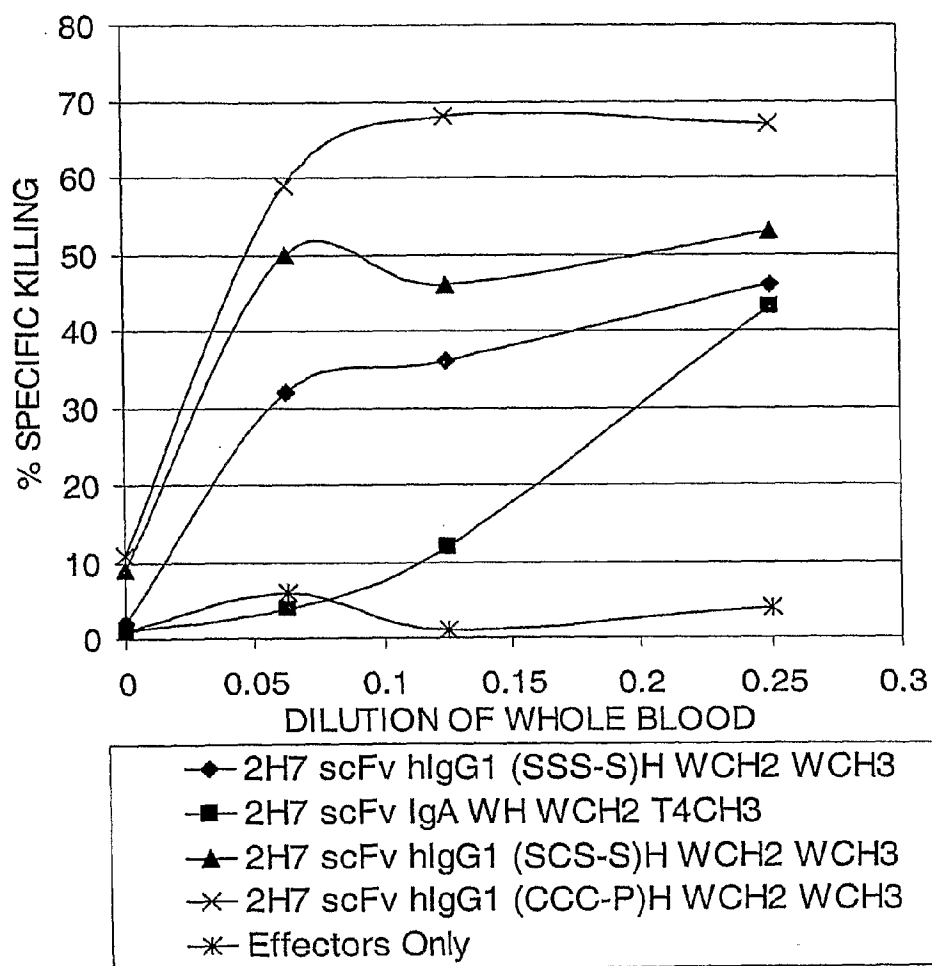


Fig. 38

ADCC Assay of anti-CD20 constructs with alternative tails
(Whole Blood Effectors / BJAB Targets)

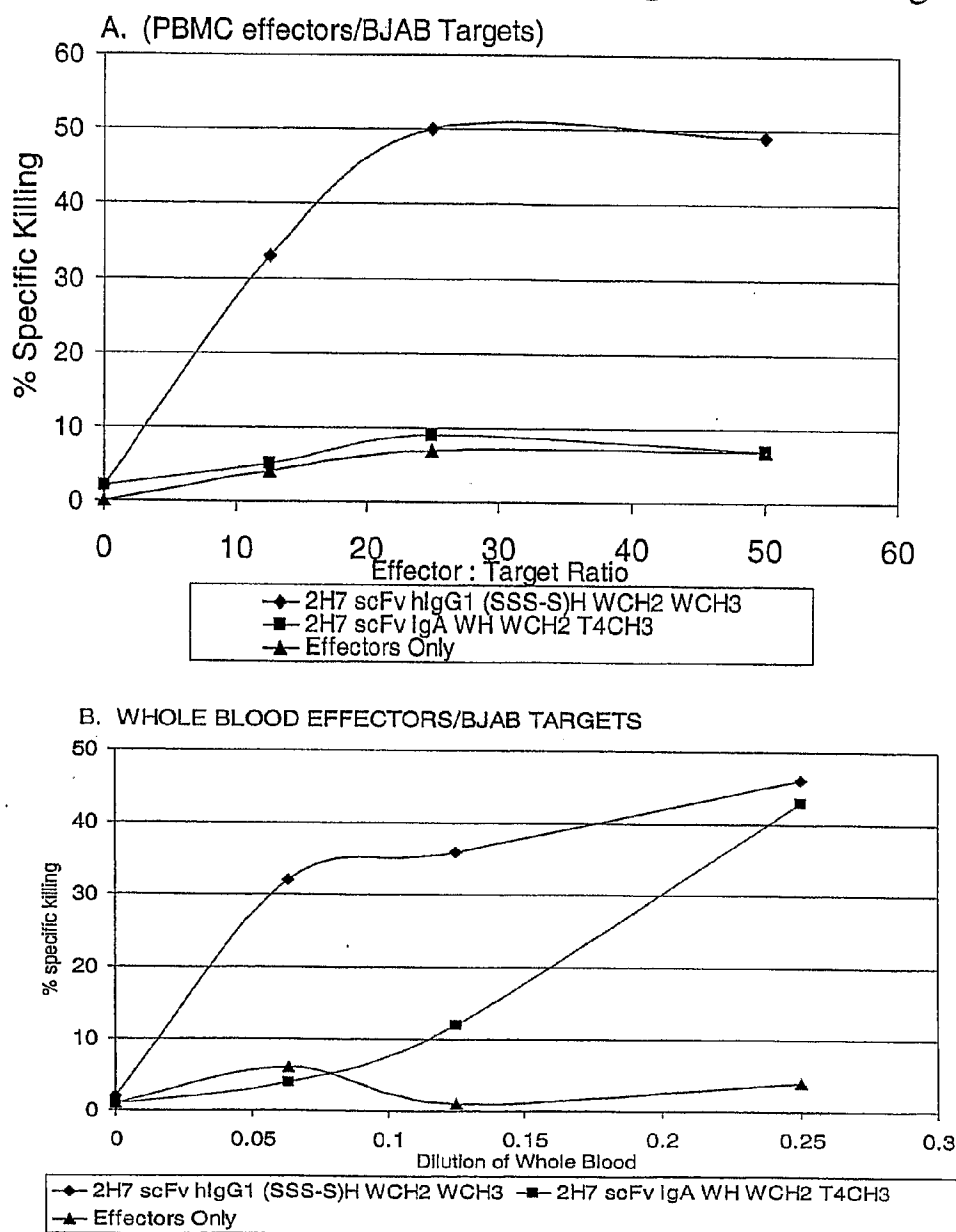


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Fig. 39

ADCC Assay of Anti-CD20 scFvIg Constructs
Using Different Effector Populations Against BJAB Targets

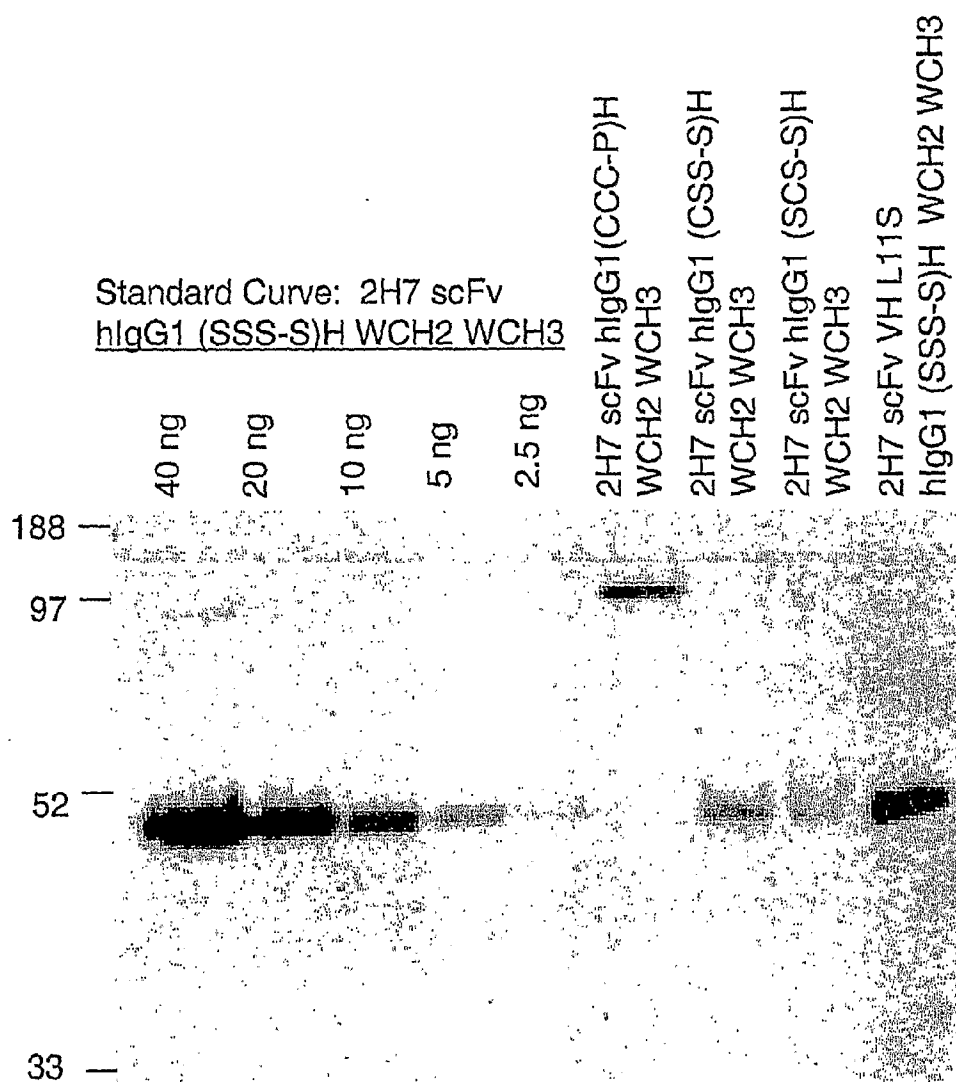


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Fig. 40

Immunoblot of 2H7 scFv Ig constructs from COS
Transfections (1 μ l/well) compared to a Concentration Standard

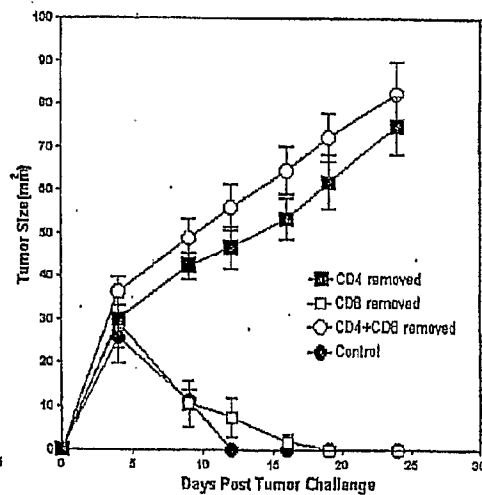
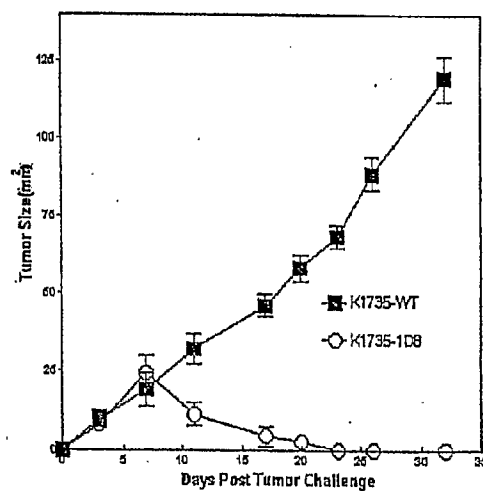
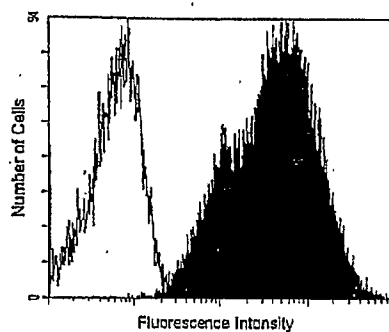


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Figures 41A, 41B and 41C

A.

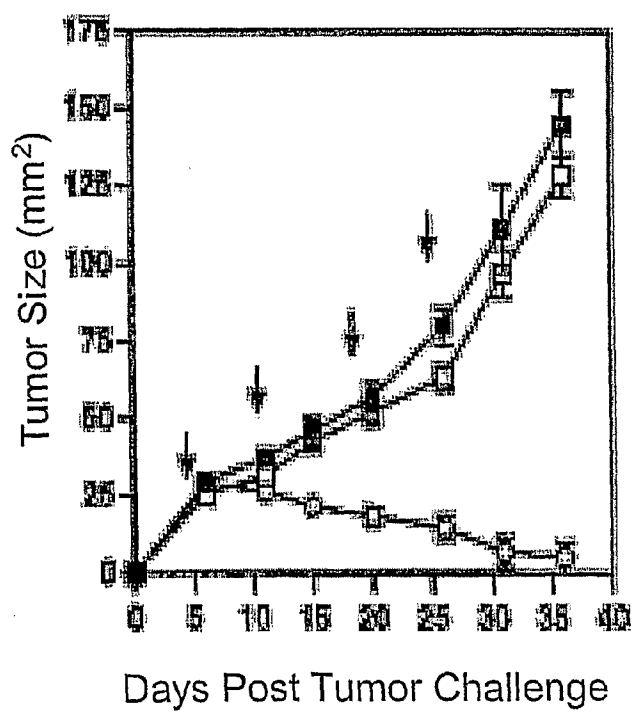


B.

C.

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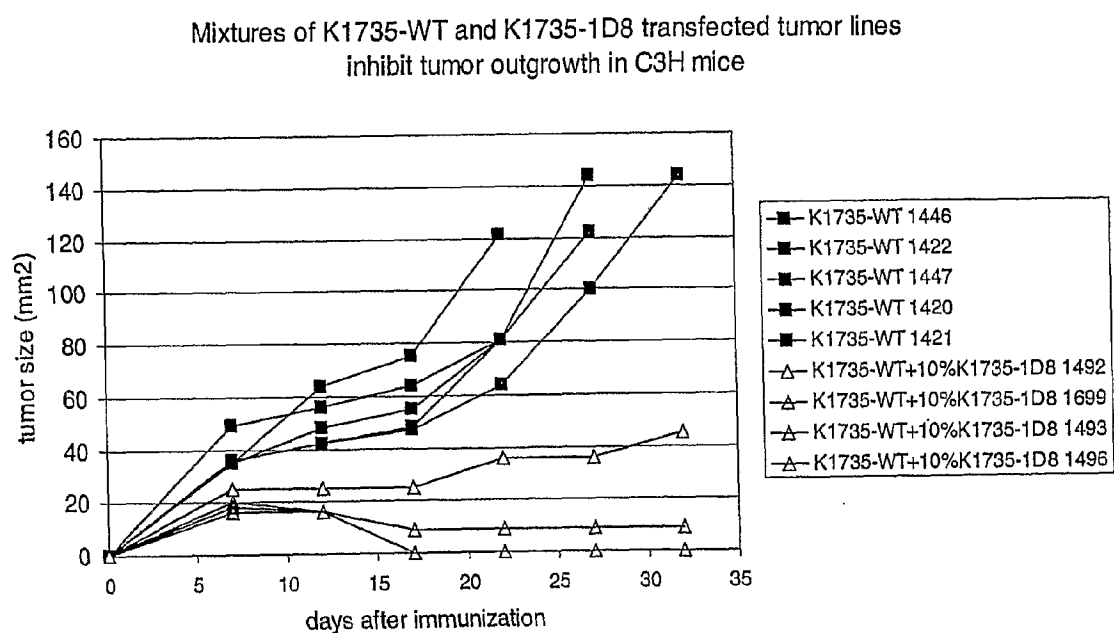
Fig. 42



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Fig. 43



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Fig. 44

Expression of anti-mouse CD137 (1D8) scFv-hIgG1 (SSS-S)H P238SCH2 WCH3
On the surface of panned Ag104-1D8 Transfected Tumor Cells

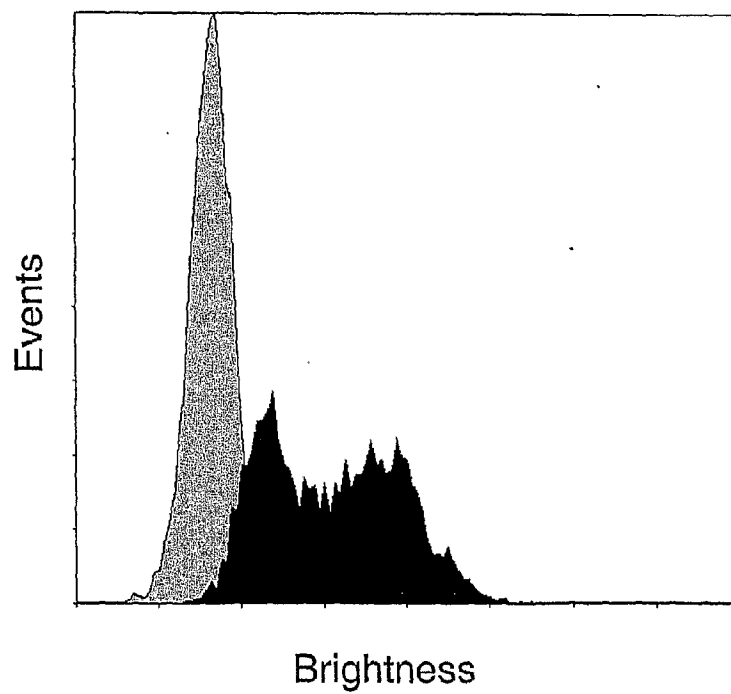


Fig. 45

Coomassie Stained SDS-PAGE Gel of 2H7 scFv Ig
Constructs

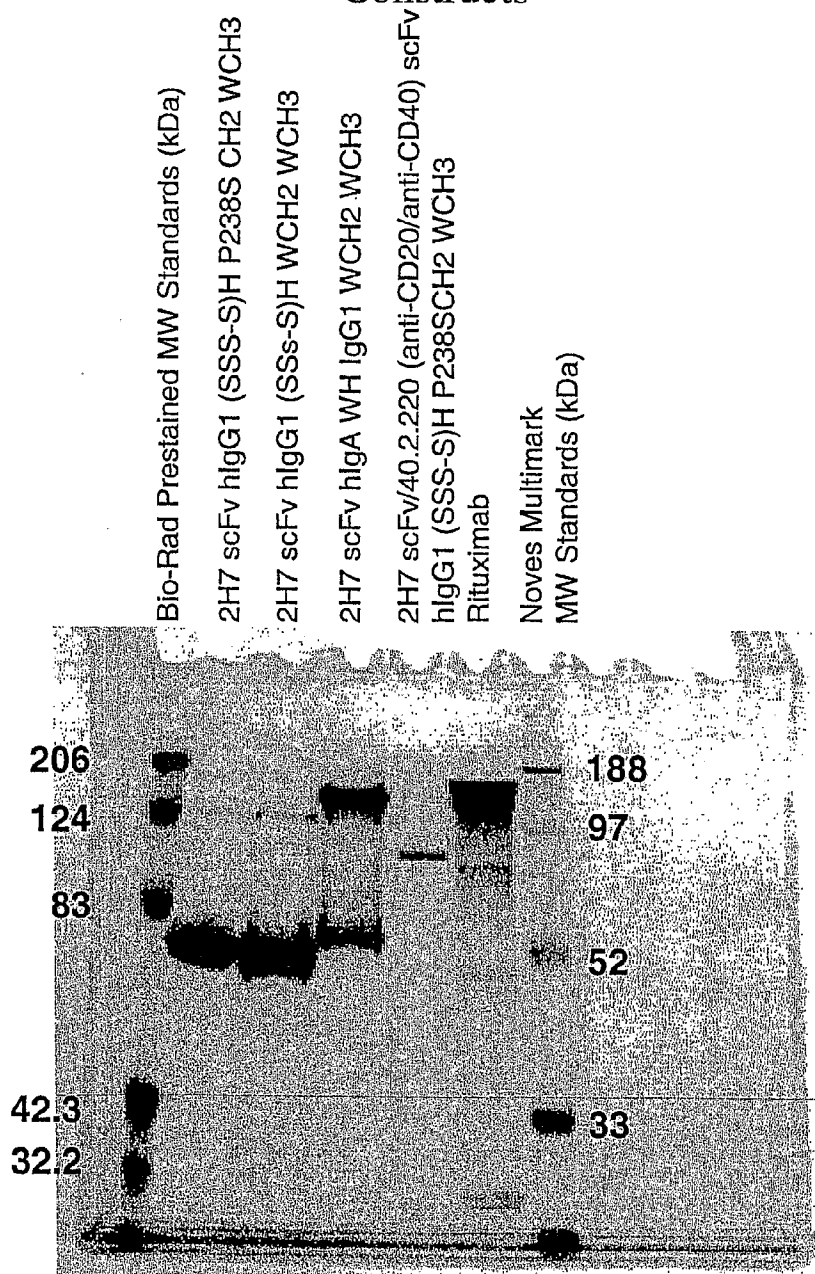
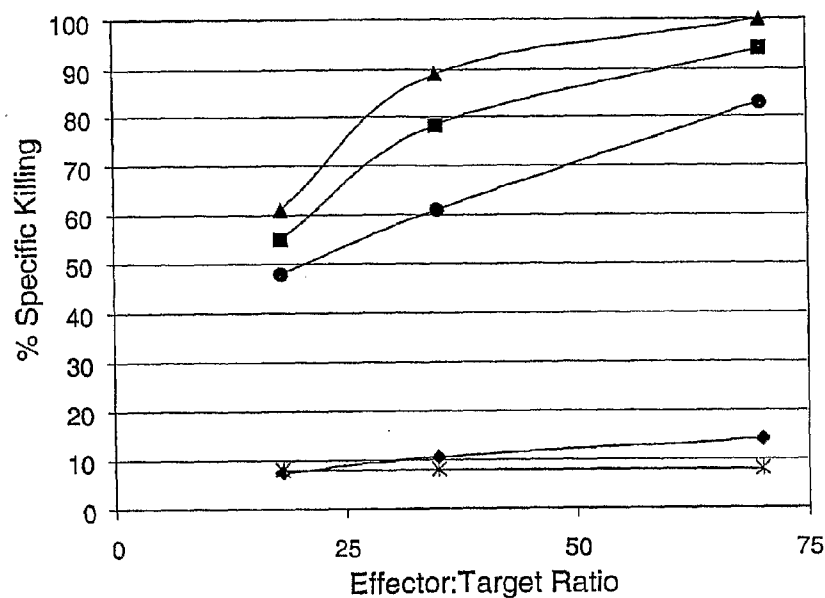


Fig. 46

ADCC mediated by 2H7 scFvIg Constructs by human PBMC effector cells against Bjab targets



- | | |
|--|------------------------|
| ◆ 2H7 scFv hlgG1(SSS-S)H P238SCH2 WCH3 | ● RITUXIMAB |
| ▲ 2H7 scFv hlgA WH IgG1 WCH2 WCH3 | * CELLS ALONE (W/O AB) |
| ■ 2H7 scFv hlgG1 (SSS-S)H WCH2 WCH3 | |

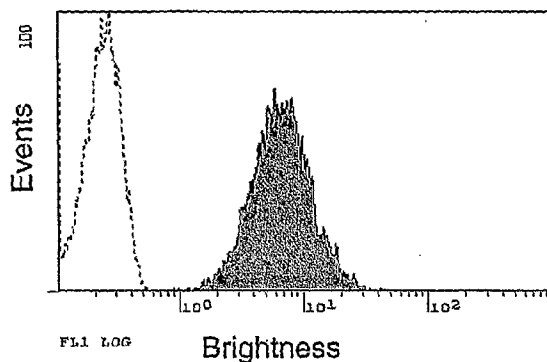
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Fig. 47

Cell surface expression of anti-human CD3 G19-4
scFv hIgG1 (SSS-S)H P238SCH2 WCH3-
hCD80TM/CT on Reh and T51 Cells.

Reh anti-CD3 (G19-4) scFv hIgG1 (SSS-S)H
P238SCH2 WCH3-hCD80TM/CT



T51 G19-4 scFv hIgG1 (SSS-S)H
P238SCH2 WCH3-hCD80TM/CT:

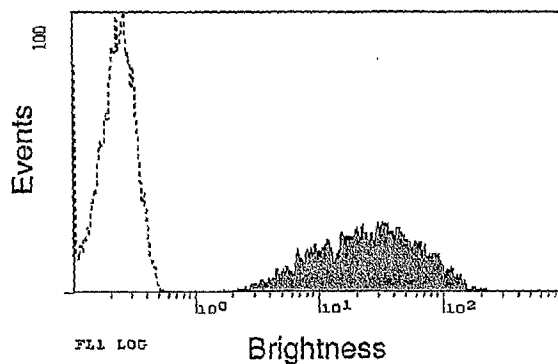
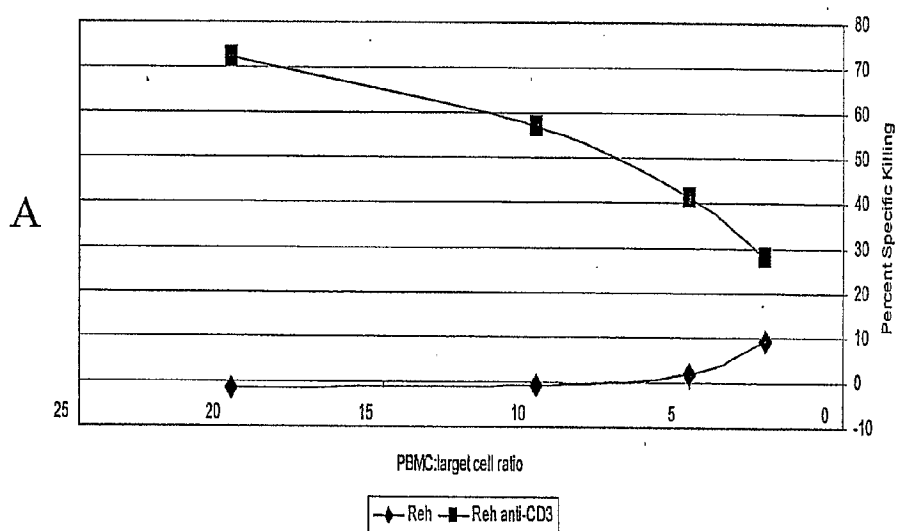


Figure 48.

Targeting of Cytotoxicity to Transfected Cell Lines by Surface expression of CD3 scFvIg

Cytotoxic activity of resting PBMC towards transfected Reh cells



Cytotoxic activity of resting PBMC towards transfected T51 lymphoblastoid cells

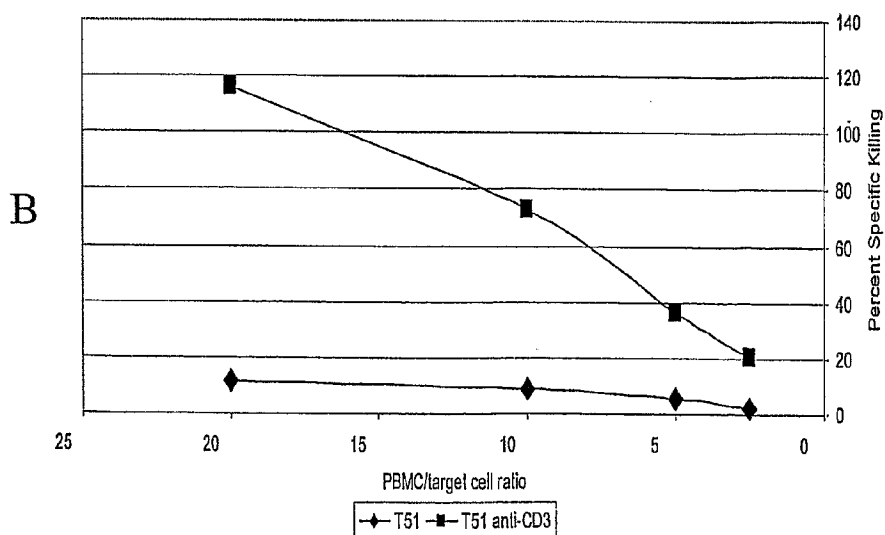
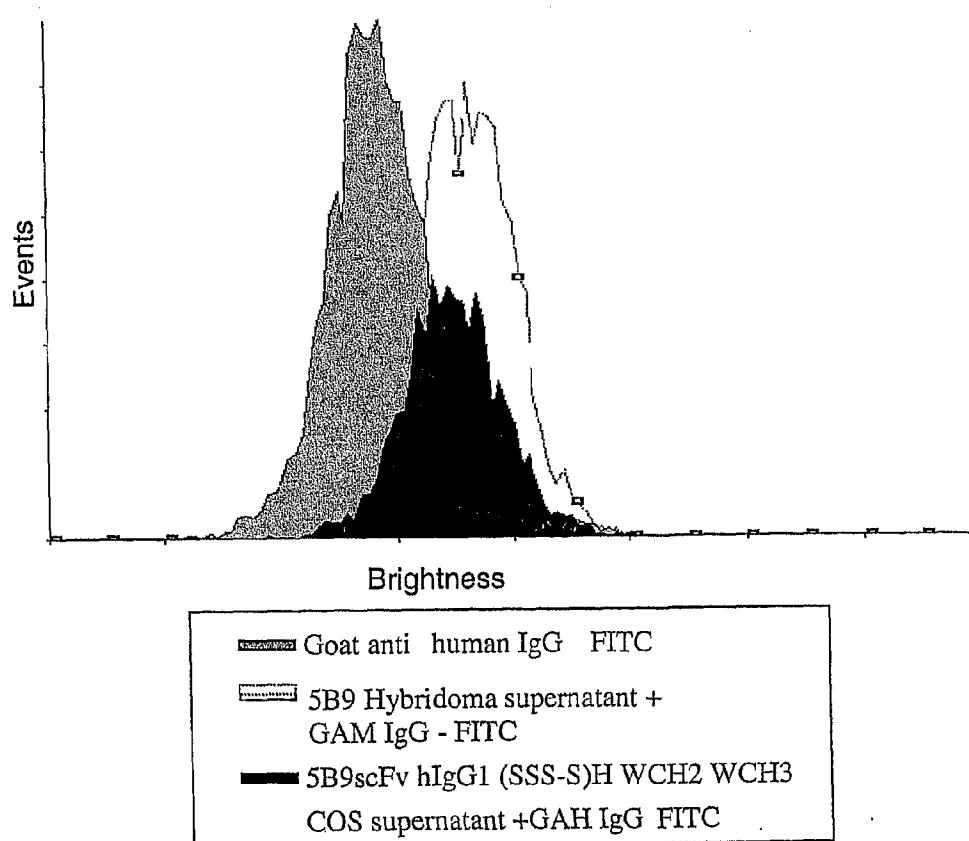


Fig. 49

Binding of 5B9, a mouse anti-human CD137 scFv hIgG1
(SSS-S)H WCH2WCH3 to stimulated human PBMC



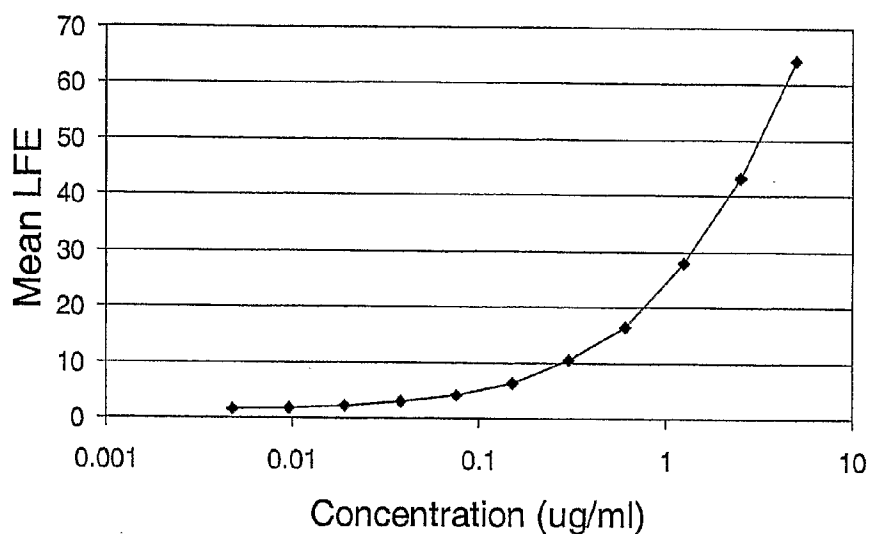
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Fig. 50

Effect of V_HL11S Mutation on CytoxB20
2H7 scFv hlgG1 (SSS-S)H WCH2 WCH3 Protein Expression

50A. Standard Curve: 2H7VH-L11S-IgG1 (SSS-S)H WCH2 WCH3



50B. CHO supernatant Brightness and Estimation of Protein concentrations from Standard Curve:

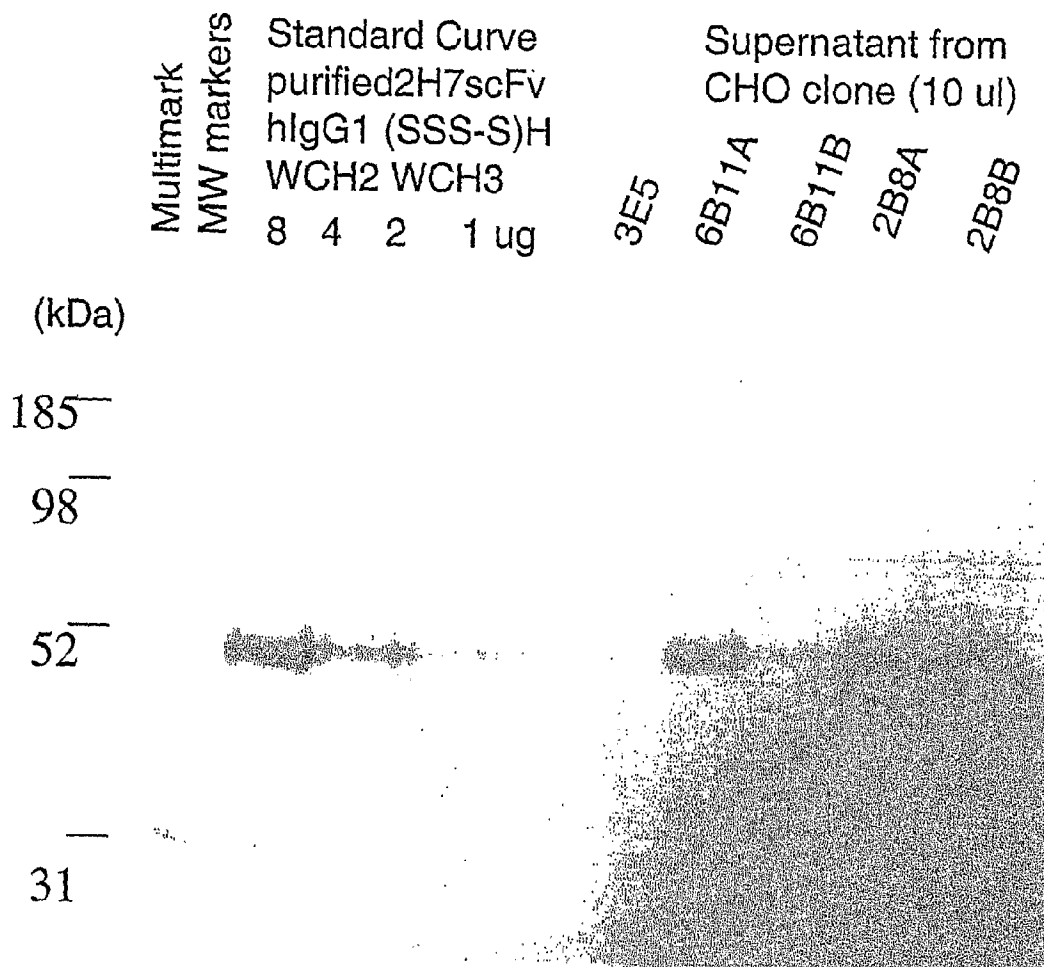
	CHO clone name				
	4F2	4F5	3E5	6B11A	2B8A
Mean LFE					
1/100	71.7	40.6	31.5	99.7	101.5
1/500	27.1	12.4	11.2	40.8	43
approx conc. µg/ml	600	225	125	1000	1250

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Fig. 51

Production Levels of 2H7scFv VH L11S hIgG1
(SSS-S)H WCH2 WCH3
From CHO Clone Culture Supernatants

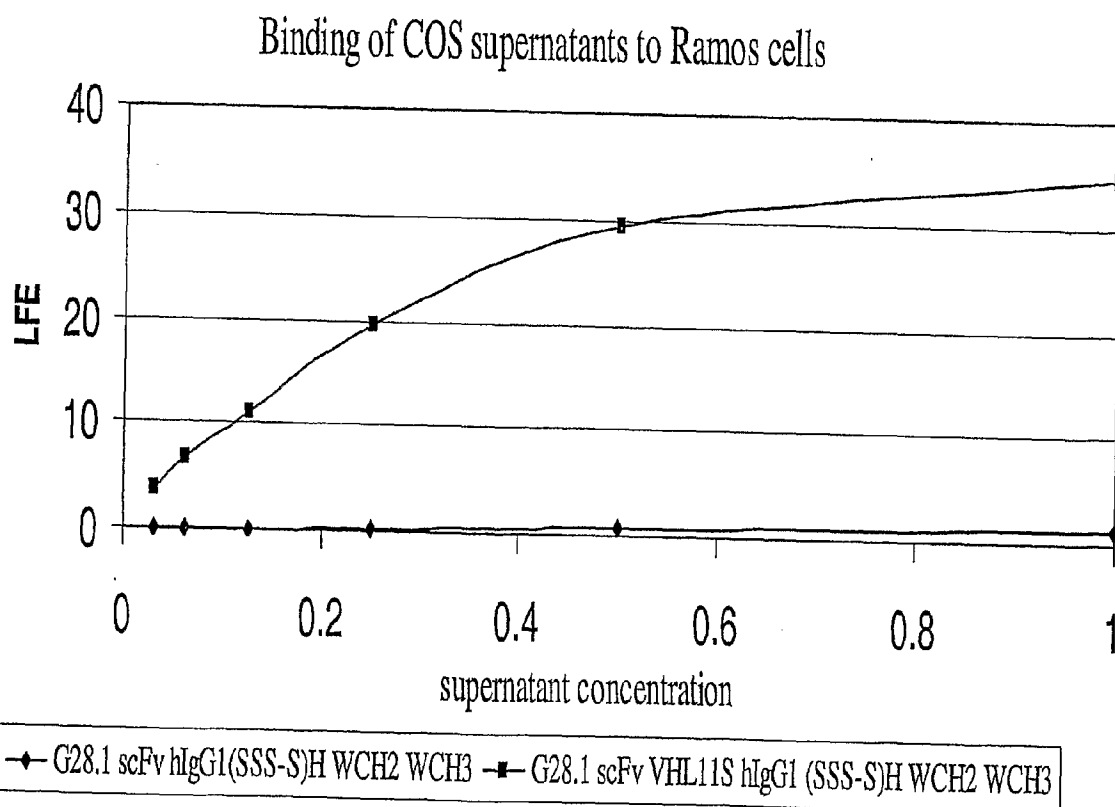


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Fig. 52

Effect of VHL11S Mutation on G28-1 scFvIg Construct Protein Production from COS cells



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Fig. 53

Immunoblot of G28-1 scFvIg Constructs

Increased Protein Levels in COS supernatants
transfected with G28-1scFv hlgG1 (SSS-S)H WCH2 WCH3
After Substitution of Leucine with Serine at position 11 of VH (VHL11S)

Fig. 53A.

Purified G28-1 (11/6/01)	G28-1 scFv hlgG1 (SSS-S)H				
scFv IgG1 (SSS-S)H	WCH2 WCH3				
WCH2 WCH3	1 ul/well				
80ng	40ng	20ng	10ng	A	B C D E

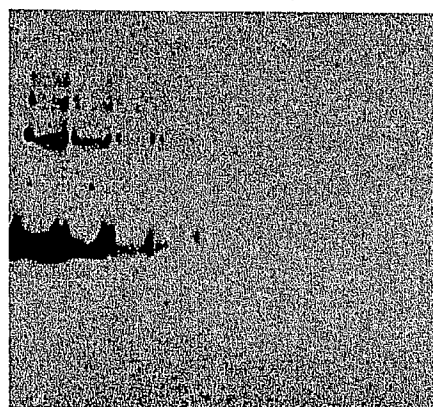


Fig. 53B.

Purified G28-1 (11/6/01)	G28-1VHL11S scFv hlgG1 (SSS-S)H				
scFv hlgG1(SSS-S)H	WCH2 WCH3				
WCH2 WCH3	1 ul/well				
80ng	40ng	20ng	10ng	A	B C D E

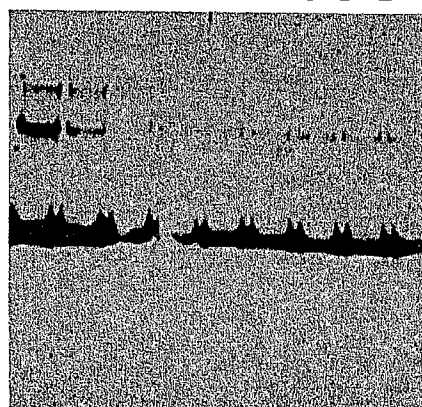


Fig. 54

Binding of 2H7 scFvIg Constructs with Altered Hinges and CH3 domains to CD20 CHO Cells

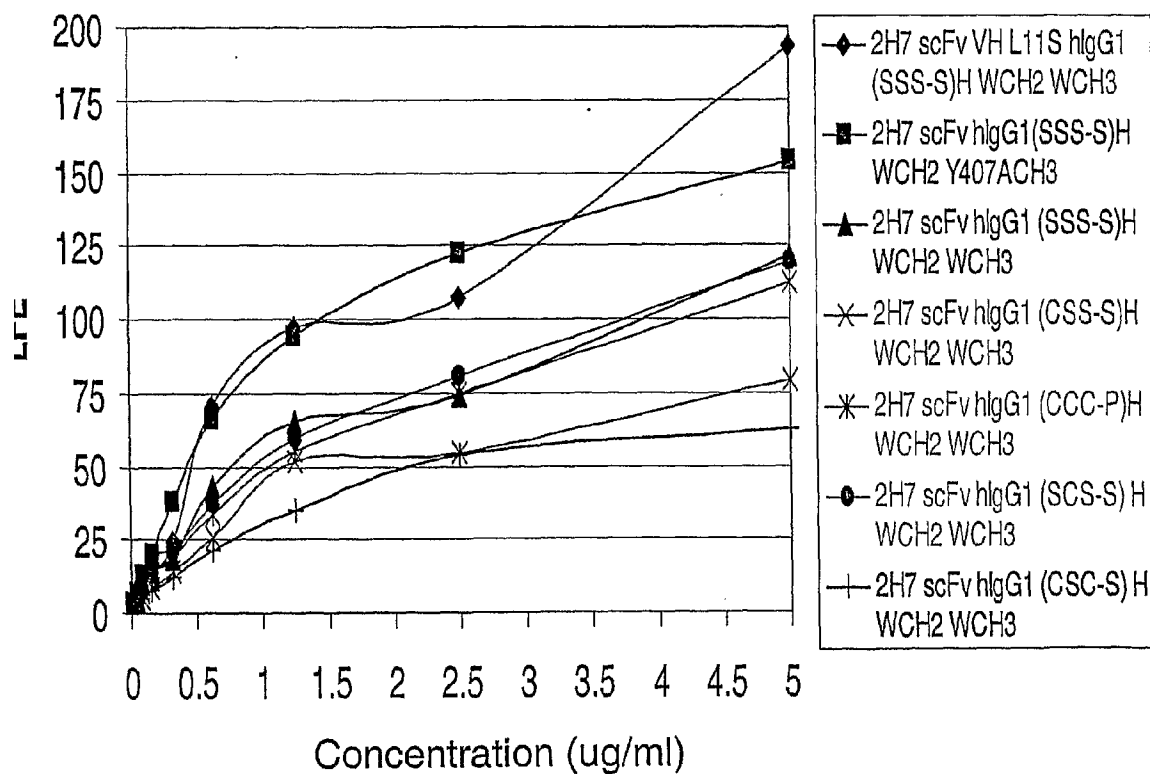


Fig. 55

ADCC Activity of 2H7 scFvlg constructs Against
BJAB Targets and PBMC Effectors

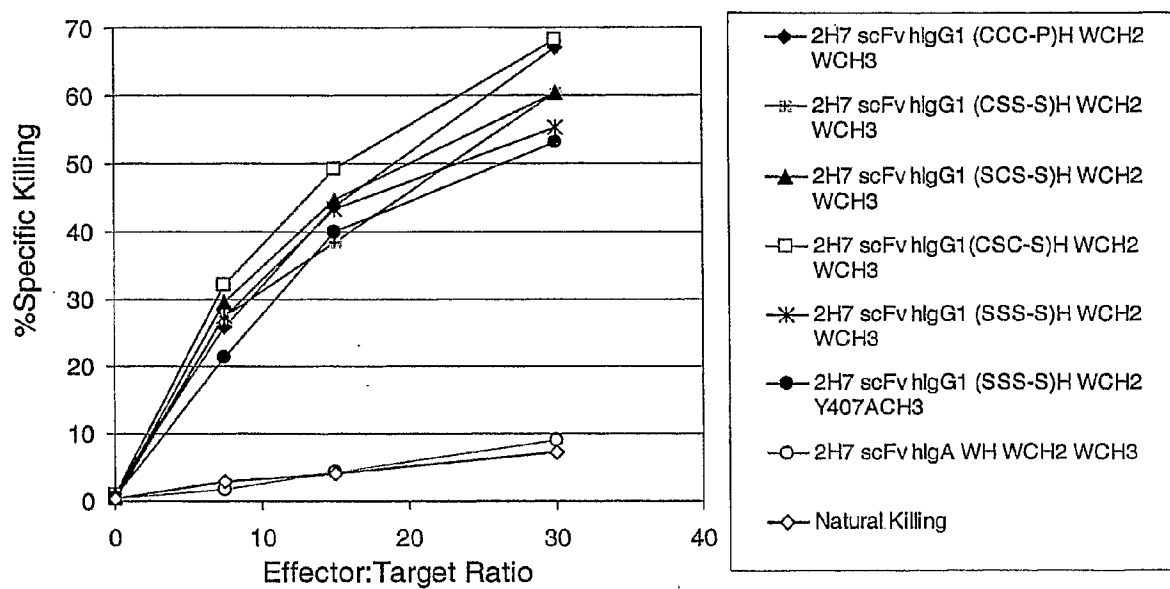
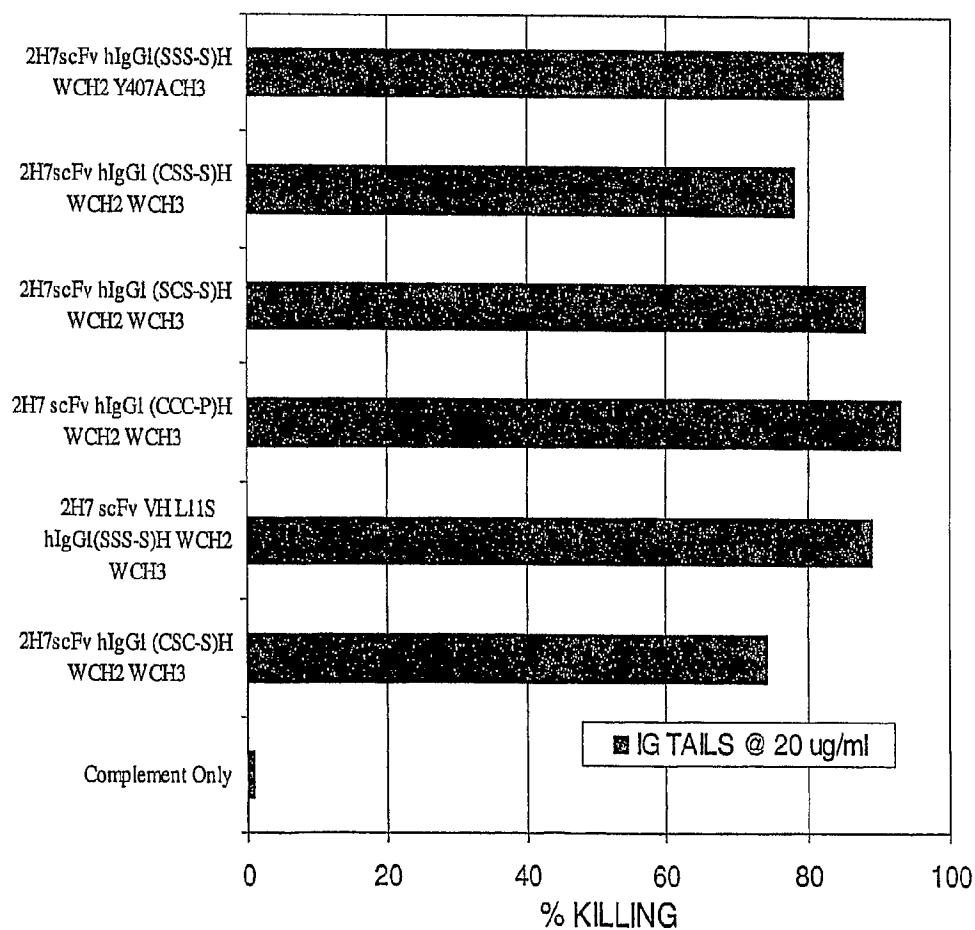


Fig. 56

Complement Activity of 2H7 scFvlg Constructs With Ramos Target Cells



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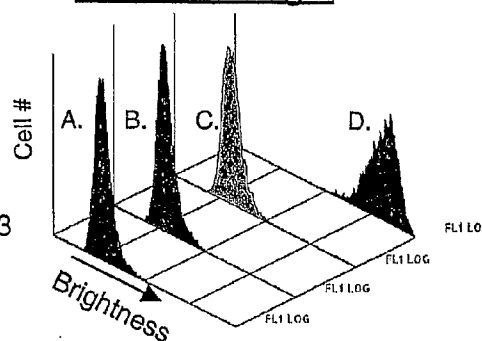
PCT/US2003/041600

Fig. 57

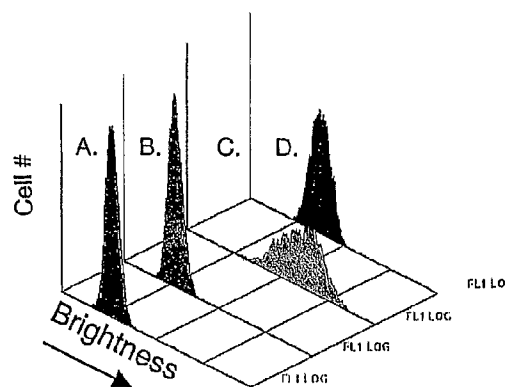
Binding of 2H7 scFvIg Derivatives to CD20CHO Cells

- A. ■ No fusion protein
- B. ■ 2H7 scFv hlgE CH2CH3CH4
- C. ■ 2H7 scFv hlgA WH WCH2 WCH3
- D. ■ 2H7 scFv hlgG1 (SSS-S)H WCH2 WCH3

FITC anti-hlgG



FITC anti-hlgA



FITC anti-hlgE

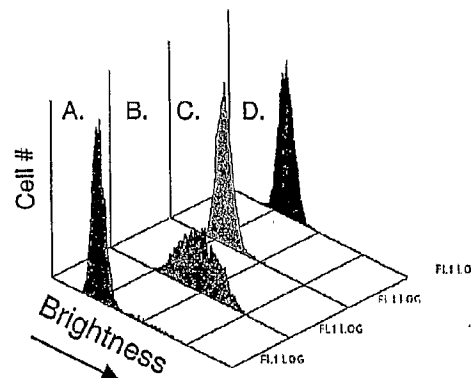


Fig. 58

Fig. 58A. 2H7 scFv VH L11S human IgE (WCH2 WCH3 WCH4)
Binding to CD20 CHO at 30 ug/ml

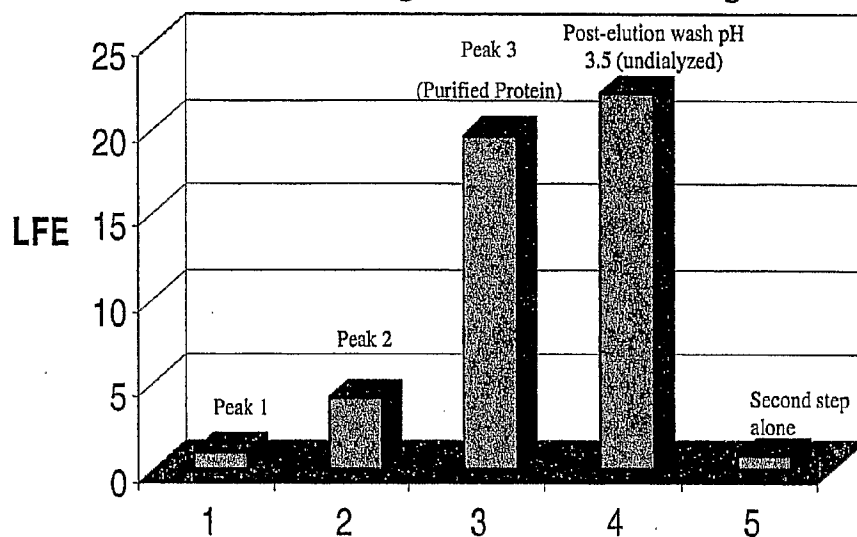


Fig. 58B. ADCC Activity of 2H7 VHL11S IgE (WCH2 WCH3 WCH4)
Protein Fractions with **PBMC** Effectors and Bjab Targets

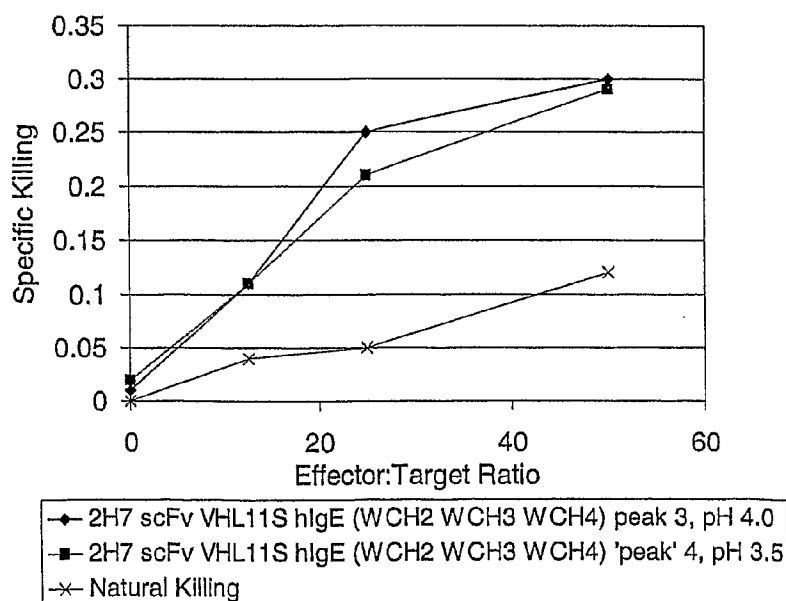
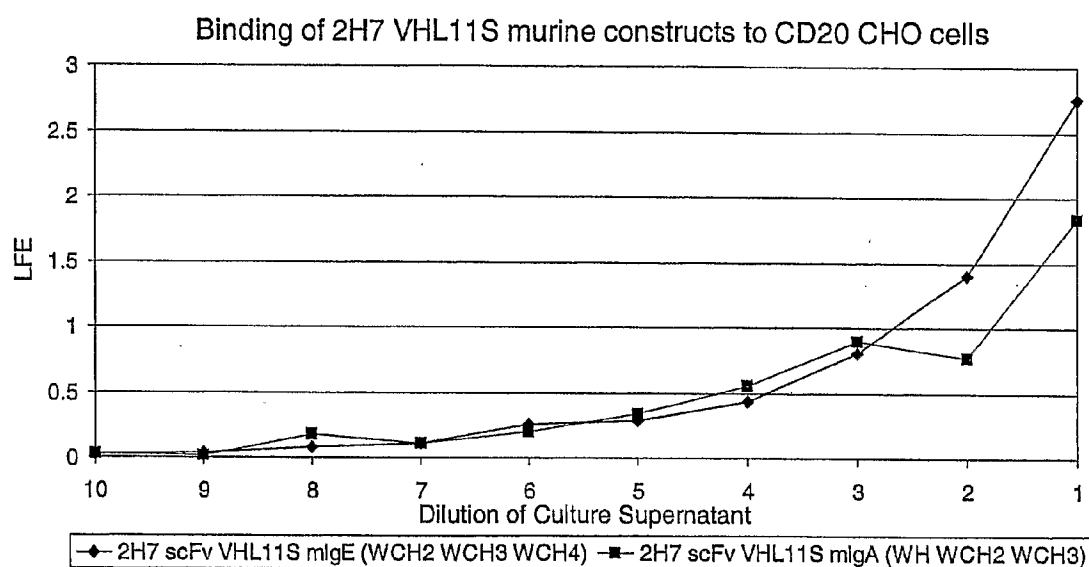


Fig. 59

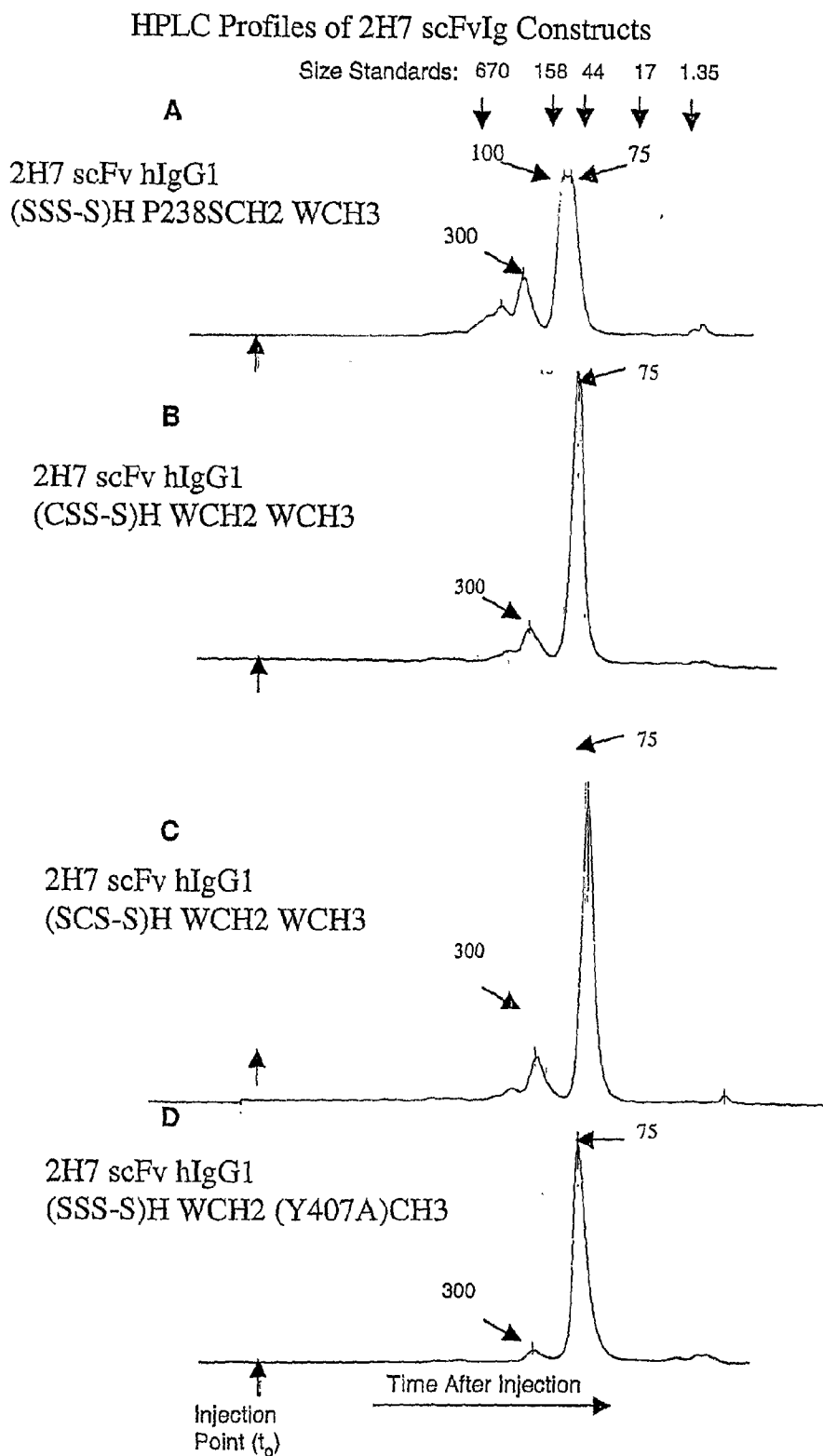
Binding Data for COS derived α -CD20 (2H7) scFv VHL11S
mIg E (WCH2 WCH3 WCH4) and
mIgA (WH WCH2 WCH3) Tailed Molecules



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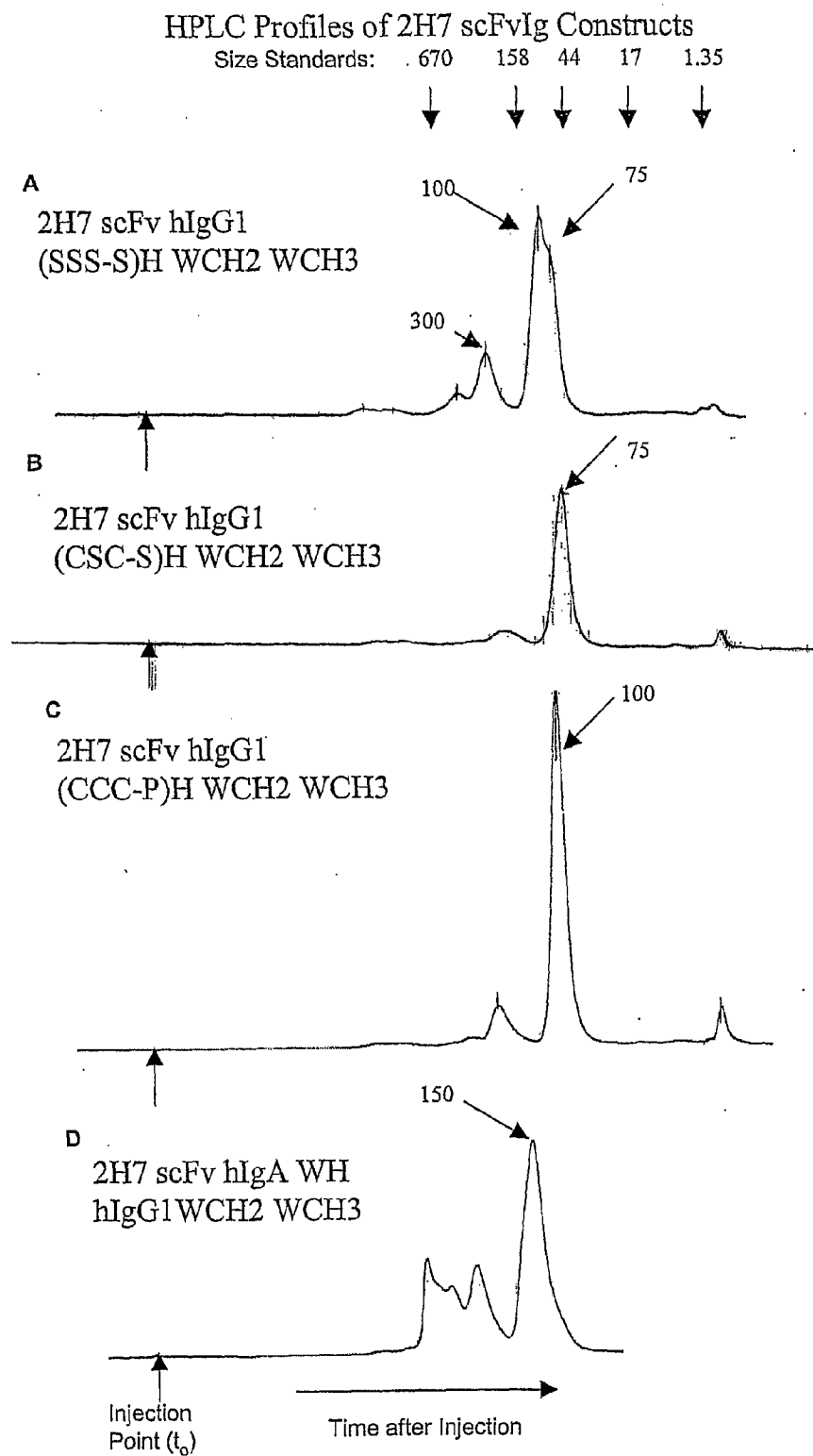
Fig. 60



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Fig. 61



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Fig. 62

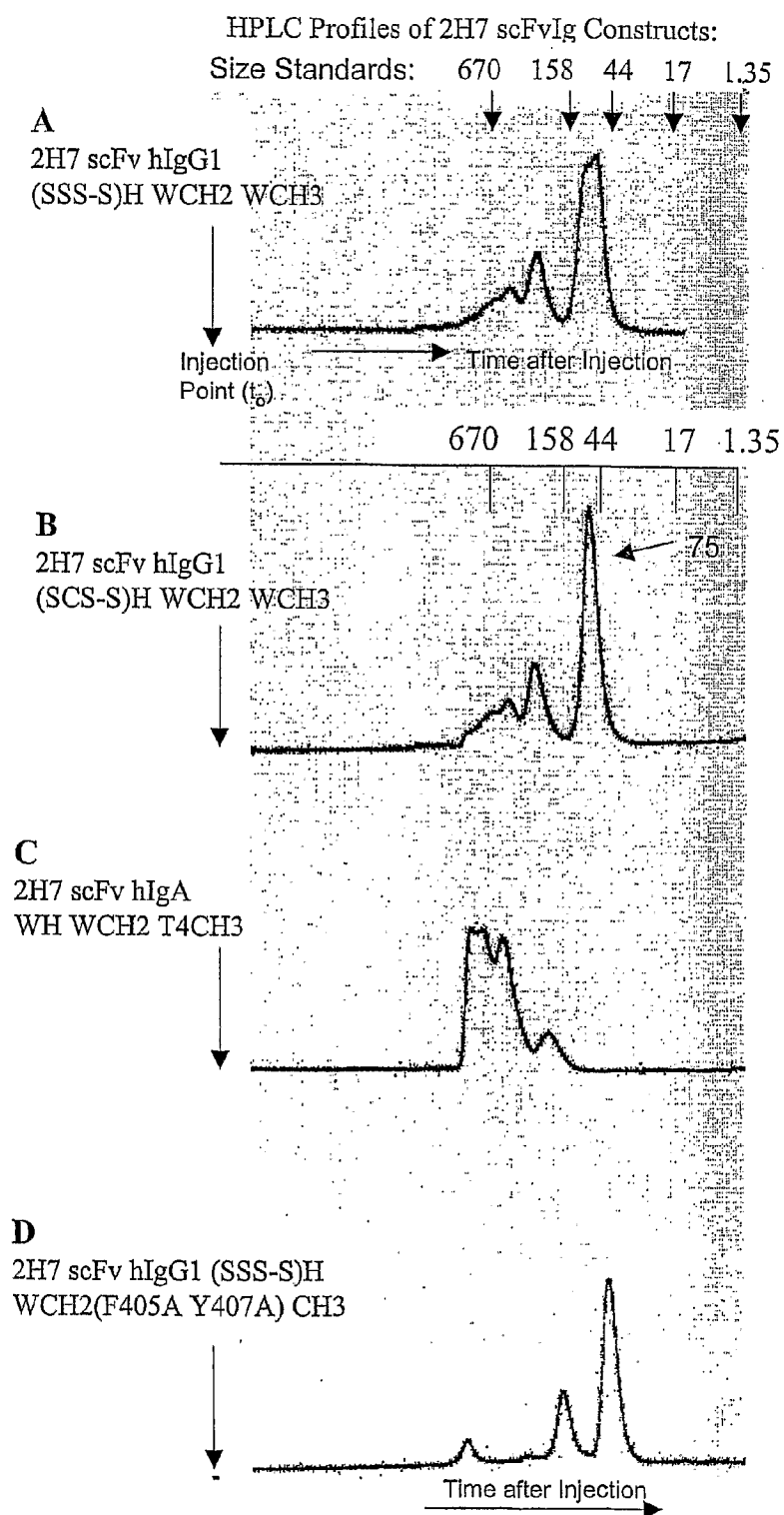


Fig. 63

Binding of Purified Proteins from COS Supernatants
to CD20 CHO cells:
Differential Effects of CH3 Mutations on Binding

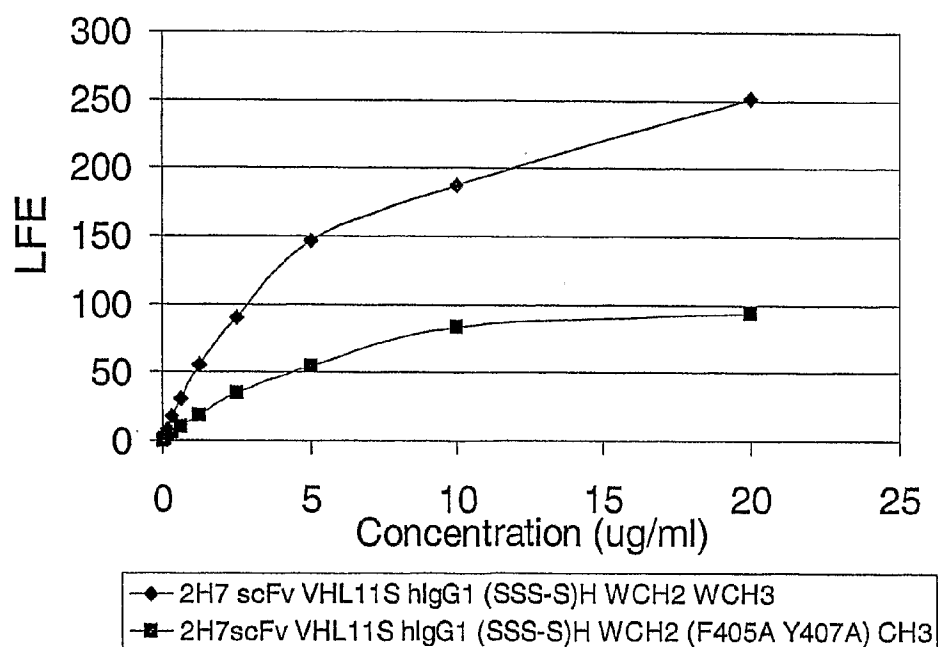
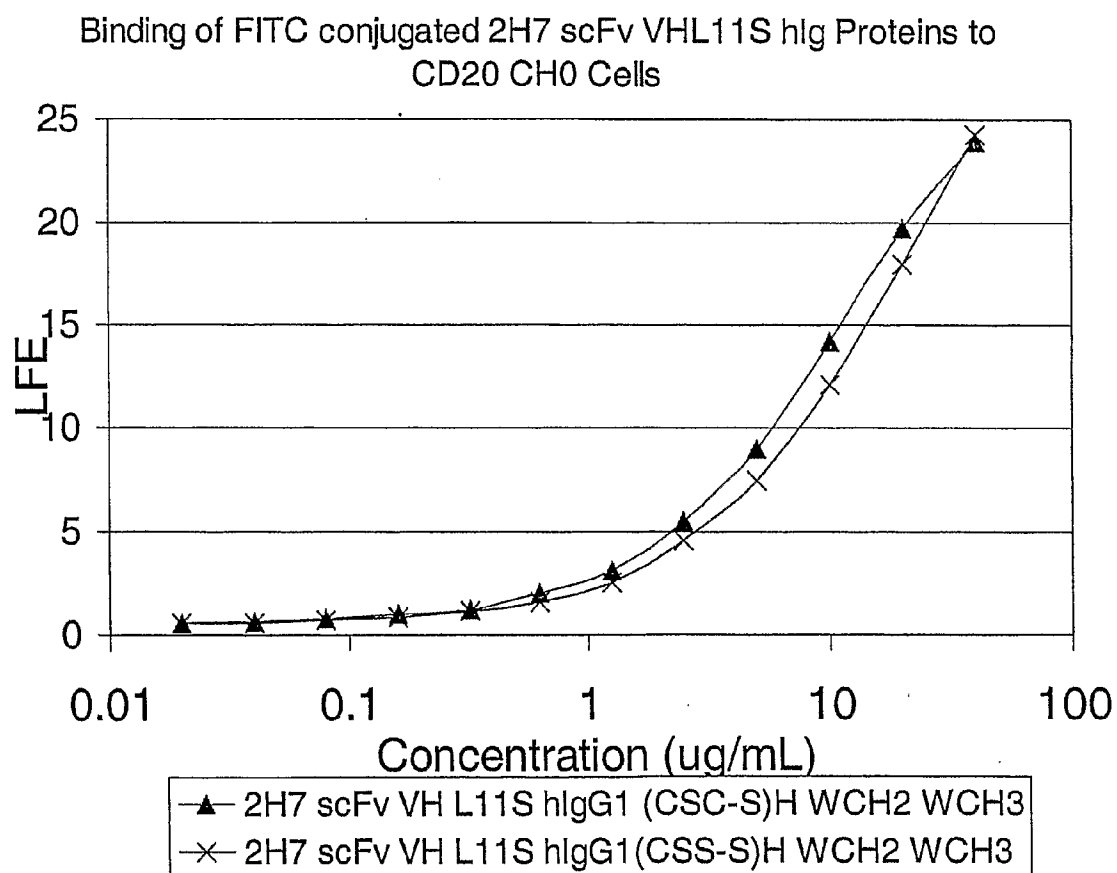


Fig. 64

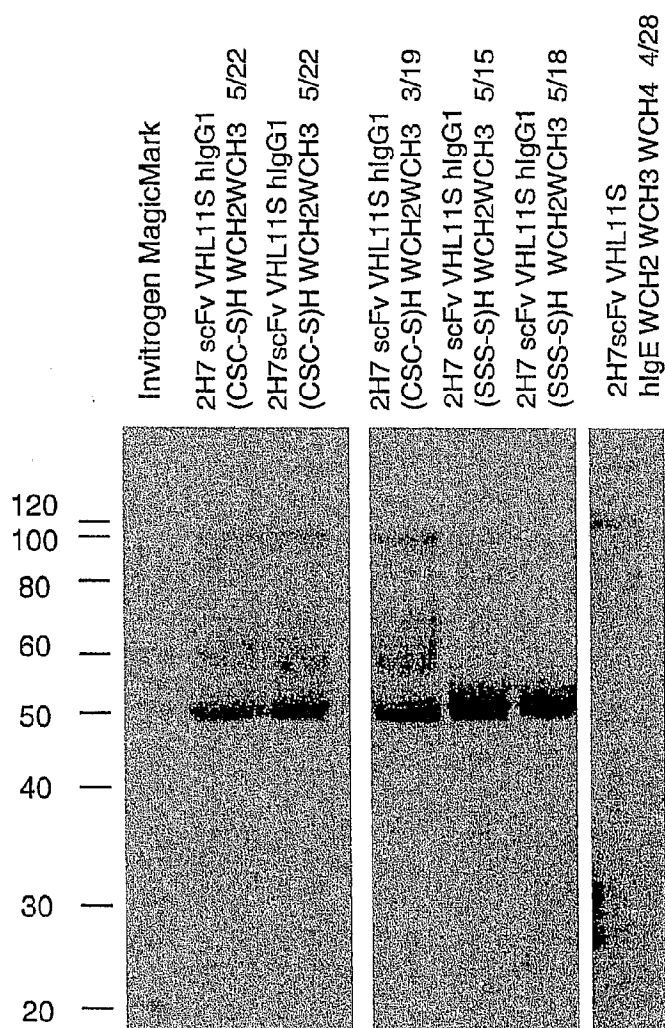


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Fig. 65

Nonreducing SDS-PAGE on Protein A-Purified Lots
of 2H7 scFv VHL11S hlg Constructs (10 ug/lane)



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Fig. 66

Alterations in Human IgG Fc sequence
that differentially change effector function efficiency

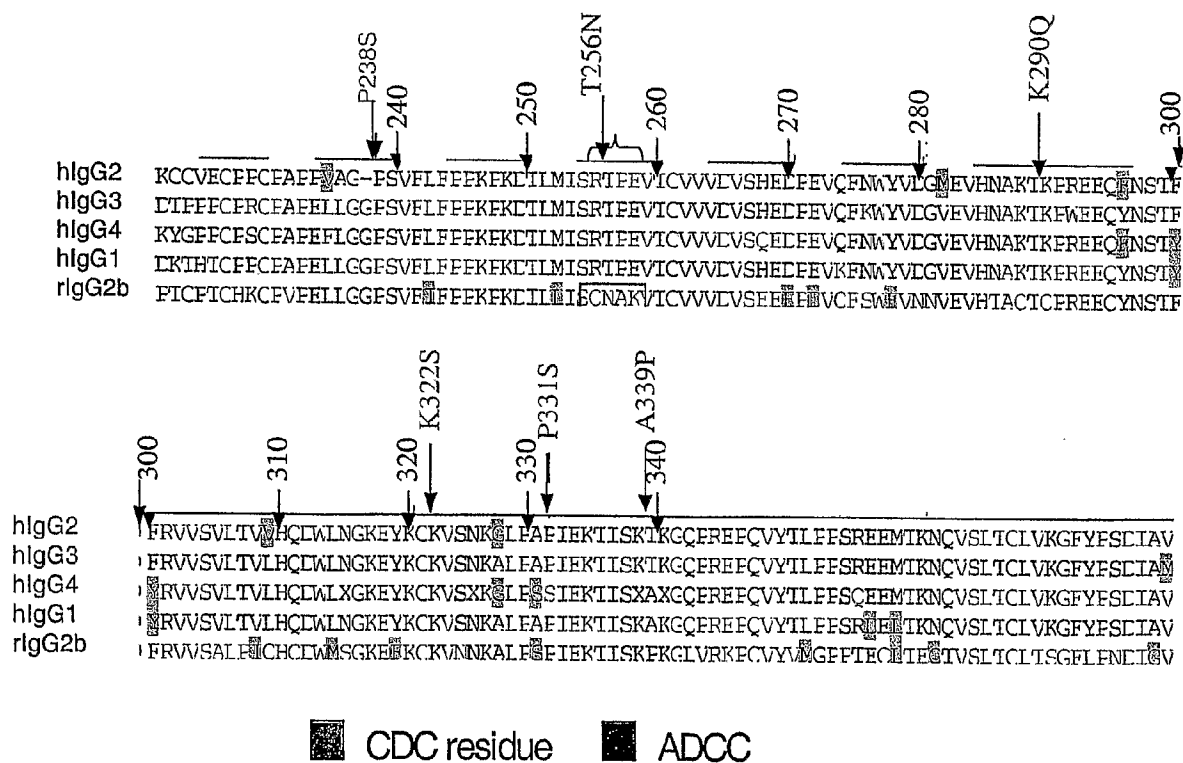


Figure 67.

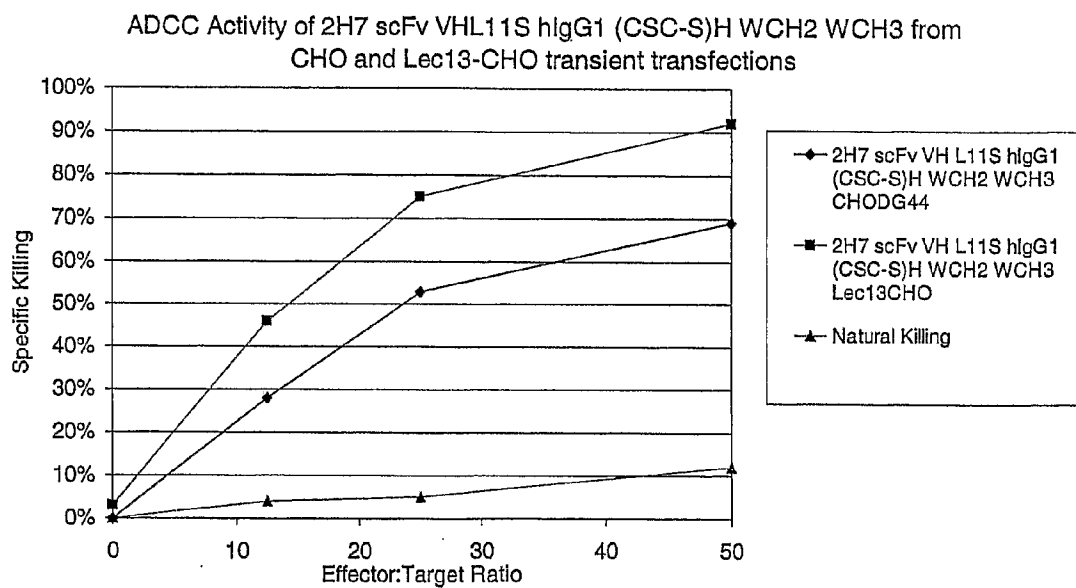


Fig. 68

CD16(ED) hIgG1(SSS-S)H P238S CH2 WCH3 high and low affinity alleles expressed as soluble molecules

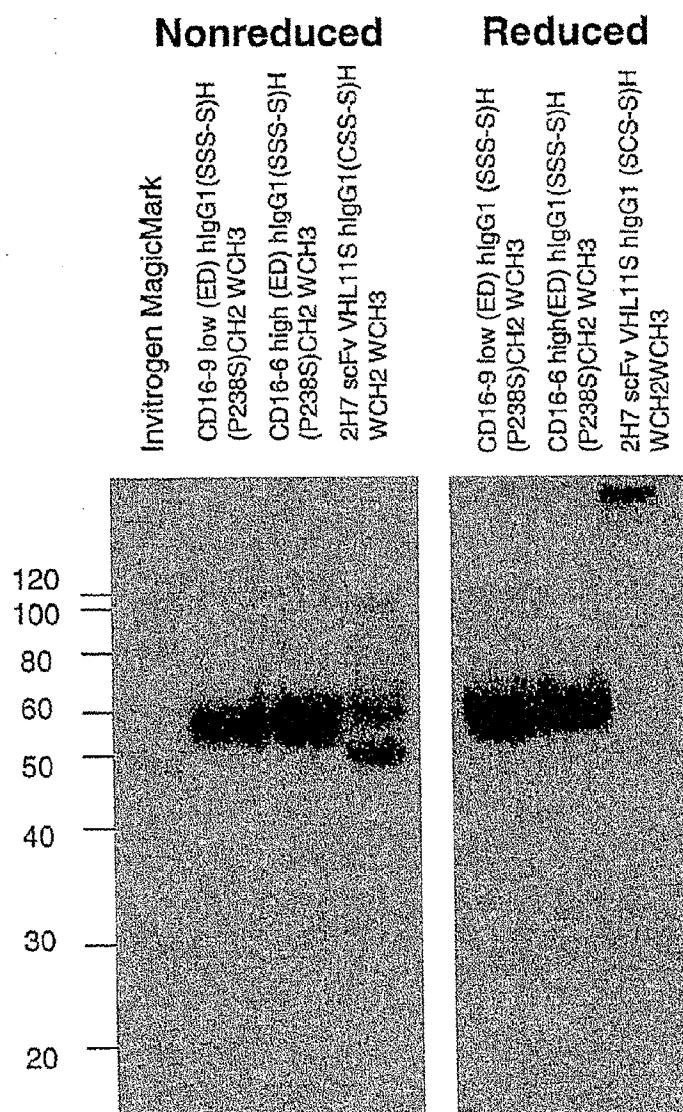
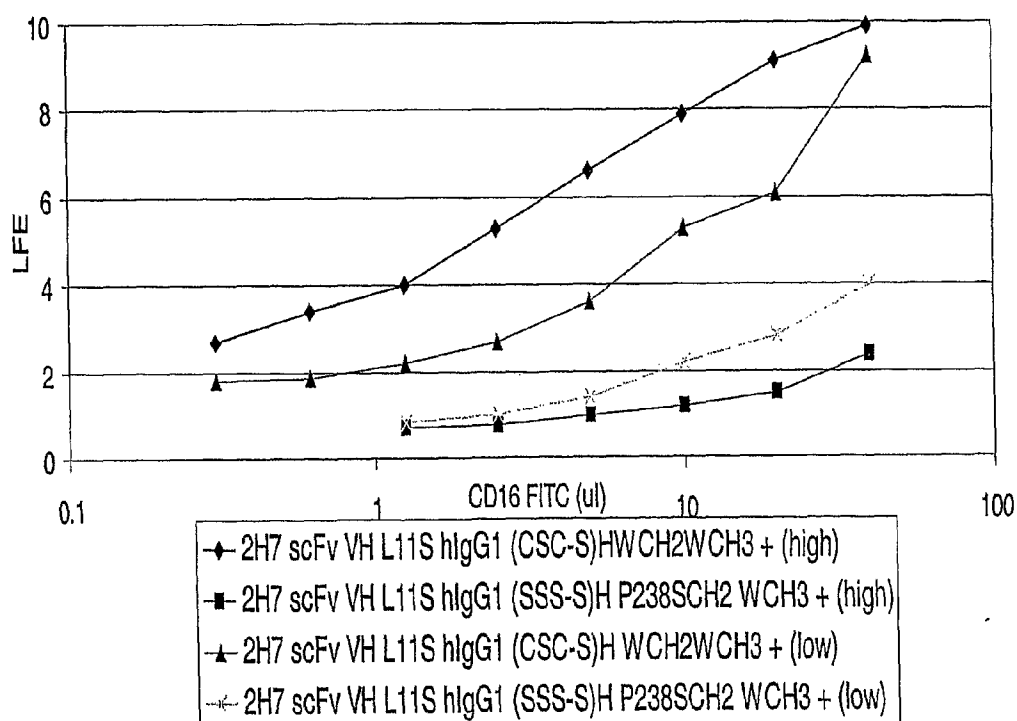


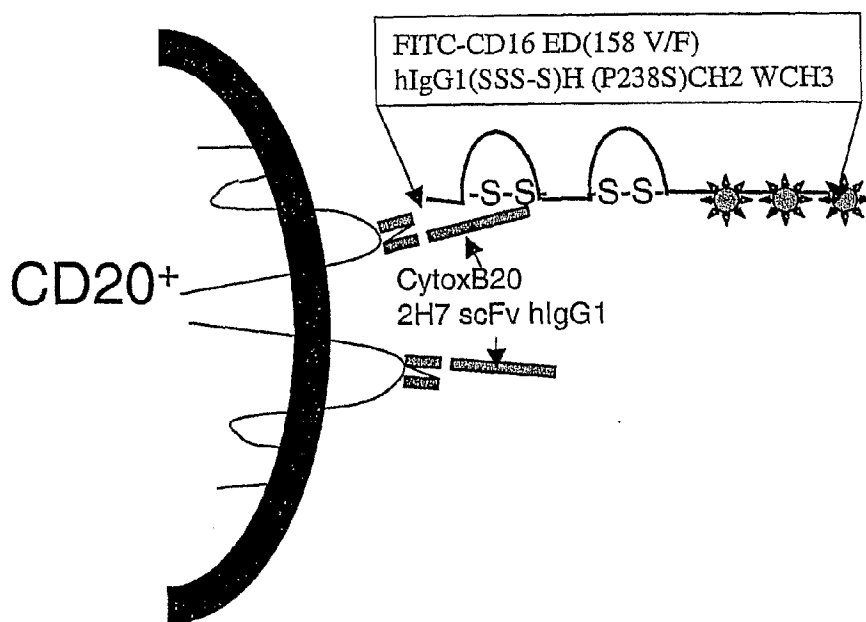
Fig. 69

Binding of soluble CD16-FITC high and low affinity fusion proteins
to 2H7 scFv VHL11S hlgG1 (CSC-S)H WCH2WCH3 or
(SSS-S)H (P238S)CH2WCH3 on CD20CHO Targets



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Binding of FITC Labeled, Recombinant Human CD16(ED) extracellular domain -Ig Fusion Protein to CytosB Derivatives on CD20 CHO Cells



Expression of surface displayed SMIPs links modified cDNAs with the altered fusion proteins

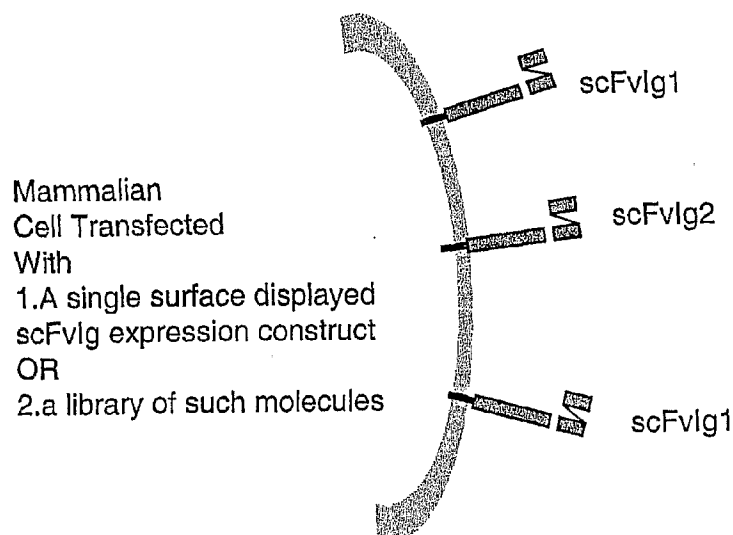


Fig. 71

CD37 mAbs and scFvIg Induce Apoptosis

scFvIg	Bjab Staining	Annexin V Positive	
	No scFvIg	17.5	
	2H7 MH	27	
	G28-1 MH	30.6	
	G28-1 IgAH	28.9	
	HD37 MH	29.1	
	(2H7+G28-1)MH	41	
	(2H7+HD37) MH	37.1	
	(G28-1+HD37) MH	35.3	
mAbs			plus GAM
	Ramos	AnnexinV Positive	AnnexinV positive
	cells alone	3	3.3
	2H7 Mab	1.4	3.1
	G28-1 Mab	18.3	8.7
	HD37 Mab	3.7	3.1
	G28-5	3.9	8.3
	2H7+G28-1	32.3	35.7
	2H7+HD37	5	10.5
	2H7+G28-5	5.7	19.4
	HD37+G28-1	26.9	50
	HD37+G28-5	8.2	18.4
	G28-1+G28-5	39.5	68.3

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Caspase 3 Activity in Ramos Cells after 4 Hour Incubation With Cytotoxic B20G SMIP

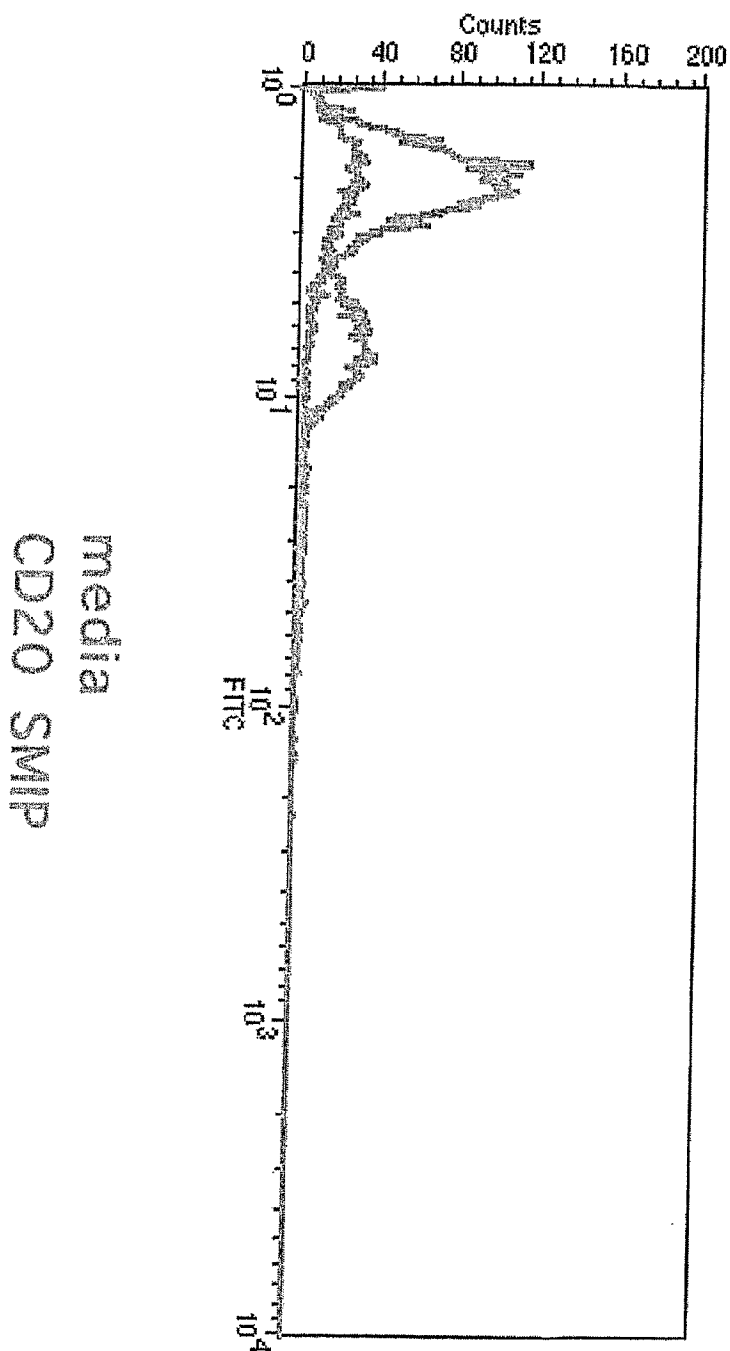


Fig. 72

Complement Dependent Cytotoxicity Mediated by CytoxB20G Derivatives

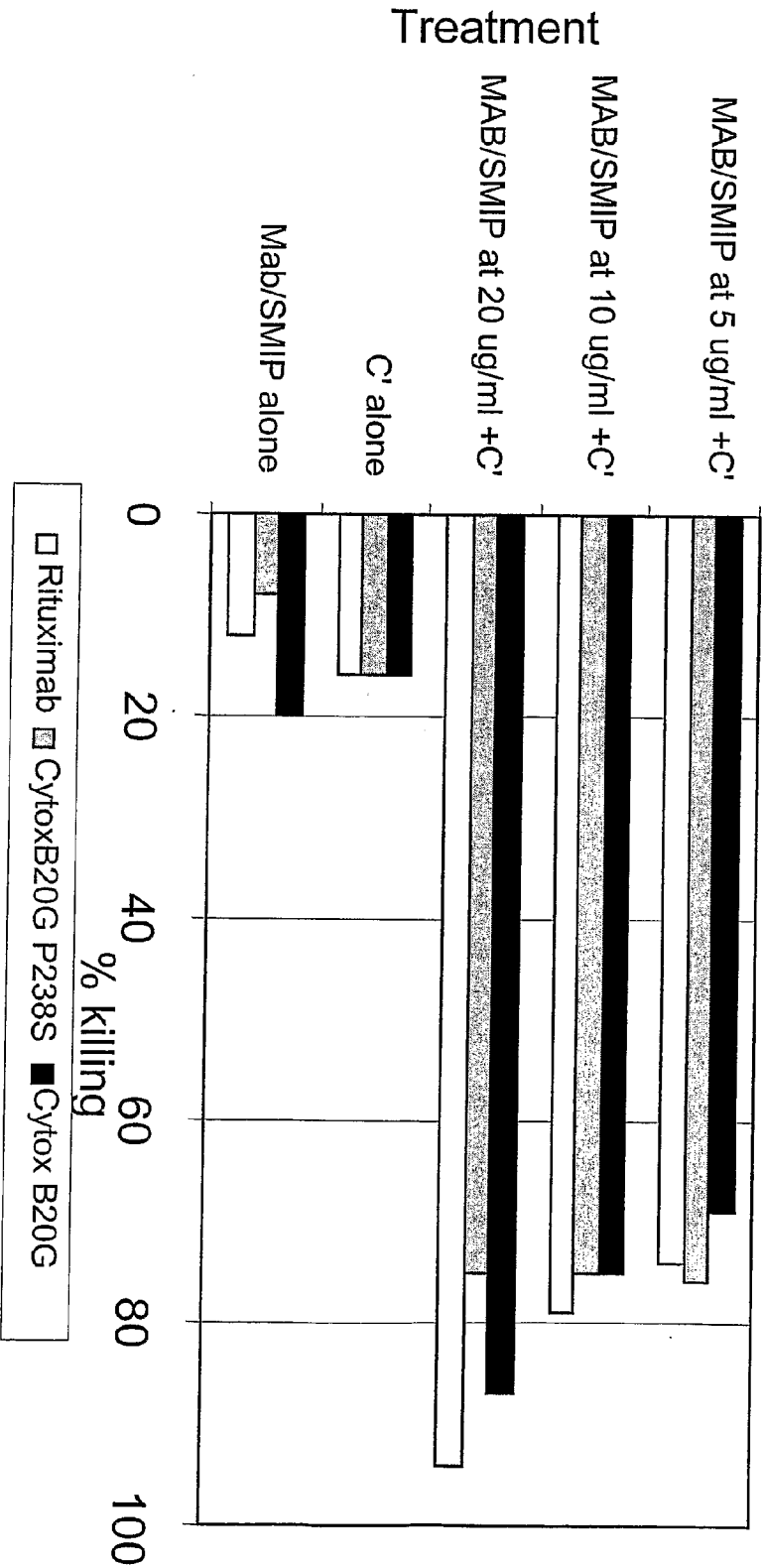


Fig. 73

Figure 76: CDC Activity of CytosB20G SMIPs. CytosB20G, CytosB20GP238, or Rituximab were incubated at increasing concentrations with 10^4 Bjab Target Cells and a 1:10 dilution of rabbit complement (PelFreez) in a volume of 100 microliters for sixty minutes. Aliquots were stained with trypan blue (Invitrogen), and counted using a hemacytometer to determine the percentage of the cell population killed during treatment. Negative controls with cells and only one reagent were also included.

ADCC Activity of CytotoxicB20G SMIPs

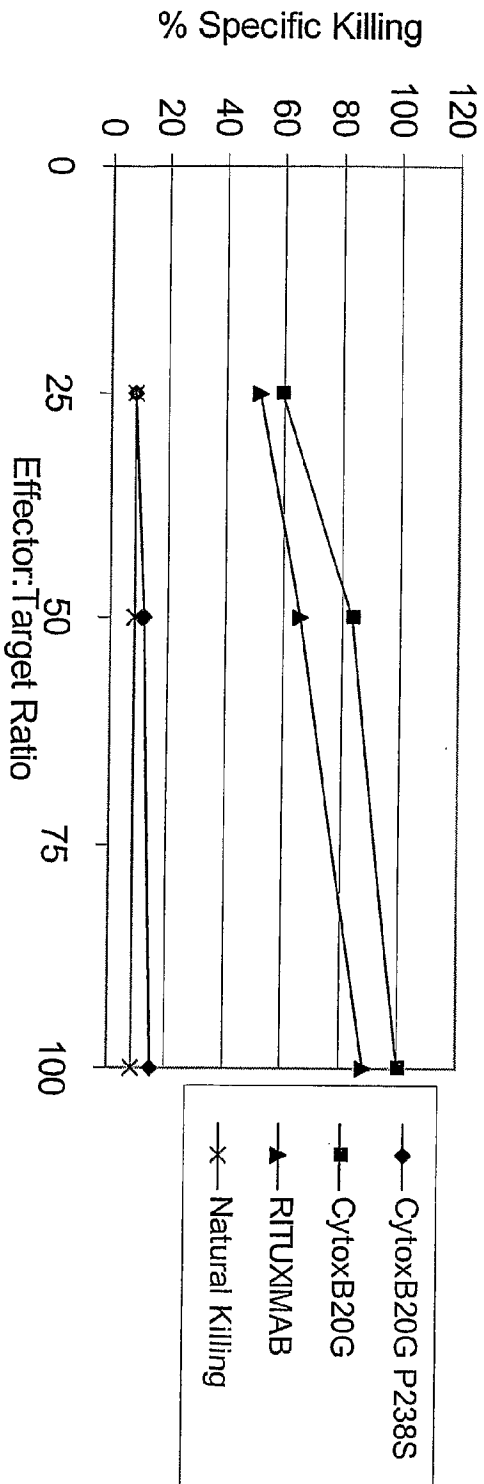


Fig. 74

Figure 77: ADCC Activity of CytotoxicB20G SMIPs. ADCC activity of CytotoxicB20G or Rituximab was measured *in vitro* against BJAB B lymphoma cell line as target and using fresh human PBMC as effector cells. Effector to target ratios were varied as follows: 100:1, 50:1, and 25:1, with the number of BJAB cells per well remaining constant but varying the number of PBMC. BJAB cells were labeled for 2 hours with ^{51}Cr and aliquoted at a cell density of 5×10^4 cells/well to each well of flat-bottom 96 well plates. Purified fusion proteins or rituximab were added at a concentration of 10 $\mu\text{g}/\text{ml}$, and PBMC were added at 1.25 $\times 10^6$ cells/well (25:1), 2.5 $\times 10^6$ cells/well (50:1), or 5 $\times 10^6$ cells/well (100:1), in a final volume of 200 μl . Natural Killing was measured at each effector:target ratio by omission of SMIP or MAb. Spontaneous release was measured without addition of PBMC or fusion protein, and maximal release was measured by the addition of detergent (1% NP-40) to the appropriate wells. Reactions were incubated for 5 hours, and 100 μl culture supernatant harvested to a Lumaplate (Packard Instruments) and allowed to dry overnight prior to counting cpm released on a Packard Top Count NXT Microplate Scintillation Counter.

Binding of soluble FITC-CD16 to CytoxB20G on CD20 CHO Cells

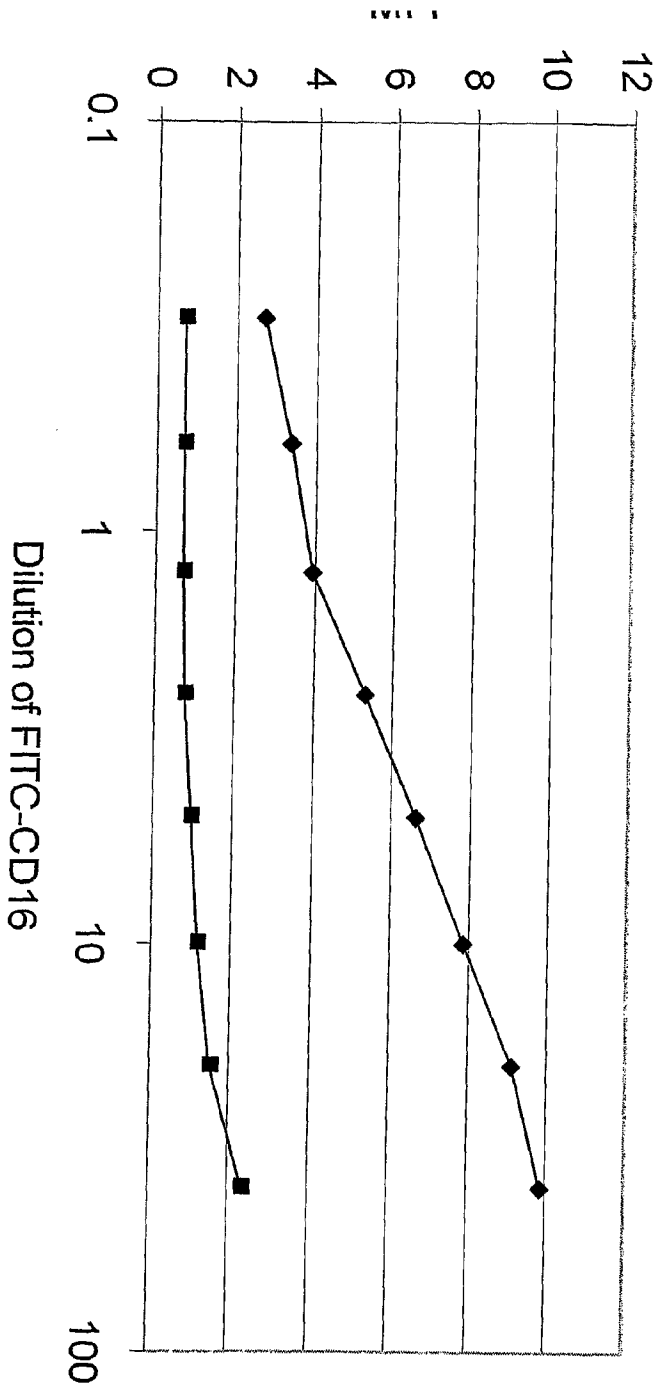


Fig. 75

re 78: Binding of soluble FITC-CD16 to CytoxB20G on CD20 CHO cells. CD20 CHO cells (10^6) were incubated with saturating amounts of CytoxB20G or CytoxB20G P238S (10 μ g/ml) for one hour on ice in PBS/2% FBS. Cells were washed in PBS/2% FBS and incubated with serial dilutions of 0.5 mg/ml FITC-CD16 for one hour on ice. Cells were washed and specific binding measured by flow cytometry using a Beckman-Coulter Epics C machine. Results were analyzed using Expo analysis software and normalized fluorescence units graphed as a function of concentration.

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CytoxB20G and Cytox-B20G P238S SMIPs bind to U937 Cells Expressing FcγRI High Affinity FcR

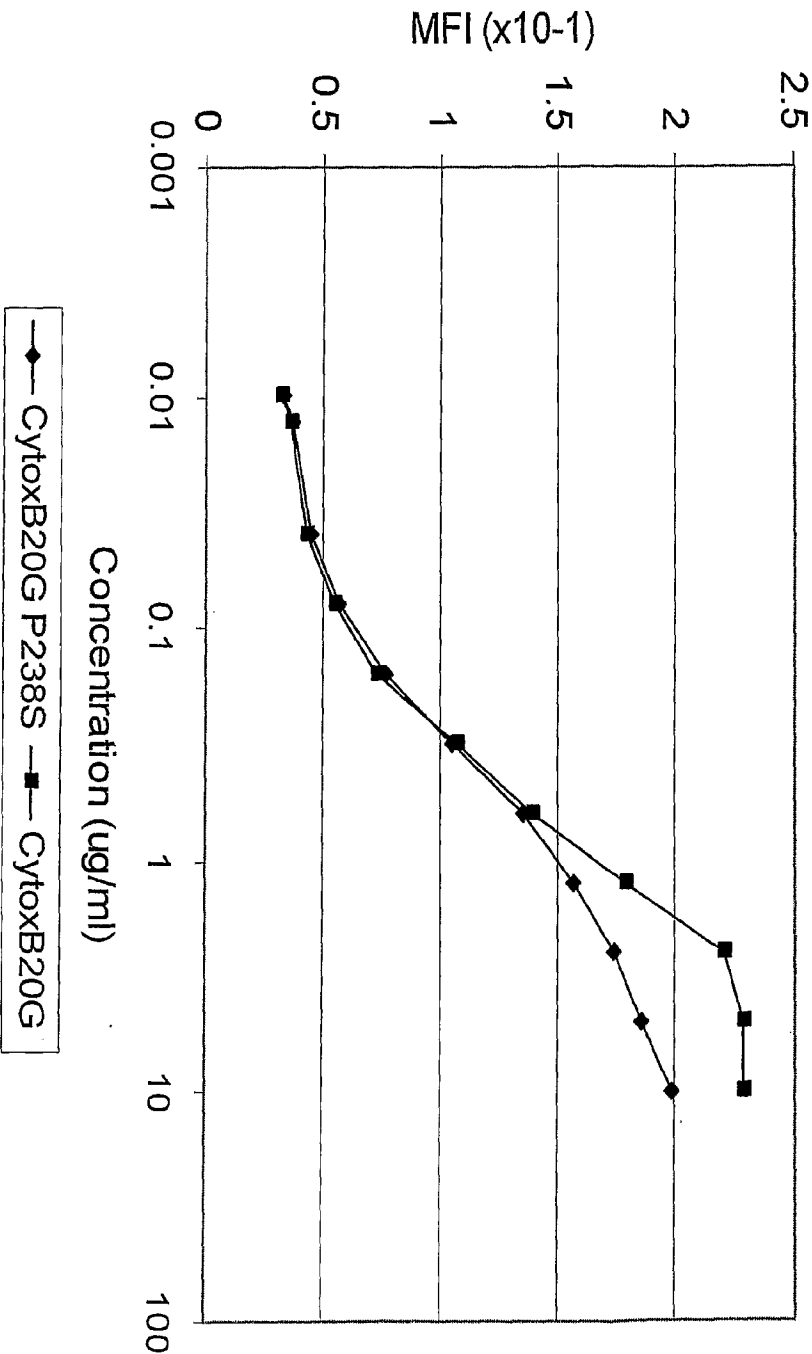


Fig. 76

re 79: Cytox-B20G SMIPs bind similarly to U937 cells expressing the high affinity FcR (FcγRI, CD64). U937 cells expressing CD64 were incubated in PBS/2%FBS for one hour on ice with Cytox-B20G or Cytox-B20G P238S. Cells were washed and incubated for one hour on ice with FITC-goat anti-human IgG1 (Fc specific) (Caltag) at a final dilution 100. Cells were washed and fluorescence analysed on a Beckman-Coulter EpicsC flow cytometer. Data was analyzed using Expo analysis software, and fluorescence intensity graphed as a function of SMIP concentration.

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B Cell Depletion Mediated by Cytox B20G SMIPs

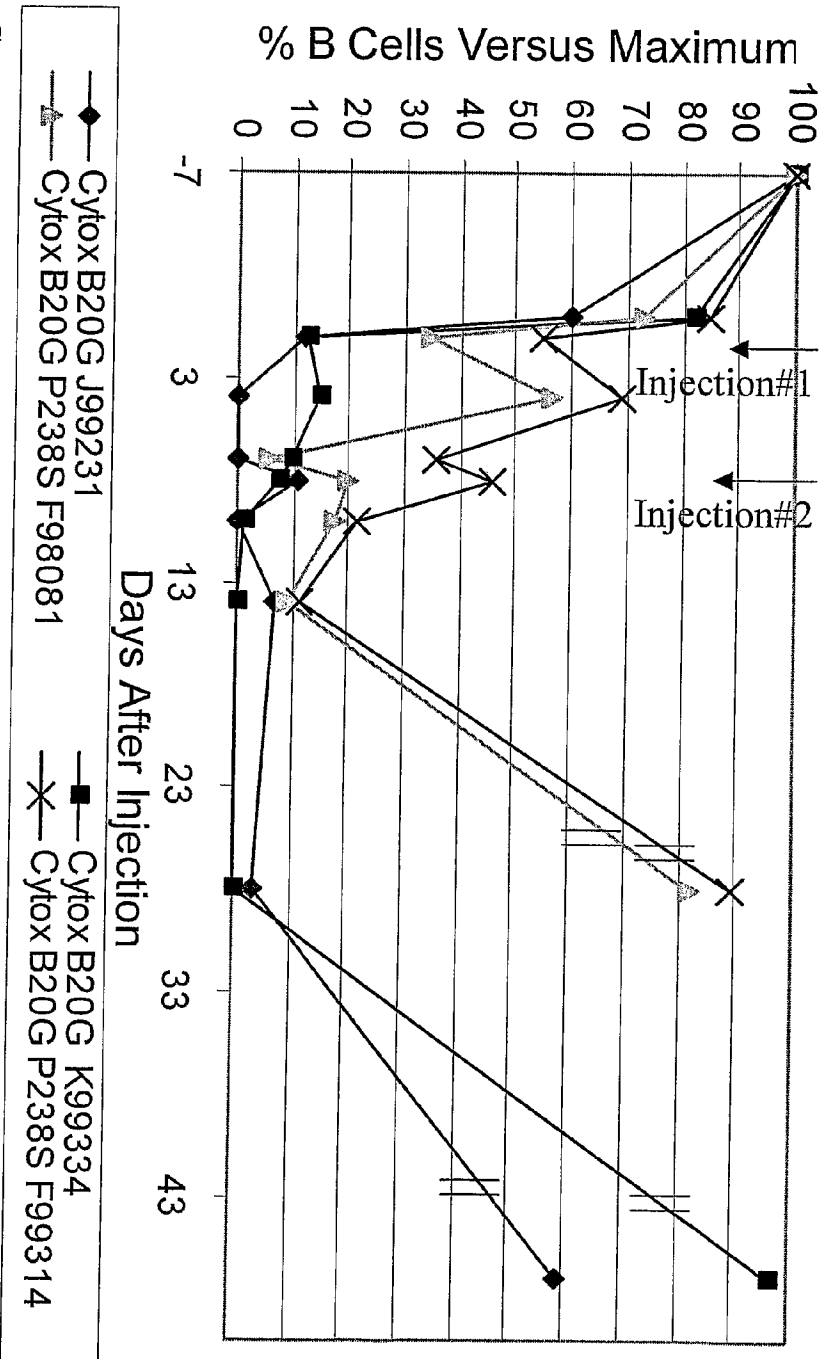


Fig. 77

10: Cytox B20G or Cytox B20G P238S were administered to macaques by intravenous injection at 6 mg/kg, with injections given one week apart. The effect on circulating B cells was measured by detection of CD40 positive B cells in peripheral blood. Blood samples were drawn from injected animals at days -7, 0, 1, 3, 7, 8, 10, 14, 28, and 43. B cell number was estimated by performing CBC (complete blood counts) and two color flow cytometry analysis on monkey FITC or PE conjugates of antibodies against CD40, CD19, CD20, IgG, CD3, CD8 were used in various combinations. Data are plotted as the number of CD40 positive blood B cells tabulated in thousands of cells per ml over time relative to the initial pre-injection time point level of B cells (maximum).

Figure 81: SEC on CytoxB37G SMIPs containing SSS and SSC hinge Domains from Human IgG1

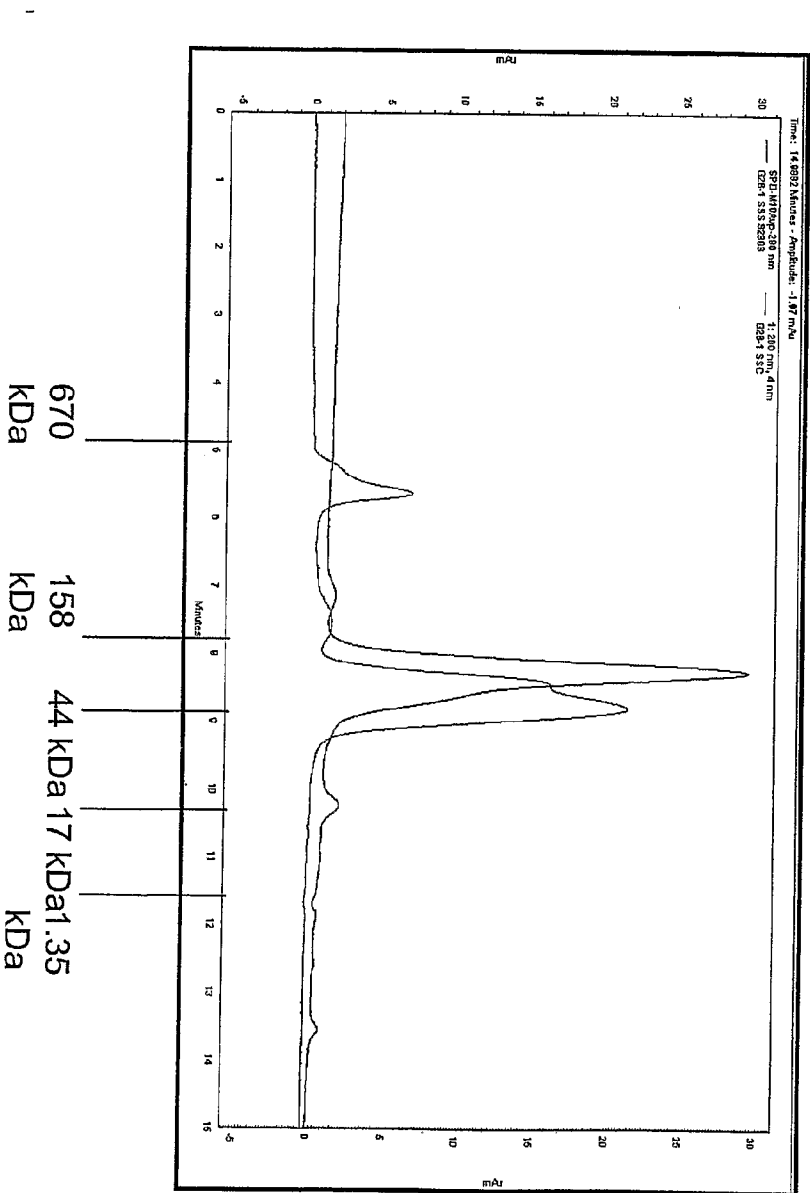


Fig. 78

Figure 81: SEC (Size Exclusion Chromatography) CytoxB37G SMIPs were purified from CHO culture supernatants by Protein A affinity chromatography. Purified aliquots of 10-25 µg were subjected to HPLC over a Tosoh Biosep, Inc. TSK 3000 SWXL HPLC column, pore size 5 µm. The flow rate was 1 ml/min, in PBS, pH 7.2 running buffer. Migration rates of molecular weight standards are indicated below the tracing. The CytoxB37G (SSS)H SMIP indicated in blue, while the CytoxB37G (CSS)H is indicated in red.

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Figure 82: Binding of CytoxB37G SMIPs to B Cell Lymphoma Cell Lines

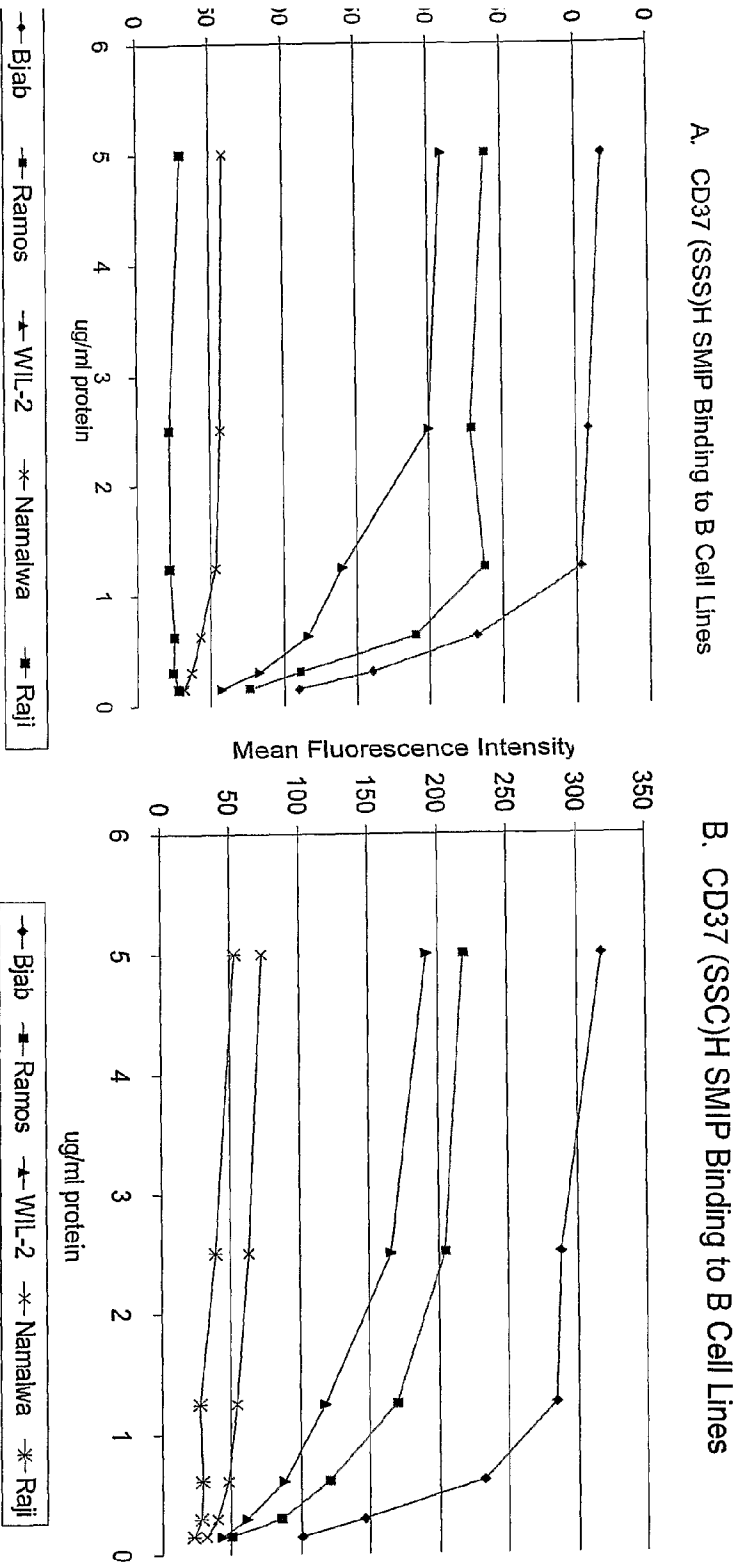


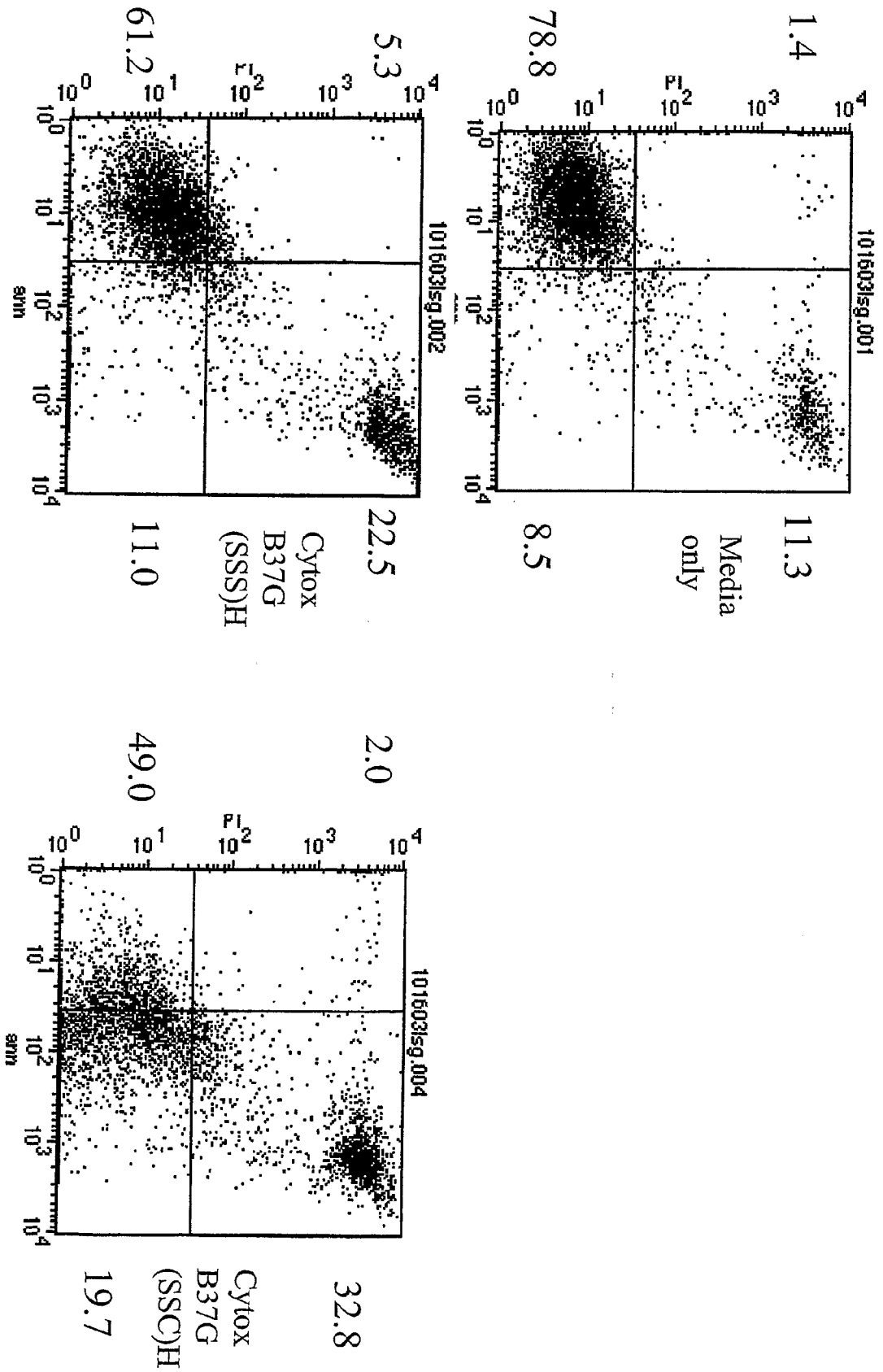
Fig. 79

Figure 82: Binding of CytoxB37G SMIPs to B cell lymphoma cell lines. Serial dilutions of purified CytoxB37G (SSS)H G or CytoxB37G (SSC)H G SMIPs were incubated with 10^6 cells of each cell type for 60 minutes on ice in PBS/2%FBS. Samples were washed twice, and incubated with a mixture of FITC goat anti-human γ and FITC goat anti-human IgG F(ab')₂ (CalTag) at 1:100 each, on ice for 45 minutes. Samples were washed and analyzed by flow cytometry using a FACScalibur (Becton-Dickinson)

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Fig. 80: AnnexinV-PI Staining of Ramos Cells Incubated
24 hours with CD37 SMIPS



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Figure 84: Thymidine Incorporation (Growth Inhibition) in Ramos B-cells after a 48 Hour Incubation with anti-CD37 SMIPs

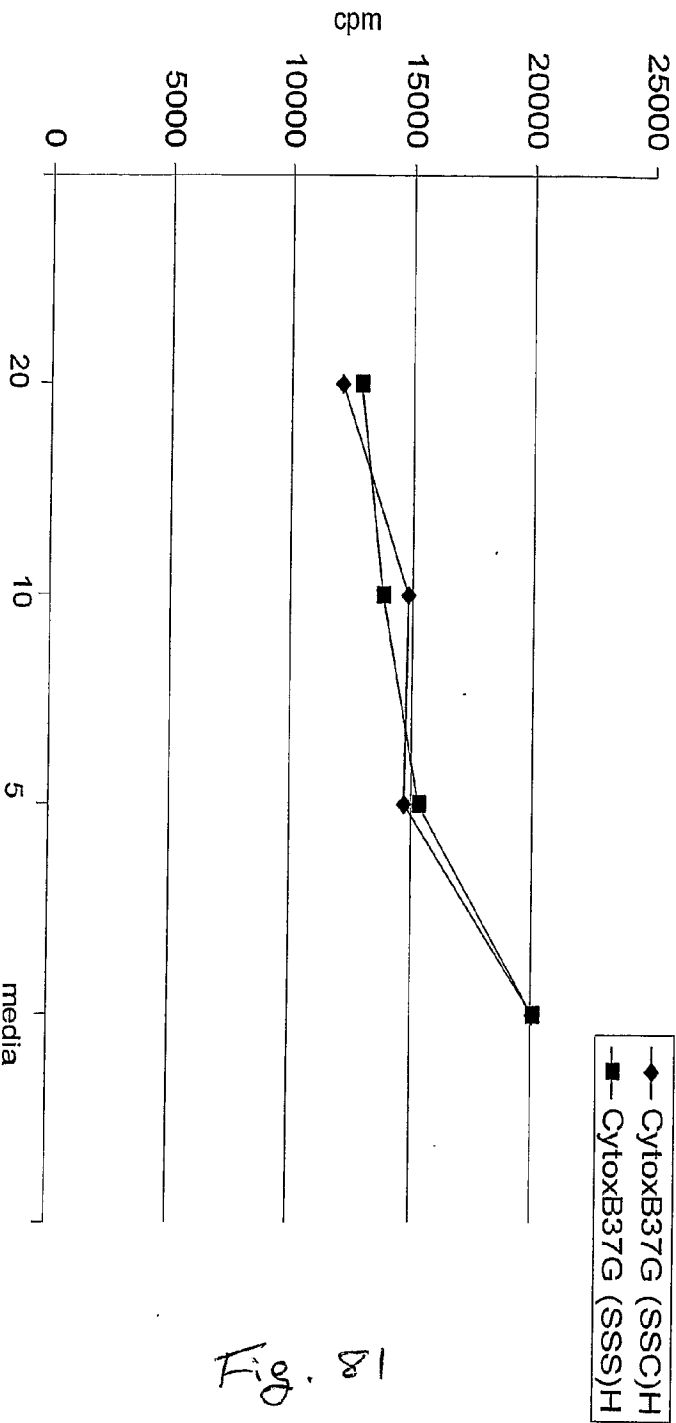


Fig. 84

Figure 84: Ramos B cells were incubated with serial dilutions of purified CD37G SMIPs containing either the IgG1 hinge identified as (SSS)H or (SSC)H. Cultures were incubated in 96 well flat bottom microculture dishes (Costar) at 37°C, 5%CO₂ for 36 hours prior to pulsing with ³H-thymidine for the next 12 hours of a 48 hour incubation (0.75 µCi/well). Cells were harvested onto 96-well GFC plates using a Packard harvester, dried, and 25 µl Microscint scintillation fluid added to each well prior to counting on a TopCount NXT microplate (Packard) scintillation counter. Data are plotted as cpm versus protein concentration. Each SMIP show increasing inhibition of proliferation with increasing protein concentration.

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Figure 85: The Induction of Apoptosis in Ramos B-cells after a 20 hour incubation with different combinations of CD20 and CD37 targeted SMIPs

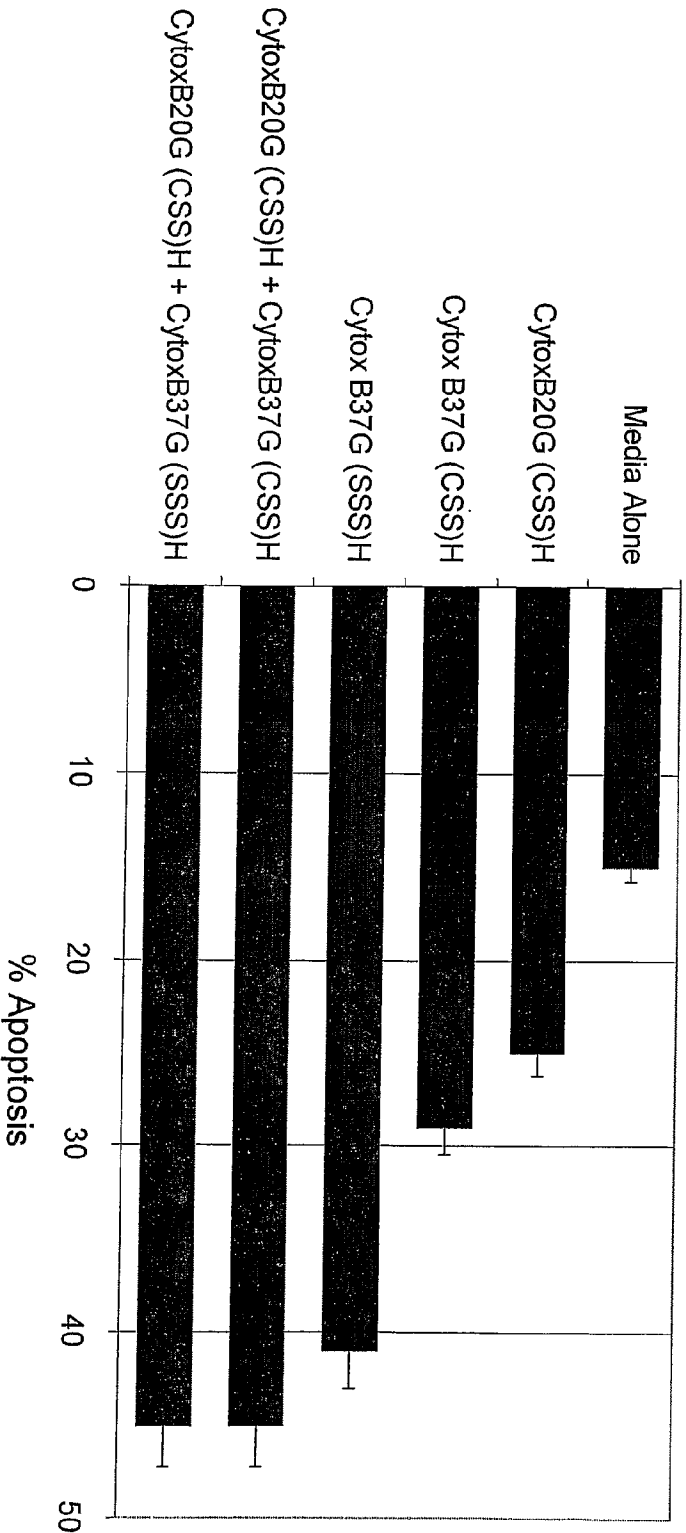


Fig. 82

Figure 85: Ramos B cells were incubated with CD20 and/or CD37 targeted SMIPs (10 µg/ml) in solution for 20 hours. Cells were then harvested, washed, and incubated in annexin V and propidium iodide using a staining kit from Immunotech prior to two color flow cytometry using a FACScalibur flow cytometer (Becton-Dickinson). The graph shows the percentage of annexin V positive cells identified by their staining in the right quadrants of the dot plots.

Figure 86: Complement Mediated Killing of Ramos Cells by CD37 SMIPs

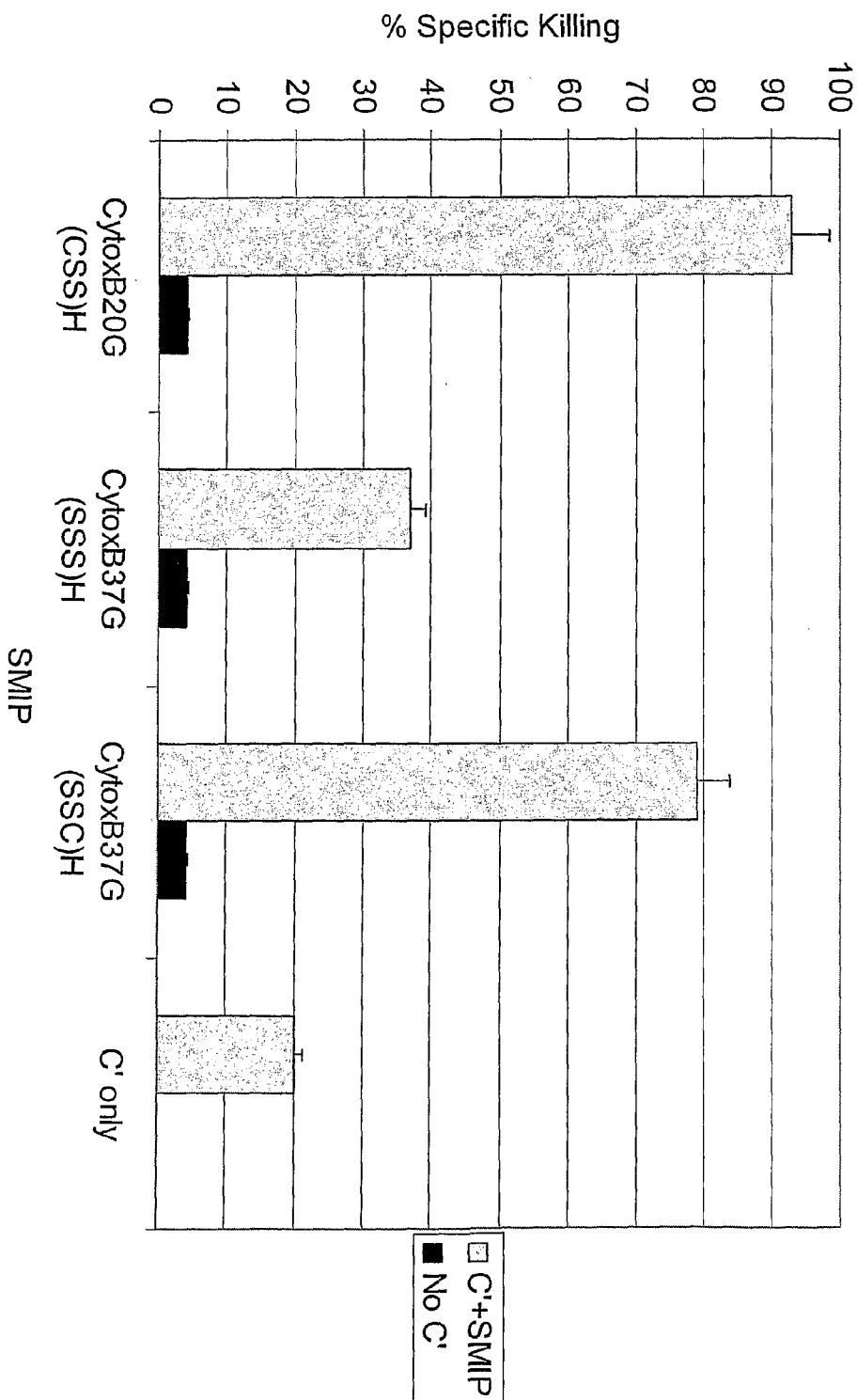


Fig. 83

Figure 86: CDC Activity of CytoxB37G SMIPs. CytoxB20G, CytoxB37 (SSS)H G, CytoxB37 (SCS)H, CytoxB37 (CSS)H, or CytoxB37 (SSC)H were incubated at 10 µg/ml with 10⁴ Ramos Target Cells and a 1:10 dilution of rabbit complement (PelFreez) in a volume of 150 µl for 90 minutes. Aliquots were stained with trypan blue (Invitrogen), and counted using a hemacytometer to determine the percentage of the cell population killed during treatment. Negative controls with cells and only one reagent were also included.

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Figure 87: ADCC Activity of CD37 SMIPs Against Ramos Targets

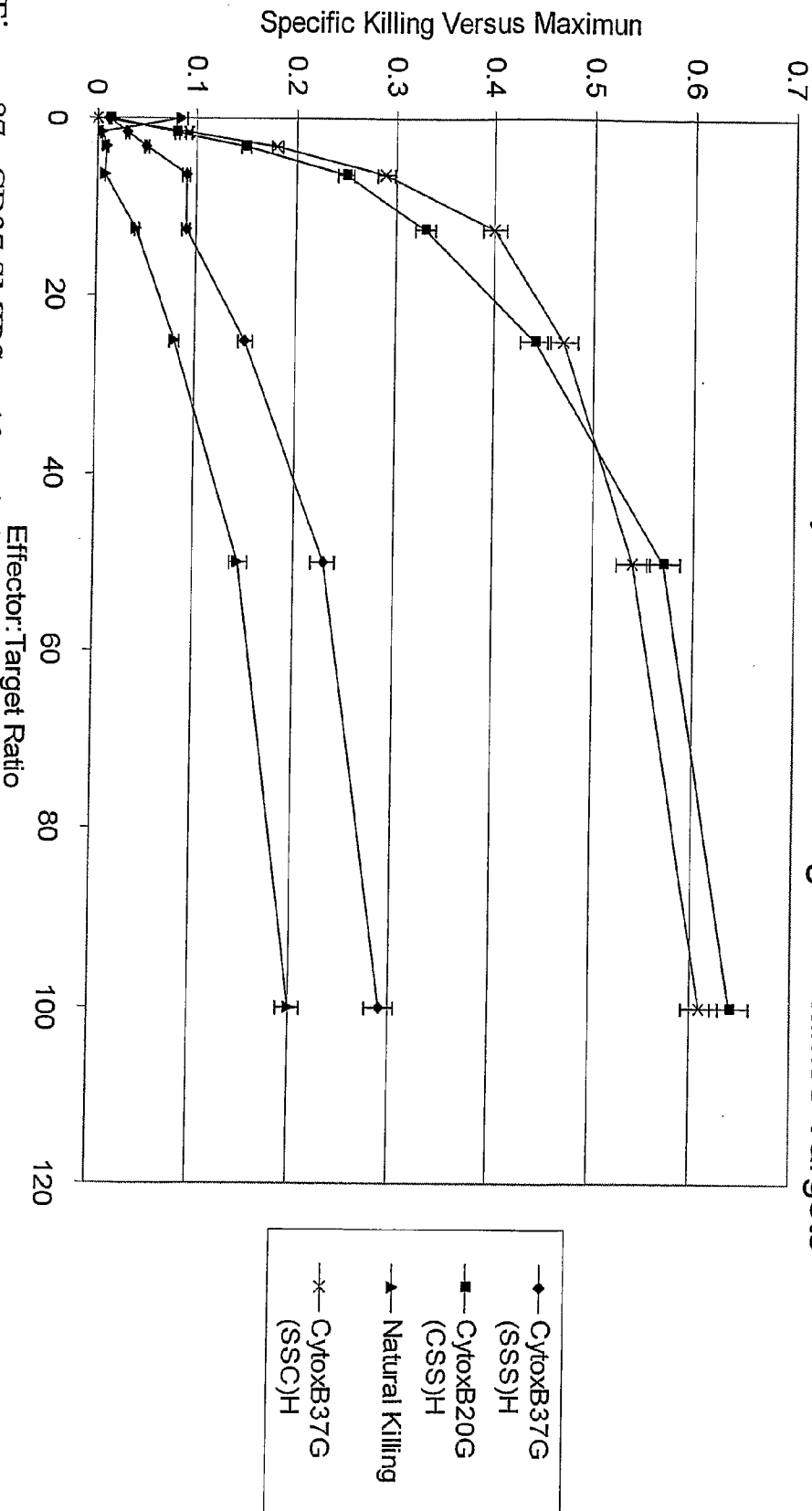


Fig. 84

Figure 87: CD37 SMIPs at 10 $\mu\text{g/ml}$ were incubated in flat-bottom 96 well plates with 10^4 ^{51}Cr -labeled Ramos cells and resting human PBMCs at different effector:target ratios ranging from 0 to 100. All incubations were performed in triplicate at each effector:target ratio. Natural Killing was measured at each effector:target ratio by omission of SMIP. Spontaneous release was measured without addition of PBMC or fusion protein, and maximal release was measured by the addition of detergent (1% NP-40) to the appropriate wells. Reactions were incubated for 6 hours, and 100 μl culture supernatant harvested to a Lumaplate (Packard Instruments) and allowed to dry overnight prior to counting cpm released on a Packard Top Count NXT Microplate Scintillation Counter.

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US03/41600

A. CLASSIFICATION OF SUBJECT MATTER

IPC(7) : C12N 15/00; A61K 39/395; C07K 16/00
US CL : 530/387.3, 388.85, 391.3; 424/130.1; 536/23.4; 435/320.1, 69.6

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
U.S. : 530/387.3, 388.85, 391.3; 424/130.1; 536/23.4; 435/320.1, 69.6

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)
Please See Continuation Sheet

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	HAYDEN et al. Single-chain mono- and bispecific antibody derivatives with novel biological properties and antitumor activity from COS cell transient expression system. Therapeutic Immunology. 1994, Vol. 94, pages 3-15, especially Figure 1, Methods.	1-7, 20-28, 31-40, 53-57, 59, 62-63, 65-75, 116-119, 129-137, 140-150, 161-169, 171-181, 238, 240-243, 251-259, 261-267, 282-285, 287-295, 399-411
Y	US 6,147,203 A (PASTAN et al.) 14 November 2000 (14.11.2000), see entire document, especially abstract, column5-6.	1-7, 20-28, 31-40, 53-57, 59, 62-63, 65-75, 116-119, 129-137, 140-150, 161-169, 171-181, 238, 240-243, 251-259, 261-267, 282-285, 287-295, 399-411

☒ Further documents are listed in the continuation of Box C.

☐ See patent family annex.

* Special categories of cited documents:	"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
"A" document defining the general state of the art which is not considered to be of particular relevance	"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
"E" earlier application or patent published on or after the international filing date	"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	"&" document member of the same patent family
"O" document referring to an oral disclosure, use, exhibition or other means	
"P" document published prior to the international filing date but later than the priority date claimed	

Date of the actual completion of the international search

29 October 2004 (29.10.2004)

Date of mailing of the international search report

02 NOV 2004

Name and mailing address of the ISA/US

Mail Stop PCT, Attn: ISA/US
Commissioner for Patents
P.O. Box 1450
Alexandria, Virginia 22313-1450

Facsimile No. (703) 305-3230

Authorized officer

Larry R. Helms

Telephone No.

571-272-1600

INTERNATIONAL SEARCH REPORT

PCT/US03/41600

C. (Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	US 6,074,644 A (PASTAN et al.) 13 June 2000 (13.06.2000), see entire document, especially column 20.	26, 28, 32, 135, 137, 142, 257, 259, 262
Y	US 5,677,425 A (BODMER et al.) 14 October 1997 (14.10.1997), see entire document, especially abstract, column 3-4.	65-75, 116, 172-180, 288-295
Y	US 6,482,919 B2 (LEDBETTER et al.) 19 November 2002 (19.11.2002), see entire document.	180

INTERNATIONAL SEARCH REPORT

PCT/US03/41600

Continuation of B. FIELDS SEARCHED Item 3:

CAPLUS, MEDLINE, WEST, BIOSIS

Search terms: inventor name, scfv, hinge, cysteine, fusion protein, CD19, CD3, deleted hinge, altered hinge, IgG1, IgA, IgE, disulfide stabilized, constnat region.